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CRUISE REPORTS *DR FRIDTJOF NANSEN*
EAF-Nansen/CR/2018/4



**SURVEY OF REGIONAL RESOURCES AND ECOSYSTEM OFF
SOUTHEAST AFRICA**

Tanzania

6 – 18 April 2018



**Tanzania Fisheries Research Institute
Dar es Salaam, Tanzania**

**Institute of Marine Research
Bergen, Norway**

The EAF-Nansen Programme

The EAF-Nansen Programme "Supporting the application of the Ecosystem Approach to Fisheries Management considering climate and pollution impacts" (GCP/GLO/690/NOR) aims to further strengthen the knowledge base and the overall institutional capacity for the implementation of the Ecosystem Approach to Fisheries (EAF) in developing countries, with additional attention to the impact of climate variability and change, pollution and other anthropogenic stressors.

The programme, that started implementation in May 2017, builds on earlier phases, and is governed by an agreement between the Food and Agriculture Organization of the United Nations (FAO), the Institute of Marine Research (IMR), Norway and the Norwegian Agency for Development Cooperation (Norad). The three pillars of the new programme are: Science, Fisheries management, and Capacity development. A new state of the art research vessel, *Dr Fridtjof Nansen* is an integral part of the programme. A science plan, covering 11 research themes, guides the programme scientific work.

The programme works in partnership with countries, regional organizations, other UN agencies as well as other partner projects and institutions.

Le Programme EAF-Nansen

Le Programme EAF-Nansen "Appuyer la mise en œuvre de l'approche écosystémique de la gestion des pêches en tenant compte des impacts du climat et de la pollution" (GCP/GLO/690/NOR), vise à renforcer la base de connaissances et la capacité institutionnelle pour la mise en œuvre de l'approche écosystémique des pêches (AEP) dans les pays en développement, en accordant une attention particulière aux effets de la variabilité et du changement climatique, de la pollution et d'autres facteurs de stress anthropiques.

Le programme, qui a débuté en mai 2017, s'appuie sur les phases précédentes et est régi par un protocole d'accord entre l'Organisation des Nations Unies pour l'alimentation et l'agriculture (FAO), l'Institut de recherche marine (IMR) de Norvège et l'Agence norvégienne de Coopération au développement (Norad). Les trois piliers du nouveau programme sont : la science, l'aménagement de la pêche et le développement des capacités. Un navire de recherche à la pointe de la technologie, le nouveau *Dr Fridtjof Nansen*, fait partie intégrante du programme. Un plan scientifique, couvrant 11 thèmes de recherche, guide les travaux scientifiques du programme.

Le programme travaille en partenariat avec les pays, les organisations régionales, d'autres agences des Nations Unies ainsi que d'autres projets et institutions partenaires.

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CRUISE REPORTS DR FRIDTJOF NANSEN

**SURVEY OF REGIONAL RESOURCES AND ECOSYSTEM OFF
SOUTHEAST AFRICA**

Tanzania

6 – 18 April 2018

by

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EXECUTIVE SUMMARY

This survey was part of a regional coverage of the eastern coast of Africa (South Africa to Tanzania), aimed at collecting a wide range of data and samples on marine ecosystems from about 20 to 1000 meters depth. Unfortunately, and due to the bottom topography and unavailability of detailed sea charts for Tanzania waters, the survey could not cover areas shallower than about 50 meters depth.

Data collection covered hydrography, meteorology, microplastics, phytoplankton, zooplankton, jellyfish, fishery biology including eggs and larvae, genetics, food safety and fish abundance. Much of the data and samples will be processed in the context of the EAF-Science Plan and related research themes. Hence, this survey report only provides an overview of what has been done during the survey and presents some preliminary results.

The hydrographic conditions were quite similar along the coast of Tanzania showing strong stratification of water masses and low primary productivity. Highest numbers of plastic particles were observed outside Dar es Salaam and northwards between Zanzibar and the main land up to the northern part of Zanzibar. The highest numbers of fish larvae and eggs were observed from Dar es Salaam, through the Zanzibar Channel, and up to Pemba.

Acoustic measurements of pelagic fish showed low abundances, with the shorthead anchovy (*E. heteroloba*) as the most abundant species. It should be noted that the survey did not cover more shallow areas where there is indication of substantially higher densities of Pelagic 1 species than over deeper waters. Furthermore, larger fish, tuna and tuna-like fish in offshore waters cannot be covered through surveys of this type.

Pelagic 2 species were found in a narrow belt in the Southern area in relatively low concentrations, in two smaller areas in near Mafia island and the highest abundances in the Zanzibar Channel. Total biomass was estimated to ~25 000 tonnes.

A total of 335 different species/groups were caught in 30 bottom trawl hauls, mostly fish but also some invertebrates, which reflects the high diversity found in these waters.

CHAPTER 1. INTRODUCTION

1.1 The survey area

The R/V *Dr Fridtjof Nansen* undertook a survey of regional resources and ecosystem in the Indian Ocean off Southeast Africa in 2018. The areas surveyed included the continental shelf and upper slope of coastal East Africa (Leg 1), the Mascarene Bank (Leg 2) and parts of the Bay of Bengal region (Leg 3). Leg 1 covered the continental shelf and upper slope of eastern Africa, with oceanographic transects in the Agulhas Current region and off Tanzania to the Seychelles (Figure 1). This report describes the ecosystem survey along the coast of Tanzania (Leg 1.4).

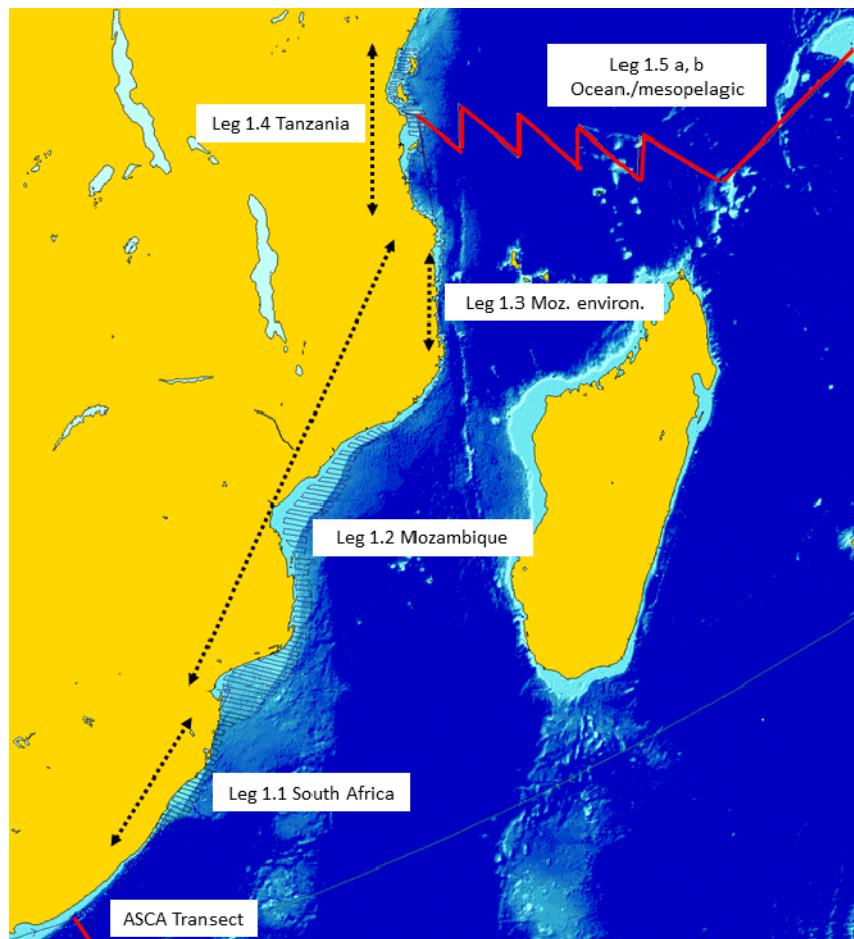


Figure 1. RV *DR Fridtjof Nansen* survey programme 2018, Leg 1.

1.2 Survey objectives

Hydrography:

- Mapping the hydrographic and environmental conditions in the survey area, viz. temperature, salinity, oxygen, chlorophyll, nutrients, pH and total alkalinity.

Phytoplankton, zooplankton, ichthyoplankton and jellyfish:

- Mapping the distribution, abundance and composition of phytoplankton, zooplankton (including jellyfish), and species composition of fish eggs and larvae.

Pelagic and demersal stocks:

- Obtaining information on abundance, size composition and distribution of the main pelagic fish species, and of the pelagic sub-groups *PEL 1 (clupeids, engraulids)* and *PEL2 (carangids, scombrids, barracudas, hairtail)*, using acoustical methods in a systematic grid survey strategy and targeted trawling.
- Obtaining information on abundance, size composition and distribution of the main demersal fish species, crustaceans and squids, using a swept area method with bottom trawls
- Collecting information on maturity stages for the main species of pelagic fish, demersal fish, crustaceans and squids
- Collecting samples for genetic analysis for stock identification of selected species
- Collecting stomach samples for analysis of contents (diet), including microplastics.

Mesopelagic fish:

- Identifying the main species and collect samples for identification and isotope analysis.

Marine debris and pollution:

- Recording occurrence of marine debris (surface);
- Collecting samples for levels of nutrients and contaminants including microplastics;
- Mapping occurrence of microplastics and describe associated neuston communities.

Contaminants and parasites:

- Collecting samples of fish species consumed locally, and other indicator species such as soles, for analysis of contaminant levels and nutrient values, and identifying and quantifying selected parasites in commercially important fish.

1.3 Participation

Pwani University, Kenya:

Jackline Adhiambo.

State Department of Fisheries and Blue Economy, Kenya:

John Kiarie Njuguna.

Kenya Marine and Fisheries Research Institute, Kenya:

Charles Mitto Kosore.

Instituto Nacional de Investigacao Pesqueira, Mozambique:

Celso Billy Isac Montanha.

Institute of Marine Research, Norway (IMR), Norway:

Vidar Fauskanger, Lucilla Giulietti, Tore Johannessen (Cruise leader), Helene Lødemel, Lars Johan Naustvoll, Hege Rognaldsen, Silje Elisabeth Seim, Olaf Johan Sørås, Jan Frode Wilhelmsen, Diana Zaera.

Ministry of Fisheries and Marine Resources, Somalia:

Abdi Ali Yare.

The University of the Western Cape, South Africa:

Yasmeen Parker.

Tanzania Fisheries Research Institute, Tanzania:

Charles Nyarongo Ezekiel, Valeli Joseph, Benedicto Boniphace Kashindye, Mary Alphonse Kishe-Machumu, Shigalla Mahongo (Co-cruise leader), Patroba Patrick Matiku, Hakim Davis Matola, Catherine Adam Mwakosya, Salome Daniel Shayo.

University of Dar es Salaam, Tanzania:

Margareth Kyewalyanga, Pooja Solanki, Omar Juma Suleiman.

State University of Zanzibar, Tanzania

Mohammed Suleiman Mohammed.

Tanzania Navy, Tanzania

Mussa Ally Yege.

1.4 Narrative

R/V *Dr Fridtjof Nansen* departed from Dar es Salaam on 6 April 2018 at 14 00 UCT. The first transect, just north of the border between Mozambique and Tanzania, was reached on 7 April 2018 at 1500 UCT. From the southern border, the Tanzanian coast was surveyed northwards, but only to 5°S due to maritime security regulations. *Dr Fridtjof Nansen* anchored in Menai Bay on the south-west side of Unguja Island in Zanzibar from the evening of 13 April 2018 to noon the following day, for calibration of echo-sounders. Due fish schools interfering with the calibration, and later on strong tidal currents bringing the calibration spheres out of position, the attempt was aborted as it was unsuccessful. *Dr Fridtjof Nansen* returned to Dar es Salaam on 18 April 2018 at 0600 UCT (9 am local time).

The wind conditions were generally good during the survey and did not limit sampling. Except for a few instances where strong currents prevented operating nets (plankton and fish

eggs and larvae) close to the shore, all hydrographical stations were carried out as planned. The shelf along the coast of Tanzania is generally very narrow (Figure 2), sloping steeply from shallow waters (20-30 m) to 300-400 m depth (Figure 2). The slope is characterized by uneven topography which does not allow bottom trawling in most places. Hence, the bottom trawling programme of four hauls per transect had to be substantially reduced. The topography in shallower waters along the coast of Tanzania is also generally “hilly”. Furthermore, the maps describing depths and bottom topography are relatively coarse with many shallow bottom structures missing. Therefore, except for the Zanzibar Channel and the delta area north of Mafia where the bottom is relatively smooth, surveying bottom depth <50 m was considered unsafe and was generally avoided. Due to reduced bottom trawl sampling, it proved impossible to carry out the planned programme to estimate abundance of fish stocks. Instead, more focus was put on describing species composition and diversity of demersal fishes and invertebrates.

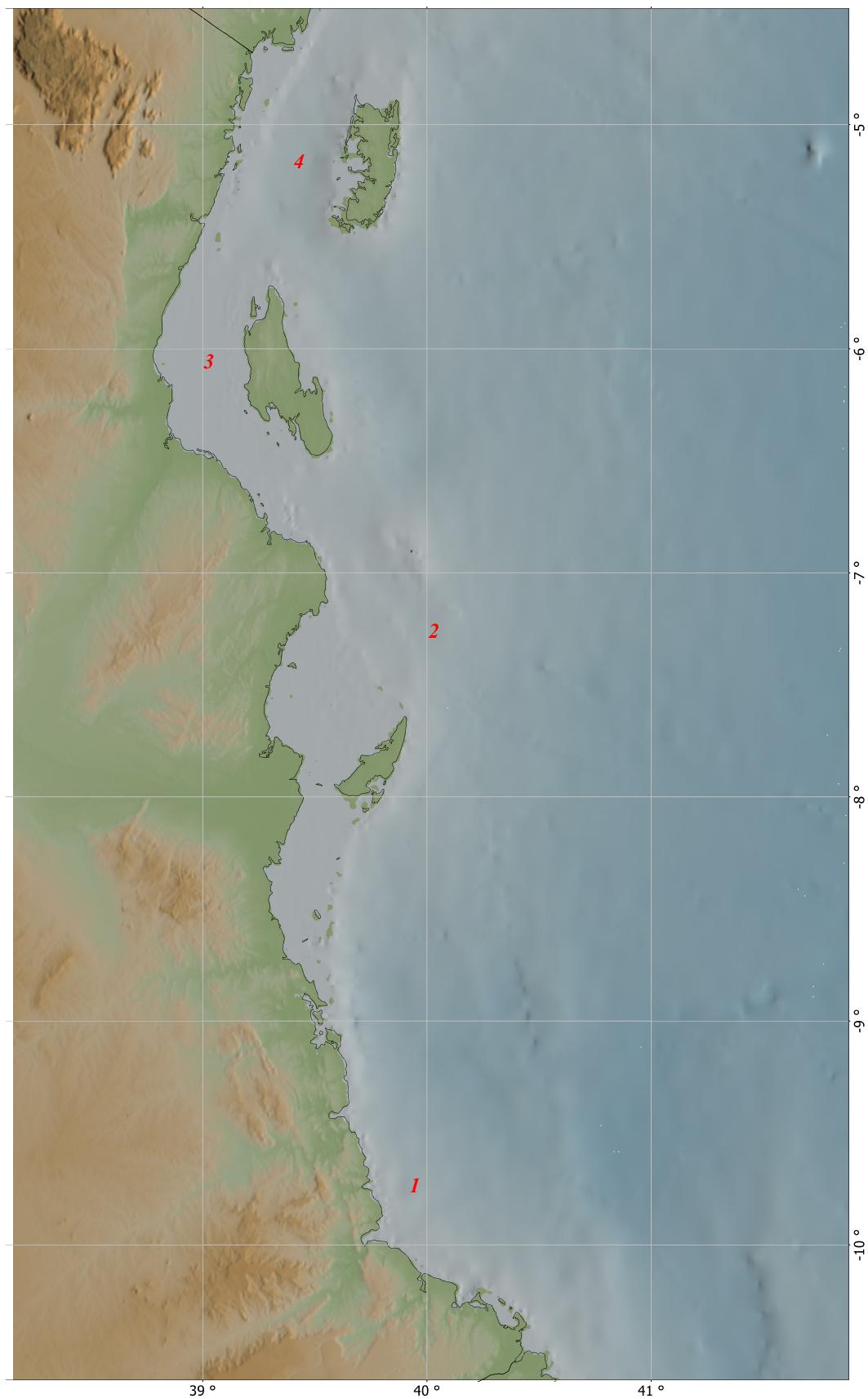


Figure 2. Depth contours along the coast of Tanzania: 0-20 m is indicated in blue along the shore, and survey depths (20-1000 m) are indicated in light blue, where lines indicate 50, 100, 200, 500 and 1000 m isobaths. 1-4 (in red) indicate sub-areas used during surveys with the first *Dr Fridtjof Nansen* in 1982-83: 1 – the southern area (south of 9°S), 2 – Mafia, 3 – Zanzibar, 4 – Pemba.

1.5 Survey effort

The survey was run along pseudo-parallel acoustic transects, perpendicular to the coastline, approximately 15 nautical miles apart, and from about 50 m to 1000 m depth (Figure 3a). In the Zanzibar Channel (between Zanzibar and the mainland), which is an important fishing area, the distance between two transects was reduced to 10 nautical miles. Both sides of the islands of Zanzibar (Unguja and Pemba), but not north of 5°S for security reasons (piracy).

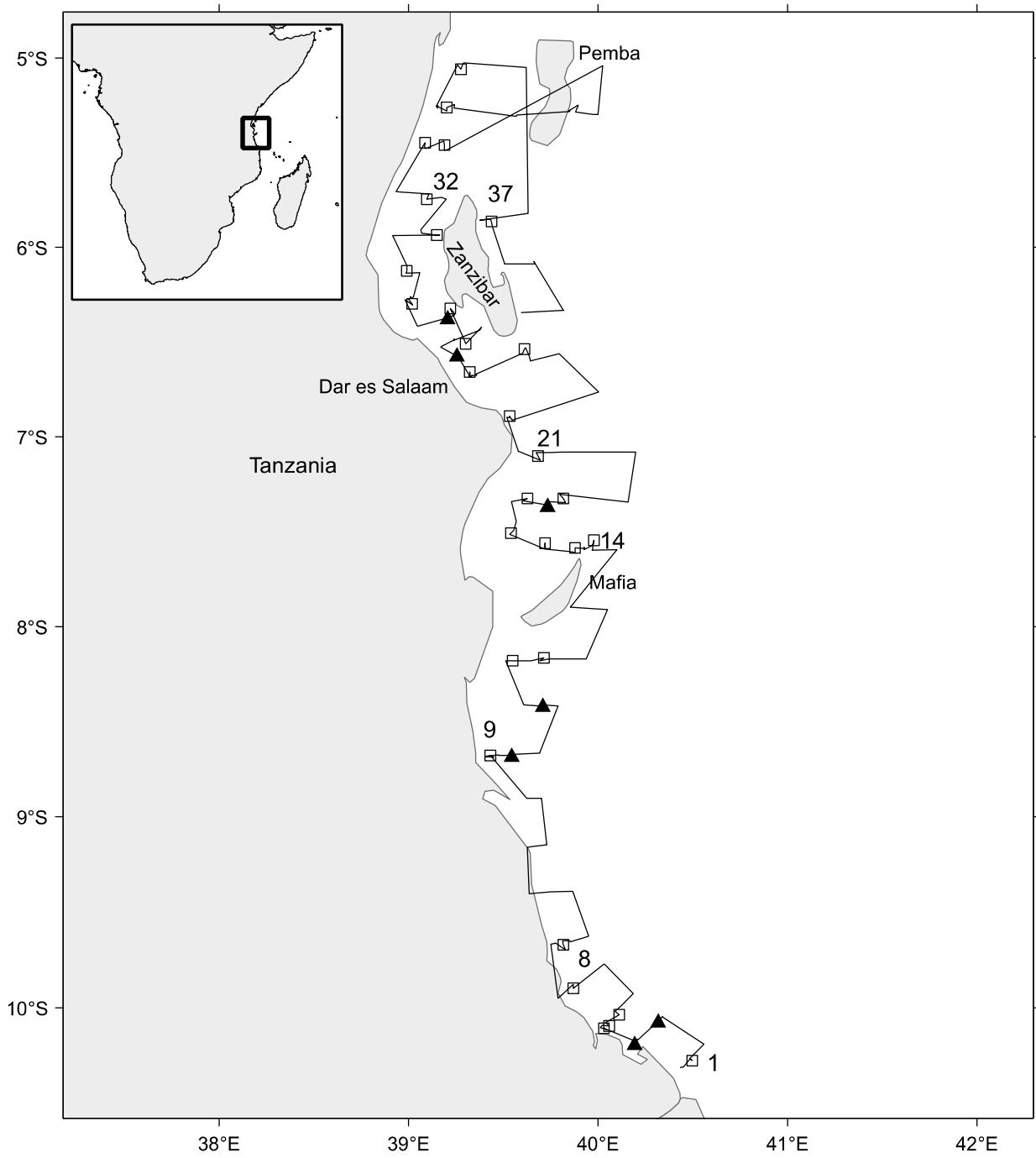
Pelagic trawling was done irregularly, either to identify echo registrations or to check ‘blindly’ if fish were mixed with plankton in the upper layers of the water column. Pelagic trawl with floats was occasionally used to catch fish close to the surface. Bottom trawling was carried out where the bottom was sufficiently smooth, generally deeper than 200 m, or <50 m in the Rufiji delta area north of Mafia and the Zanzibar Channel (Table 2). Figure 3a shows the course track and trawl stations.

In addition to continuous acoustic recording for pelagic and mesopelagic for fish biomass estimates, continuous recordings of data from the multibeam bottom mapping echo sounder EM710, thermosalinograph and weather station was carried out.

At every second transect, hydrographic observations were carried out to a depth of 1000 m. These transects included CTD casts and sampling of phytoplankton, zooplankton, fish eggs and larvae, and microplastics. Figure 2b shows the position of hydrographical stations, and Table 1 effort during the survey in terms of number of trawl stations, CTD casts and samples of microplastics (Manta net), phytoplankton, zooplankton (WP2) and fish eggs and larvae (Multi net).

Fish were also sampled for analyses of food safety and parasites. The number of samples for food safety are given in Table 3, and number of samples and preliminary results from the parasite studies are represented in Table 4.

All collected data are the property of the United Republic of Tanzania.



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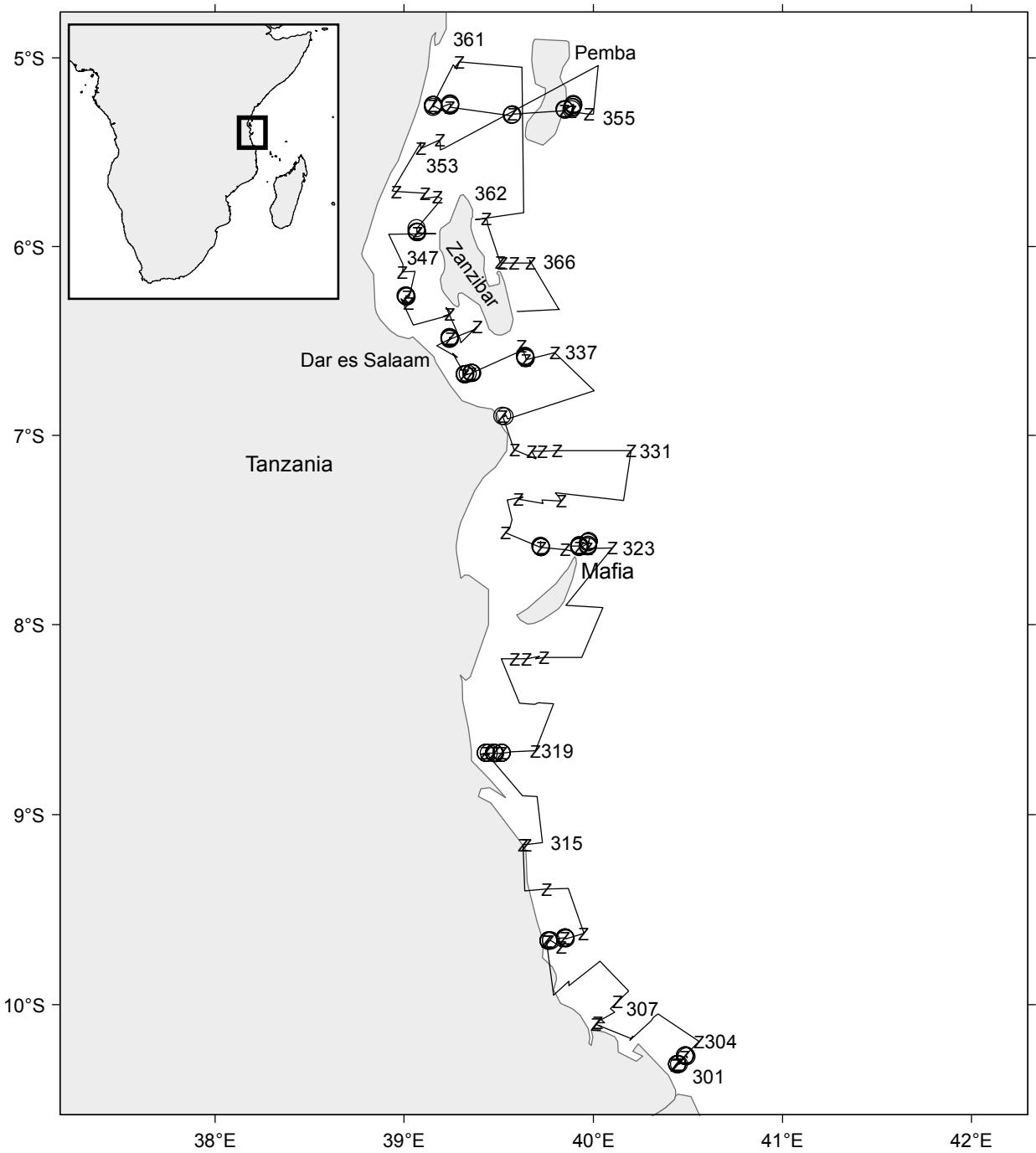
6–18 April 2018

Trawl st. no 1-37

□ Bottom trawl

▲ Pelagic trawl

Figure 3a. Course tracks with trawl stations along Tanzania.



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6–18 April 2018

Z CTD st.no 301-366
○ Plankton st. (several gears)

Figure 3b. Course track with hydrographic and plankton stations: Z indicates CTD and O CTD, plankton, Multi net for eggs and larvae, Manta net for microplastics and chemical seawater analyses.

Table 1. Survey effort: Phyto -phytoplankton net, WP-2 – zooplankton net, Multi – net for eggs and larvae, Manta – net for microplastic particles in the surface, BT-bottom trawl, PT- pelagic trawl.

	NM sailed	CTD	Phyto	WP-2	Multi	Manta	BT	PT
The southern area	224	15	4	8	4	4	6	2
Mafia	545	25	9	18	9	9	13	3
Zanzibar	304	15	3	4	3	3	7	2
Pemba	289	15	3	6	5	4	4	0
Total	1 362	65	19	36	21	19	30	7

Table 2. Bottom trawl hauls per depth stratum in the various areas.

Area	20-50 m	50-100 m	100-200 m	200-1000 m	Sum
The Southern area	0	1	0	5	6
Mafia	5	2	0	6	13
Zanzibar	4	2	0	1	7
Pemba	0	1	0	3	4
Total	9	6	0	15	30

Table 3. Number of fish samples collected for food safety analyses per species.

LARGE FISH

DATE	SPECIES	NO. OF FISH	TISSUE	FREECE-DRIED	STATION
08/04/2018	<i>Trichiurus lepturus</i>	5	Fillet and liver	6	5
09/04/2018	<i>Trichiurus lepturus</i>	5	Fillet and liver	6	7
10/04/2018	<i>Saurida undosquamis</i>	25	Fillet and liver	30	13
12/04/2018	<i>Trichiurus lepturus</i>	25	Fillet and liver	30	21
12/04/2018	<i>Saurida undosquamis</i>	15	Fillet and liver	18	21
15/04/2018	<i>Scomberomorus commerson</i>	8	Fillet and liver	7	33
	Total	83		97	

SMALL FISH

DATE	SPECIES	NO. OF FISH	TISSUE	FREECE-DRIED	STATION
08/04/2018	<i>Decapterus kuroides</i>	150	Whole fish	6	3
09/04/2018	<i>Stolephorus heterolobus</i>	300	Whole fish	3	9
10/04/2018	<i>Spratelloides gracilis</i>	450	Whole fish	3	12
11/04/2018	<i>Upeneus taeniopterus</i>	150	Whole fish	6	16
13/04/2018	<i>Encrasicholina punctifer</i>	900	Whole fish	3	25
14/04/2018	<i>Spratelloides gracilis</i>	450	Whole fish	3	28
14/04/2018	<i>Stolephorus heterolobus</i>	300	Whole fish	3	28
15/04/2018	<i>Carangoides malabaricus</i>	150	Whole fish	6	31
15/04/2018	<i>Upeneus taeniopterus</i>	150	Whole fish	6	31
15/04/2018	<i>Decapterus macrosoma</i>	150	Whole fish	6	32
	Total	3 150		45	

Table 4. Number of fish investigated for parasites.

SPECIES	DATE	NO. OF FISH	TISSUE EXAMINED	PARASITE INVESTIGATED	INFECTED (Y= yes N= no)	FREEZED SAMPLES (Y= yes N= no)	TRAWL STATION
<i>Scomberomorus commerson</i>	09/04/2018	12	Fillet	<i>Kudoa</i> spp.	N	N	9
<i>Scomberomorus commerson</i>	15/04/2018	8	Fillet	<i>Kudoa</i> spp.	N	N	33
<i>Lophoides mutilus</i>	08/04/2018	5	Fillet, Viscera	Anisakids	Y	Y	6
<i>Trichiurus lepturus</i>	08/04/2018	9	Fillet, Viscera	Anisakids	y	y	6
<i>Trichiurus lepturus</i>	08/04/2018	19	Fillet	<i>Kudoa</i> spp.	y	y	7
<i>Trichiurus lepturus</i>	12/04/2018	50	Fillet	<i>Kudoa</i> spp.	y	y	21
<i>Trichiurus lepturus</i>	12/04/2018	20	Fillet, Viscera	Anisakids	y	y	22
<i>Polysteganus coeruleopunctatus</i>	08/04/2018	5	Fillet, Viscera	Anisakids	y	y	5
<i>Polysteganus coeruleopunctatus</i>	10/04/2018	6	Fillet, Viscera	Anisakids	y	y	13
<i>Polysteganus coeruleopunctatus</i>	12/04/2018	5	Fillet, Viscera	Anisakids	y	y	22
<i>Decapterus macrosoma</i>	08/04/2018	2	Fillet, Viscera	Anisakids	y	y	5
<i>Saurida undosquamis</i>	08/04/2018	5	Fillet, Viscera	Anisakids	y	y	6
<i>Saurida undosquamis</i>	09/04/2018	6	Fillet, Viscera	Anisakids	y	y	8
<i>Saurida undosquamis</i>	10/04/2018	11	Fillet, Viscera	Anisakids	y	y	13
<i>Saurida undosquamis</i>	10/04/2018	9	Fillet, Viscera	Anisakids	y	y	14
<i>Saurida undosquamis</i>	11/04/2018	17	Fillet, Viscera	Anisakids	y	y	17
<i>Saurida undosquamis</i>	14/04/2018	16	Fillet, Viscera	Anisakids	y	y	27
<i>Saurida undosquamis</i>	15/04/2018	7	Fillet, Viscera	Anisakids	y	y	30
<i>Ephinephelus areolatus</i>	11/04/2018	4	Fillet, Viscera	DIGENEAN	y	y	17
<i>Octopus sp.</i>	12/04/2018	2	Fillet, Viscera	Anisakids	N	N	21

CHAPTER 2. METHODS

2.1 Meteorological data

Meteorological data was logged continuously from the AANDERAA Smartguard meteorological station and included wind direction and speed, air pressure, relative humidity, air temperature and shortwave solar radiation. All data were logged to the Nansis tracklog system averaged every 60 seconds. A problem with the sensor systems may have had impact on the quality of the data. Hence, these data will not be presented until the sensors have been calibrated.

2.2 Oceanography

2.2.1 Thermosalinograph

The SBE 21 thermosalinograph ran continuously during the survey, obtaining samples of sea surface (at 4 m depth) salinity and relative temperature every 10 seconds. An attached in-line C3 Turner Design Submersible Fluorometer measured turbidity and chlorophyll-*a* levels.

2.2.2 Current speed and direction measurements (ADCP)

Two hull-mounted Acoustic Doppler Current Profiler (VMADCP) from RD Instruments ran during the survey. The frequency of the VMADCP are 75 and 150 kHz. The system was run in narrow band mode and data was averaged in 16 and 4 m vertical bins at 75 and 150 kHz respectively and stored on files for post survey processing. The 150 kHz was run continuously while the 75 kHz was turned off during the last part of the survey due to interference with the ping rate of the EK80 echosounder.

2.2.3 Temperature, salinity, oxygen, fluorescence and sampling of water

Vertical profiles of temperature and salinity were obtained by a Seabird 911 CTD, while *in situ* concentrations of dissolved oxygen were measured using a CTD-mounted SBE 43 oxygen sensor. Real time logging and plotting was performed using the Seabird Seasave software installed on a PC. Attached to the CTD was also an uncalibrated Chelsea Mk III Aquatracka fluorometer, which measures *in situ* fluorescence on relative scale (measure of chlorophyll-*a*). 12 Niskin water-bottles (10 l), attached to a CTD-mounted rosette, were used to collect water at standard sampling depths: 5, 25, 50, 75, 100, 150, 200, 250, 300, 400, and 500 m.

2.2.4 Hydrographical transects

The sailing order suggested CTD casts at bottom depths of 30 m, 75 m, 100 m, 200 m and 500 m, at transects 30 nautical miles apart. However, as the bottom generally sloped steeply from shallow water to 200-300 m, the distance between station <200 m was often negligible. Hence, at most transects the 75 and 200 m stations were cancelled, and in some instances also the 30 m station was not sampled. Additional CTD casts were carried out at 1000 m depth, and in relation to bottom trawl sampling.

At every second hydrographical transect (60 NM apart), a more extensive sampling programme was carried out at depths of 30 m (sometimes cancelled), 100 m and 500 m. The samples collected at these so-called “super-stations” are shown in Figure 4. The most northerly of these transects ($\sim 5^{\circ}\text{S}$) was carried out on both sides of the Pemba Channel and on the eastern side of Pemba as well (Figure 3). Three additional super-stations (30 m stations) were carried out along the middle of the Zanzibar Channel.

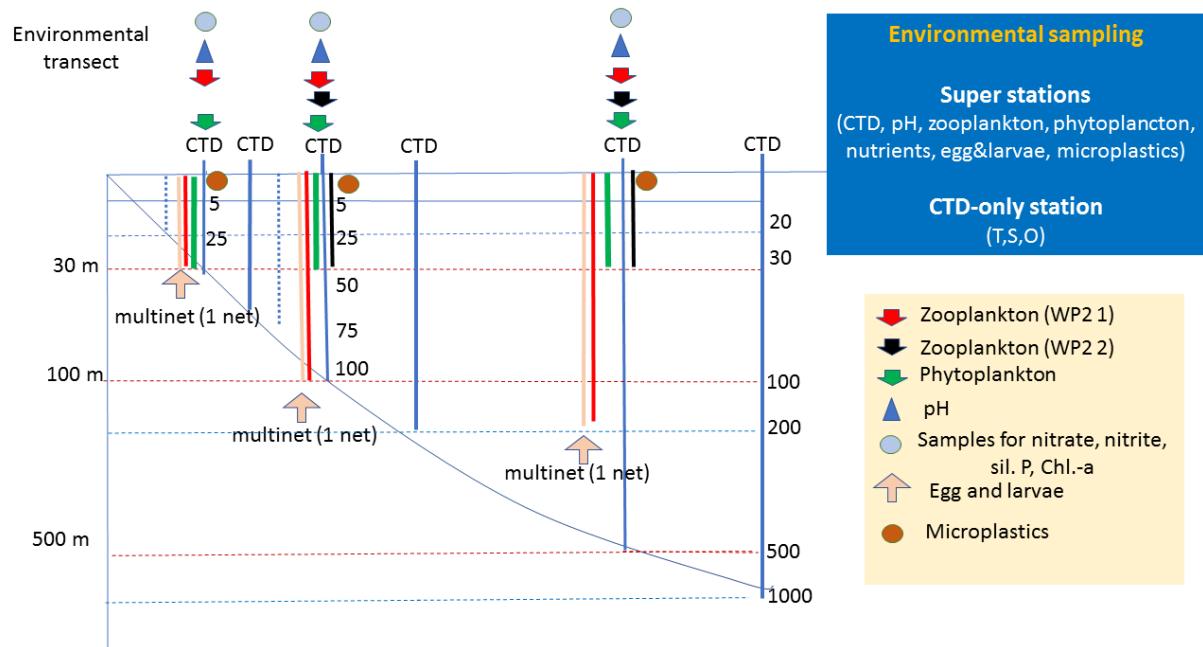


Figure 4. Sampling diagram showing the depth and the sampling at the hydrographical transect with the super-stations transects, from the inshore (left) towards the deep 500 m station (right).

The oxygen sensor on the CTD was validated using the Winkler titration (Grasshoff *et al.* 1983).

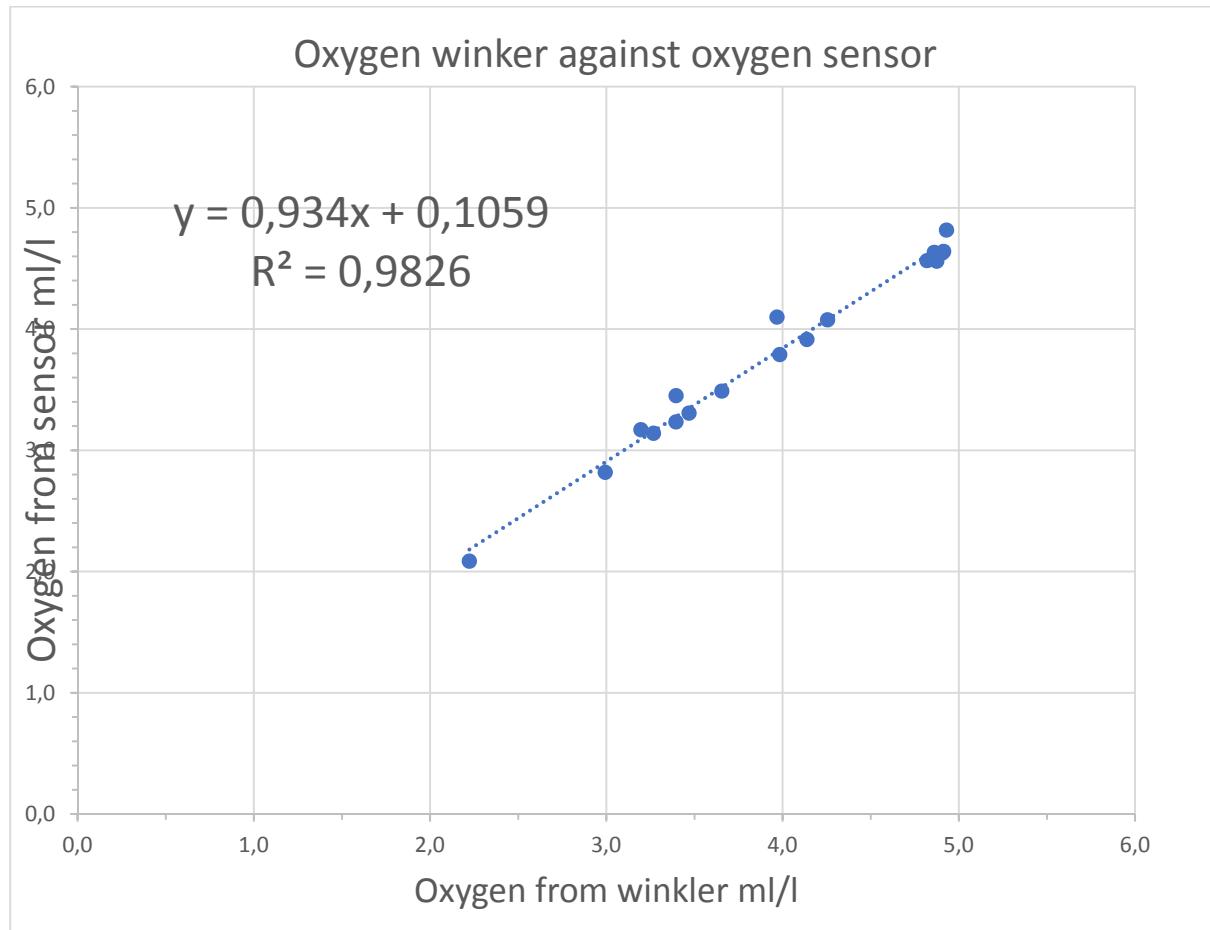


Figure 5. Oxygen sensor measurements on the CTD plotted against oxygen measurements performed on board.

2.2.5 Ocean acidification parameters (pH and alkalinity)

Seawater samples (250 ml) obtained at standard depths from the CTD-mounted Niskin-bottles were collected in borosilicate glass bottles using silicone tubing to reduce air exchange. Both pH and alkalinity were analysed on board the vessel. pH was determined spectrophotometrically using a diode array spectrophotometer and a pH sensitive indicator, m-cresol purple in 2 mM solution, as described by Clayton and Byrne (1993) and Chierici *et al.* (1999). Alkalinity was measured by titration with acid (0.05M HCl) and changes in pH were measured with an electrode (potential in mV) using tiamo software. Further processing of the data will be done on land by IMR and will provide more information on the marine carbonate system and parameters for ocean acidification.

2.2.6 Nutrients

Seawater samples (20 ml) for nutrient analyses (nitrate, silicate and phosphate) were collected from the Niskin water bottles (standard depths). The samples were stored in 20 ml

polyethylene vials, conserved with 0.2 ml chloroform, and kept cool and dark in a refrigerator. The analyses will be made on shore by IMR, using a modified Alpkem AutoAnalyzer C (O I Analytical, USA) and following standard procedures (Grasshoff, 1965). Storage may introduce loss of accuracy of the results, especially when the concentrations of nitrate, silicate and phosphate are low, such as in surface samples from the productive season. Samples from deep water are more stable, because most of the nutrients in the deep water occur in their inorganic form.

2.3 Plankton

2.3.1 Phytoplankton biomass

Chlorophyll-*a* was sampled as an indicator of phytoplankton biomass. For chlorophyll-*a* and phaeopigment measurements, seawater was collected from the Niskin-bottles at the standard depths (not below 200 meters). The water was filtered using a 0.7 µm filtration system (Munktell glassfiber filters Grade: MGF, vacuum 400 mm Hg) and stored at 20°C until analysis on shore by IMR. The assay was performed by extraction with 90% acetone followed by centrifugation, and the measurements were taken with a fluorometer (model 10 AU, Turner Designs Inc., Sunnyvale, Ca., USA), according to Welshmeyer (1994) and Jeffrey and Humphrey (1975).

2.3.2 Phytoplankton identification

Phytoplankton was collected along the hydrographic transects at stations positioned at bottom-depths of approximately 30 m, 100 m and 500 m. At each plankton-station, qualitative phytoplankton samples were collected with a net (35 cm in diameter and mesh-size of 10 µm), hauled vertically at a speed of 0.1 ms⁻¹ from the depth of 30 m to the surface (from ca. 5 m above bottom at the 30 m stations). The samples were preserved with 2 ml of 20 % formalin buffered with hexamine in 100 ml bottles (i.e. a final solution of ca. 0.4% formaldehyde). These samples are not quantitative but used to establish the taxonomic composition of the phytoplankton community.

2.3.3 Zooplankton sampling

Mesozooplankton was collected with a WP2-net along the hydrographic transects at stations positioned at bottom-depths of approximately 30 m, 100 m and 500 m. The WP2-net (56 cm diameter, mesh size 180 µm) (Fraser 1966, Anonymous 1968) was hauled vertically at a speed of ~0.5 ms⁻¹ at each station. At the shallowest and intermediately deep stations (bottom-depths of 30 m and 100 m, respectively), the sampling-strata were from near-bottom to the surface (lower sampling-depths of ~25 and 90 m, respectively). At the deepest stations with bottom-depth of ~500 m, the sampling-stratum was from the depth of 200 m to the surface.

Furthermore, a second collection with the WP2 net was performed for the depth-stratum of 30-0 m at the stations with bottom-depths of 100 m and 500 m. The purpose of these additional samplings was to enable a direct comparison of the zooplankton composition and concentrations in the uppermost layer of the water-column along the bottom-depth gradient. Each zooplankton-sample was divided into two equally large parts using a Motoda plankton

splitter (Motoda 1959). The first part of the sample was size-fractioned by using a series of sieves with the decreasing mesh-sizes of 2000 µm, 1000 µm and 180 µm, and the zooplankton retained on each sieve were thereafter dried on aluminium-trays at ~60 °C for 24 h. These samples will be dried once more and weighed on land for estimation of biomasses for the different size-groups. The second part of the sample was preserved in seawater with a final solution of 4% formaldehyde buffered with borax for subsequent species identification and quantification on land.

2.3.4 Fish eggs and larvae

Fish eggs and larvae were collected using a Hydro-Bios Multinet with 405 µm meshes. Samples were obtained along the hydrographical transects at approximately 30, 100 and 500 m depth. The net was towed obliquely from the bottom or a maximum depth of 100 m to the surface with a towing speed of 1.5 ms⁻¹. All fish larvae visible to "the naked eye" were removed from the total sample, transferred to vials and preserved in 4% borax buffered formaldehyde. After removing visible fish larvae, the Multinet sample was fractionated using a Motoda plankton splitter for enumeration of eggs under a stereomicroscope (Motoda 1959). The principle of this procedure is to split a homogenised sample into two "equal" parts, which again can be split further depending on the sample size. Small fish larvae overlooked by the "naked eye" scan were collected under the stereomicroscope and preserved in a separate vial noting the splitting factor. The remaining fish eggs were preserved in 4% borax buffered formaldehyde in vials noting the splitting factor. The rest of the sample was fixed for reference purpose and for possible later checks of overlooked egg and larvae. The fish-eggs will be sorted, and the larvae identified by regional laboratories (contact person: Margareth Kyewalyanga, University of Dar es Salaam, Tanzania).

2.4 Microplastics and debris

Microplastics are small pieces of plastic marine debris normally less than 5 mm long. Microplastics were collected at all hydrographical super-stations. At each station, the surface layer was sampled with a Manta-trawl having a rectangular opening of 19 cm × 61 cm (HxW), mesh-size 335 µm and two wings to keep it in balance and at the surface during the tow. Trawls were hauled horizontally at a speed of ~1.5 ms⁻¹ for 15 minutes. Trawling was performed on the right-hand side about mid-ship, a few meters away from the ship attempting to avoid the wake of the vessel. Geographical start and stop positions were recorded in the bridge-log. In addition, the counts of a flowmeter attached below the trawl opening were recorded at start and stop of each trawl event.

The Manta-trawl samples were washed in filtered sea-water over a sieve with mesh-size 180 µm. Microplastic particles were picked from the sample under a stereomicroscope. This was repeated twice to ensure detection of the microplastic particles. All assumed microplastic items were then placed on a gridded petri dish for examination under the stereo-microscope, photographed and, to the extent possible, also measured and described (e.g. length, shape, type and colour). The sorted microplastics were washed with distilled water and dried in pre-weighed aluminium-trays in a drying cabinet at 30 °C. The trays were packed in aluminium

foil, stored in room-temperature and shipped to the laboratory at IMR, where they will be studied in more detail. After removing the microplastics, the remaining part of the samples - mainly biological material - was preserved for studies of neuston.

2.5 Sediment sampling

Sediment samples were obtained at all bottom trawl stations. Stainless steel cylinders were mounted on the footrope of the trawl to collect bottom sediment samples at every trawl station. The samples were collected from the cylinder when the trawl was on deck and stored in plastic bags and preserved for further analyses of sedimentological and chemical composition.

2.6 Bottom mapping echo sounder

The EM 710 and EM 302 multibeam echo sounders are both belonging to a high to very high-resolution seabed mapping system. Data acquisition depth starts approximately 3 m below the transducers and the maximum acquisition depth is limited in practice to 1000–1500 m on *Dr Fridtjof Nansen*. Across track coverage (swath width) is up to 5.5 times water depth and may be limited by the operator either in angle or in swath width without reducing the number of beams. The operating frequencies are between 70 to 100 kHz. There are 128 beams with dynamic focusing employed in the near field. The transmitting fan is divided into three sectors to maximize range capability and to suppress interference from multiples of strong bottom echoes. The sectors are transmitted sequentially within each ping and use distinct frequencies or waveforms. The along track beam width is 1 degree. Ping rate is set according to depth. The receiving beam width is 2 degrees. Sound profiles were set manually in the system according to the area of operation. The data was logged to the on-board Olex plotting system.

2.7 Food safety and parasites

2.7.1 Food safety

Whole fish, fillet and different organs from various fish and octopus were sampled during this survey. At IMR, all the samples will be analysed for a wide variety of nutrients and contaminants. Some of the samples will also be analysed for correspondence between the microbiota and the metal content of the gut. One pelagic fish sample will be analysed for the content of microplastic particles.

2.7.2 Parasites

Whole fish, fillet and different organs from various fish that are regularly and irregularly consumed in the Tanzanian coast were sampled during this survey to detect the presence of parasites. All fish were measured (total body length, TL, 5 mm accuracy) and weighed (total body weight, TW, in g) before further parasitological analysis. Other host biometric parameters that were recorded on a routine basis included fish gender, state of maturity, as well as gross identification of stomach contents (empty/fish/crustaceans/mollusks/mud). All the samples were analysed for a wide variety of parasites, with a focus on those which are of concern to human health and seafood industry/fisheries (i.e. Anisakid nematodes and *Kudoa*

myxozoan). Members of both these groups include human pathogenic and/or impact fish quality reducing species. To date, “*Anisakis*” is the major biological hazard in seafood products (EFSA, 2010), since larval stages of *Anisakis* are etiological agents of human anisakiasis. On the other hand, *Kudoa* species may generate visible macroscopic cysts in the fish host’s muscle and cause *postmortem* tissue myoliquefaction, commonly referred to as ‘soft flesh’ of the fish host’s somatic musculature. Due to its repellent appearance and the spoilage of fish fillets, when it occurs in fish intended for human consumption, it can drastically reduce the marketability of the products.

The inspection procedure of nematode larvae applied in the present survey was based on the UV-press method. In brief, both flesh sides, i.e. fillets including belly flaps, and the visceral organs of each fish, were placed in separate clear plastic bags and then pressed in a hydraulic pressing device to a 1–2 mm thin layer. The bags were then deep-frozen for several hours to ensure proper core freezing, and, after thawing, examined under UV-light (366 nm). Any larvae present emerge as more or less brightly fluorescent spots or coils under UV-light. The larvae present in viscera and in the muscles were separately counted and stored in Eppendorf tubes at –20 °C until their further genetic identification to be achieved at IMR, Bergen.

Tissue samples from fish of the species *Thrichiurus lepturus* and *Scomberomorus commerson* were analysed also for the parasite *Kudoa*. Muscle samples from fresh fish were taken within 2 h after catch and stored in at –20 °C. After 48 hours, the musculature of the fish was examined for *post mortem* myoliquefaction, based on manual muscle texture testing and/or visual inspection of the muscle appearance. From every specimen showing light to clear signs of “soft flesh” at gross examination, one muscle tissue sample was taken and analysed with microscope of 100X magnification. When kudoid spores were detected, the entire mucle of fish was stored and preserved at –20 °C for further genetic identification at IMR. Tissue samples from mackerel samples will be analysed for the parasite *Kudoa*.

2.8 Top predators

There was no dedicated top predator observer on board during this cruise.

2.9 Trawl sampling

Biological sampling of fish was carried out using both pelagic and bottom trawls. Annex I gives a description of the instruments and the fishing gear used. In shallow waters (<30 m) or at night when pelagic fish was close to the surface, a small pelagic trawl with floats or a bottom trawl with floats were used for sampling. All catches were sorted, and subsamples taken to measure the number and weight by species. Species identification was based on the FAO Species Identification Guides. Individual fish were measured by total length (nearest cm below) and weight. For target species, 30 individuals were investigated with respect to sex, maturity stages and stomach fullness. In addition, for target species, the following biological samples were collected for later use: otoliths for aging, fin clipping for genetic analyses, stomachs for diet studies and liver for condition studies. Based on obtained measurements, length-weight relationships were established for acoustic estimates of the biomass of target species.

The list of species priority can be found in Annex II, and the complete records of fishing stations and catches are shown in Annex III. A full list of biological samples per species is given in Annex IV.

Jellyfish were caught as part of the trawl haul and identified to the lowest taxonomical level possible, and were counted and weighed. Jellyfish specimens that were in a good condition were photographed (top and bottom sections), before being processed and preserved for future analysis. A small piece of the oral arm tissue as well as one gonad was removed and preserved in 96% ethanol (EtOH) and stored at -20°C. Tissue samples stored in EtOH were collected for genetic studies, aimed at determining species and population structure, as well as establishing regional and global connectivity. The rest of the specimen was preserved in 10% formalin. These samples formed part of a greater morphological identification and taxonomic study.

2.10 Acoustic sampling

2.10.1 Sonar data

A Simrad SH90 Sonar was recording data continuously during the survey and stored for post processing after the survey. The sonar was set to a frequency of 26 kHz, in FM Normal mode. The sonar was operated using bow up/180 deg operation mode with the bearing of the vertical beams perpendicular to the vessel direction, and a horizontal range of 450 m and tilt angle of 3 deg. The filters built in the sonar software to improve the school representation (i.e. AGC, RCG) and ping to ping was set to default values, except for the Noise filter, which was turned off. The settings, including range and tilt, were kept the same throughout the survey, except during trawling operations when the sonar in some instances was used to target fish schools.

2.10.2 Acoustic estimates of fish biomass

Acoustic data were recorded using a Simrad EK80 Scientific Split Beam Echo Sounder equipped with keel-mounted transducers at nominal operating frequencies of 18, 38, 70, 120, 200 and 333 kHz. Annex I gives the details of the acoustic settings used during the survey.

Acoustic data were logged and post-processed using the latest acoustic data post-processing software, the Large-Scale Survey System (LSSS) Version 2.0. In cases where the target category of fish contains more than one species, the mean s_A -value allocated to the category is divided between the species in the same ratio as their contribution to the mean back scattering strength in the catches (relative amount by number at length in the catches).

The following target strength (TS) function was applied to convert s_A -values (mean integrator value for a given species or group of species in a specified area) to number of fish:

$$TS = 20 \log L - 72 \text{ dB},$$

which can be converted (see Toresen *et al.* 1998 for details) to the area form (scattering cross sections of acoustic targets):

$$C_{Fi} = 1.26 \cdot 10^6 L^{-2},$$

where L is total length in 1 cm length group i and C_{Fi} (m^{-2}) is the reciprocal back scattering strength, or so-called fish conversion function. In order to split and convert the allocated s_A - values (m^2/NM^2) to fish densities (numbers per length group per NM^2), the following formula was used:

$$\rho_i = s_A \cdot \frac{p_i}{\sum_{i=1}^n \frac{p_i}{C_{Fi}}}$$

where

ρ_i = density of fish in length group i

s_A = mean integrator value

p_i = proportion of fish in length group i

$\sum_{i=1}^n \frac{p_i}{C_{Fi}}$ = the relative back scattering cross section (m^2) of the length frequency sample of the target species, and

C_{fi} = reciprocal back scattering cross section (σ_{bs}^{-1}) of a fish in length group i .

The integrator outputs were split in fish groups using a combination of behaviour pattern as deduced from echo diagrams, the LSSS analysis and catch composition as described below.

Table 5 shows the target species/groups that were used.

Table 5. Allocation of acoustic densities to species groups. Note that only examples are listed for the various groups.

Group	Taxon	Species
Pelagic species group 1 (PEL1)	Clupeidae	<i>Dussumieri a acuta</i>
	Engraulididae	<i>Stolephorus</i> spp. <i>Encrasicholina heteroloba</i> <i>Thryssa</i> spp.
Pelagic species group 2 (PEL2)	Carangidae	<i>Carangoides</i> spp. <i>Decapterus</i> spp.
	Scombridae	<i>Rastrelliger kanagurta</i> <i>Scomber japonicus</i> <i>Scomberomorus commerson</i>
	Sphyraenidae	<i>Sphyraena</i> spp.
	Trichiuridae	<i>Lepidorhynchus caudatus</i> <i>Trichiurus lepturus</i>
Other demersal species	Demersal families	
Mesopelagic species	Myctophidae	
	Other mesopelagic fish	
Plankton	Calanoidae	<i>Calanus</i> sp.
	Euphausiidae	<i>Meganyctiphanes</i> sp.
	Other plankton	

The equations above show that the conversion from s_A -values to number of fish is dependent on the length composition of the fish. It is therefore important to get representative length distributions from the stock in the whole distribution area.

When the size classes (of e.g. young fish and older fish) are well mixed, the various length distributions can be pooled together with equal importance. Otherwise, when the size classes are segregated, the total distribution area is post-stratified, according to the length distributions, and separate estimates are made for the regions containing fish with equal size.

For a region representing a distribution of a target-species, the following basic data are needed for the estimation of abundance:

- 1) The average s_A -value for the region,
- 2) The surface (usually square nautical miles, NM^2), and
- 3) A representative length distribution of the fish in the region.

If the targeted fish consists of more than one species, a representative length distribution of each species within the region, as obtained from the trawl catches, are used. A length distribution representing the number of the various species for each catch must be calculated.

Thereafter, these distributions must be normalized to a unit number (usually 100) so they are equally weighted (independent of sample size).

A systematic approach to a) divide the s_A -value between species in a category of fish and b) produce pooled length distributions of a target species for use in the above equation and c) calculate the biomass estimates for a region, is obtained through the following procedure:

- The samples of the species in the category are respectively pooled together with equal importance (normalized).
- The mean back scattering strength (ρ/s_A) of each length frequency distribution of the target species is calculated and summed. This is automatically done in the Excel spread-sheet made available for acoustic abundance estimation on board R/V *Dr Fridtjof Nansen*, provided the data are punched in this sheet.
- The mean s_A -value allocated to the category of fish in the region is divided between the species in the same ratio as their relative contribution to the mean back scattering strength of the length groups in the sample representing the region
- The pooled length distribution is used, together with the mean s_A -value, to calculate the density (numbers per square NM) by length groups and species, using the above formula. The total number by length group in the area is obtained by multiplying each number by the area.
- The numbers are then converted to biomass using the estimated weight at length.

The combination of low s_A value recorded and few PEL1 and PEL2 in the bottom trawl catch and few pelagic trawls made the splitting by length groups unreliable. For PEL1 an overall average of 9 cm was applied in the estimates of biomass, and for carangids and associated species (PEL2) an overall average of 23 cm was used.

2.11 Bottom trawl estimates of abundance

In the bottom trawl survey, stock biomasses were estimated by the swept-area method with catch per haul as the index of abundance (see Strømme 1992). In most hauls the trawling time (with the gear at the bottom) was around 30 min. The area swept by the trawl net within 30 minutes trawl time was 0.015 NM² and it corresponds to an average horizontal trawl opening of 18.5 m efficient net width, towing at 3.0 knots. Diagrams of the bottom trawl used are shown in Annex VI. The general formula to estimate biomass B, using this method is:

$$B = \frac{A}{a} \cdot \frac{\bar{X}}{q} \quad (6)$$

A is the total area surveyed, a is the swept area of the net per haul, \bar{X} is the average catch per haul (the index of abundance) and q (trawl catchability) is the proportion of fish in the path of

the net that are actually caught. The density of the resource is estimated as biomass per unit area. In a stratified survey of k non-overlapping strata, if the mean catch per haul in stratum i and its variance are denoted by \bar{X}_i and s_i^2 respectively, then an unbiased estimate of the population mean \bar{X} is the stratified mean \bar{X}_{st} , which is given by:

$$\bar{X}_{st} = \frac{1}{N} \sum_{i=1}^k N_i \bar{X}_i = \sum_{i=1}^k W_i \bar{X}_i \quad (7)$$

where $W_i = \frac{N_i}{N} = \frac{A_i}{A}$ is the relative size of the i^{th} stratum (A_i is the area of the i^{th} stratum and A is the total area surveyed). The variance of the stratified mean is given by

$$\text{var}(\bar{X}_{st}) = \sum_{i=1}^k W_i^2 \text{var}(\bar{X}_i) = \sum_{i=1}^k W_i^2 \frac{s_i^2}{n_i} \quad (8)$$

where n_i is number of hauls in the i^{th} stratum and n is the total number of hauls in the survey. Table 6 shows the areas used in the swept-area method to estimate biomass for the different regions. A stratified semi-random design was used with depth and area as stratification factors. Estimated total biomass by species/group was obtained by summing estimates for each depth stratum.

Table 6. Areas in nm² for the various regions and depth strata.

DEPTH STRATA (NM²)	The southern area	Mafia	Zanzibar	Pemba
20 – 50	312	2 271	3 315	429
50 – 100	328	906	1 116	228
100 – 200	525	866	564	355
200 – 500	1 916	5 160	1 478	3 221
200 – 1 000	5 538	12 038	4 065	7 745

For conversion of catch rates (kg/hour) to fish densities (t/NM²), the effective fishing area was considered as the product of the wing spread and the haul length, or distance over the bottom, as measured by means of the SCANMAR® equipment based on GPS readings. The area swept for each haul was thus 18.5 m (traditionally applied wing spread for the “Nansen” bottom trawl) times the distance trawled, raised to NM²/hour. The catchability coefficient (q), i.e. the fraction of the fish encountered by the 18.5 m horizontal opening of the trawl that was actually caught, was assumed equal to 1, which leads to an estimation of the minimum biomass for comparison with previous surveys. Catchability may vary depending on the type of gear used and the type of species (e.g. gears with bobbins are apparently less efficient for species such as flat fishes and octopus, while gears without bobbins and with footrope touching the bottom are more efficient for benthic species). Departures of q from 1 can introduce biases in biomass estimates leading to wrong fisheries management advices (David Somerton, 1996).

CHAPTER 3. SURVEY RESULTS

3.1 Hydrographical conditions

Hydrographical data were collected at fixed CTD stations to 1 000 m depth, and from the Aanderaa weather station that continuously collect sea surface temperature, wind speed and direction, solar radiation etc. during the survey (not presented).

3.1.1 Data from the CTD sensor

The CTD has sensors for measuring salinity, temperature, fluorescence and oxygen at approximately every meter, thus giving a vertical profile. Measurements were performed at 65 CTD stations, and seven “super transects” were carried out (Figure 6).

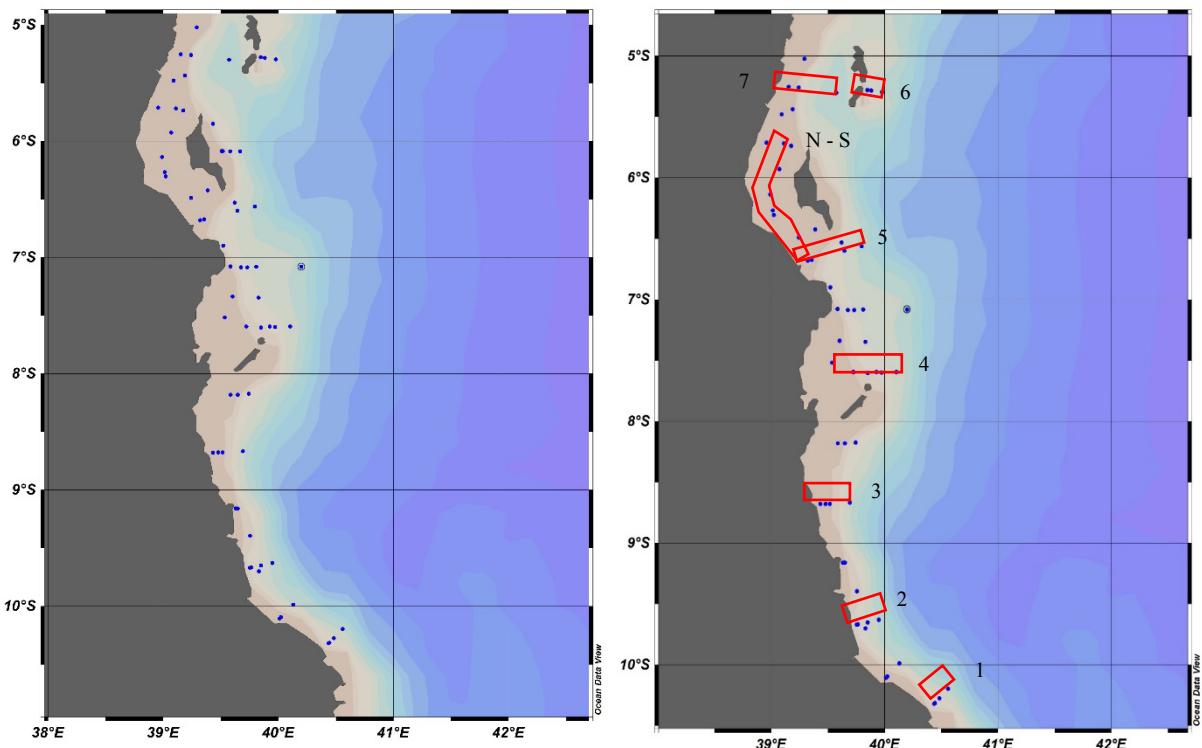


Figure 6. Map showing the position of the 65 CTD stations (left side) and the 7 selected “super transects” during survey 2018404, Tanzania.

In Figures 7 and 8 we present the different sensor parameters for “super transect 4”. Figures from all transects are given in Annex VII. Transect 4 is situated outside the Rufiji River just north of Mafia. In this area the continental shelf is substantially wider than typical for the coast of Tanzania. At the shallow stations on the shelf, both temperature and salinity were more or less identical from surface to the bottom (Figure 7). Above deeper waters, high temperatures were measured from the surface to ~65 m ($>25^{\circ}\text{C}$) at the edge of the shelf. With increasing distance offshore, this warm surface layer became gradually shallower and extended down to a depth of 30 m at the outmost station. The salinity showed much the same pattern, with mixing from surface to bottom at the shelf. At the deeper stations the salinity was <35 PSU down to 50-60 m, whereas at the outmost station of transect 4, the salinity was

~35 PSU at the surface. Outside the shelf there was an intermediate layer with high salinity water (>35.1 PSU), from ~60 m to 300 m, with lower salinity above and below. This pattern was observed at all transects, excepted for at the southern transect (transect 1) where the whole water column was mixed. The salinity differences may be influenced by the four different water masses in the Tanzanian offshore waters, having different levels of salinity (and oxygen): Tropical surface water, Arabian sea water, Antarctic intermediate water, and North Indian deep water.

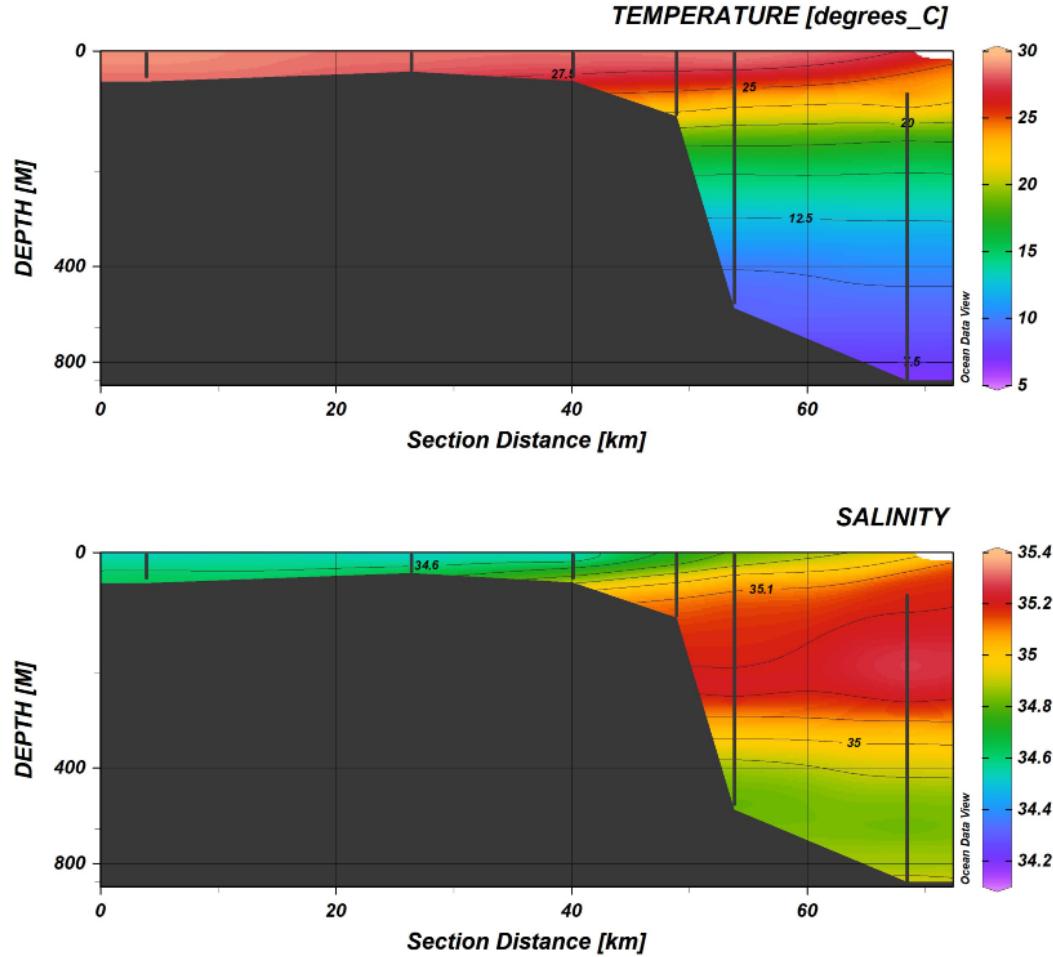


Figure 7. Sensor data (CTD) for salinity (PSU) and temperature ($^{\circ}$ C) along “super transect 4”.

The vertical profiles along the transect for oxygen and fluorescence are given in Figure 7. In the upper well mixed waters the oxygen concentration was >4 ml/l (0 to ~65 m). In the intermediate water (60 m to 560 m) the oxygen concentration was between 3 and 4 ml/l. In the deep water (> 650 m) there were measured low oxygen concentrations (2.5 – 1.5 ml/l). Similar to salinity concentrations above, the four water masses off the coast of Tanzania also influence the levels of oxygen concentration along the coast.

Fluorescence gives a proxy for phytoplankton biomass. At all transects, higher fluorescence was measured above the shelf (Figure 8), whereas for most of the stations in deeper waters

(outside the shelf) the fluorescence values indicate low biomass of phytoplankton. At most stations the fluorescence data indicates that most of the phytoplankton biomass are located from 0 to 60 m.

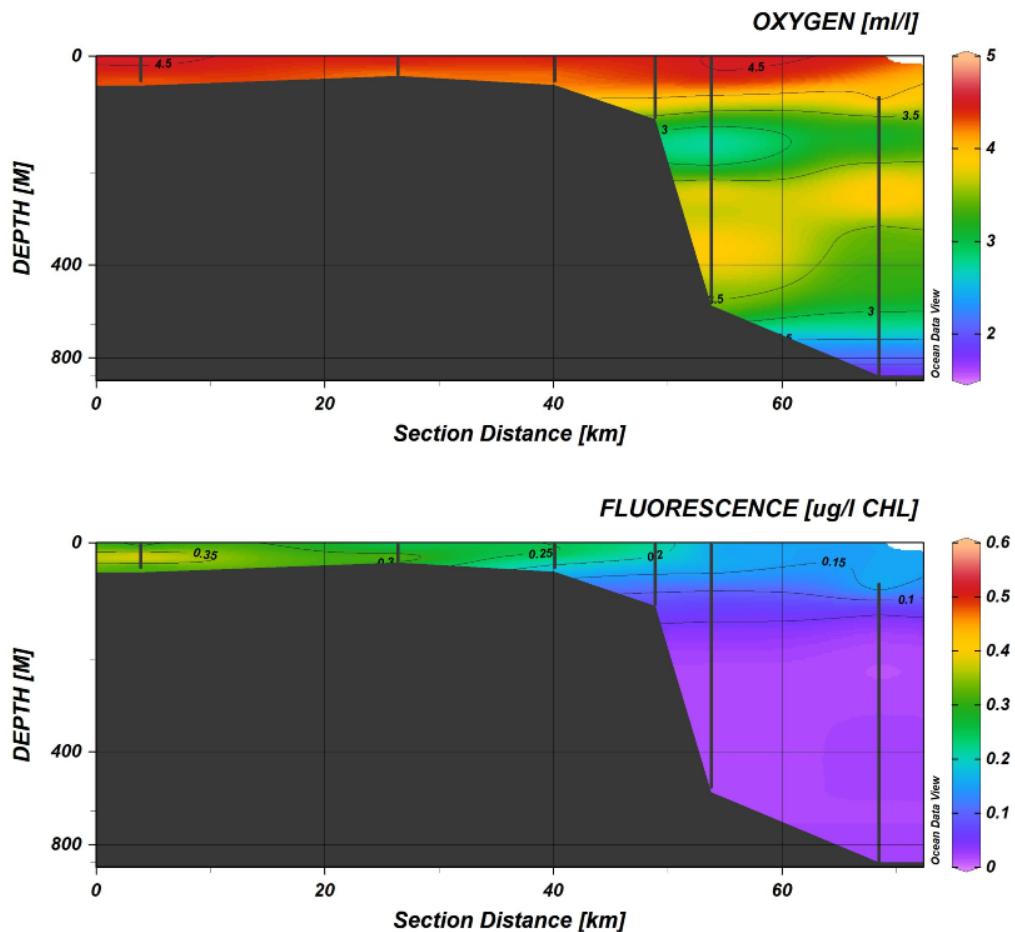


Figure 8. Sensor data (CTD) for oxygen (ml/l) and fluorescence (ug/l Chl) along “super transect 4”.

Based on all CTD stations shown in the map (Figure 6, left), surface plots are given in Figure 9 for temperature, salinity, and fluorescence, and the concentration of oxygen in the deepest sampling depth.

The salinity data shows two areas with lower salinity in the surface water, one at the southern part (transect 1) and one area north of Mafia (transect 4). Both areas are situated close to the mouths of large rivers (Ruvuma in the south and Rufiji near Mafia). For the other stations, there were only minor differences. At the outmost station outside Mafia, the measured salinity was higher, indicating transport from oceanic water to the coast.

Based on surface temperature, the Tanzanian coast could be divided into three areas. South of Dar es Salaam, close to the coast, the surface temperatures measured were highest. North of Dar es Salaam the temperatures were lower in the surface with a small area west of Pemba that had even much lower surface temperature. East of Mafia lower temperatures were measured at the outmost stations, in the same area where higher salinity was measured.

The surface plot of fluorescence shows highest biomass of phytoplankton inside the island of Mafia and north of Dar es Salaam between Zanzibar and the main land. In addition, there was higher fluorescence at station in transect 1 (southern part of Tanzania).

The surface plot of oxygen in the bottom water shows concentrations >3 ml/l along the coast. However, at the deeper station in open waters the concentration of oxygen was <2.5 ml/l. The extend of this low oxygen water should be followed up to monitor the changes over time as well as the horizontal and vertical distribution of these water masses.

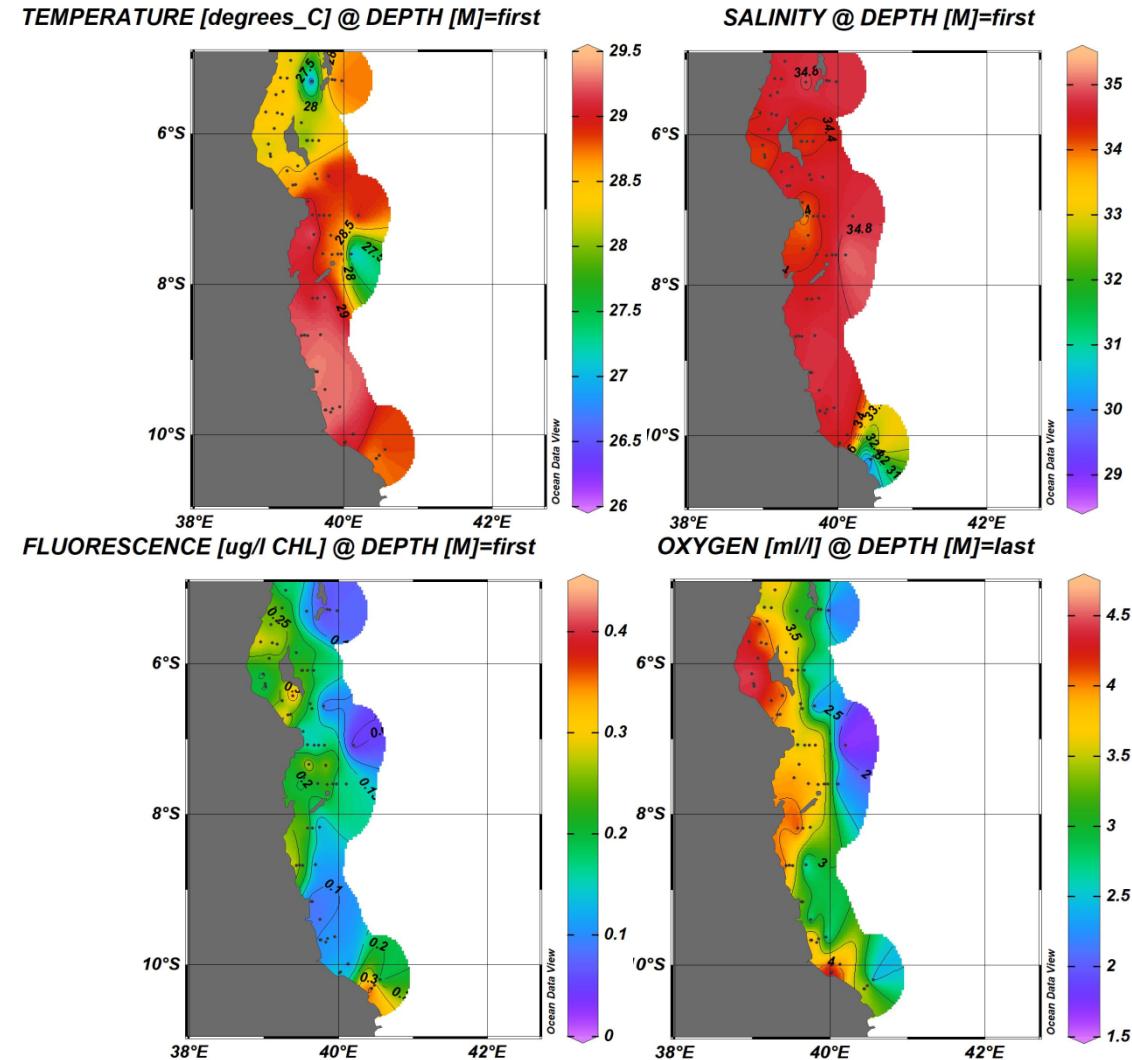


Figure 9. Surface plots of temperature, salinity, fluorescence and bottom concentration and oxygen using all CTD stations in the survey.

3.1.2 pH cross shelf distribution

Cross shelf distribution of pH is shown in Figure 10. In general, the pH was highest in the surface and decreased gradually with depth. In the south (Transect 1-3), high pH values (7.9 - 8.1) were recorded down to approximately 100 m depths. Further north (Transect 4-7),

high pH values were only recorded down to approximately 50-75 m depths, indicating a shallower mixed layer. In the deep-water masses between 500-1000 m depth, very low pH (7.5 - 7.6) values were measured.

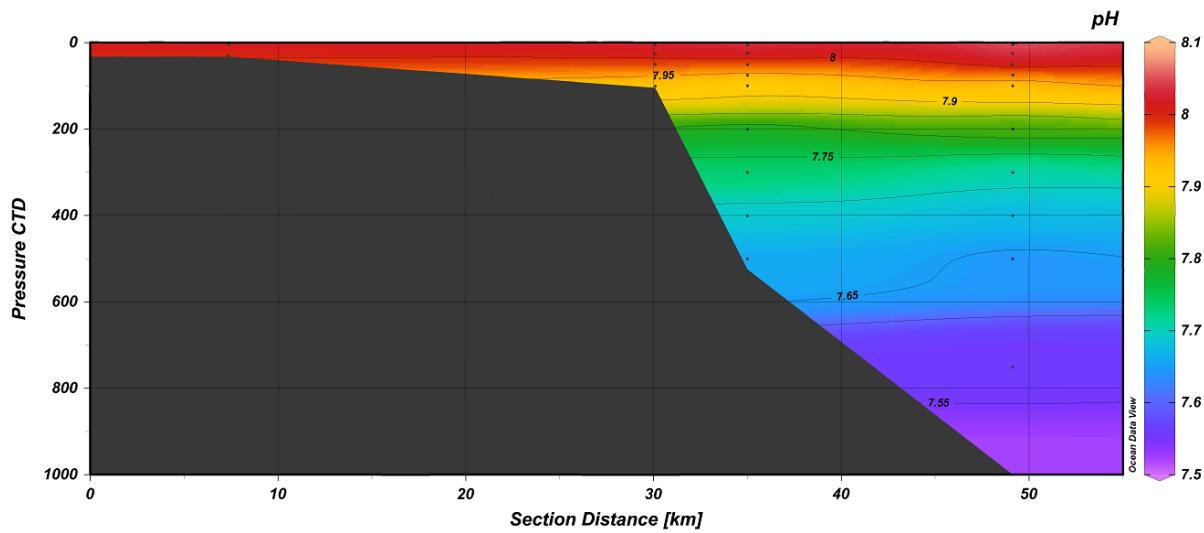


Figure 10. Cross shelf distribution of pH from Transect 4 (7.6 °S).

3.2 Microplastics

Samples for microplastics were collected at the surface using a Manta trawl retaining particles >335 µm. Microplastic particles were enumerated using a stereo microscope. Highest numbers of particles were observed outside Dar es Salaam and northwards between Zanzibar and the main land up to the northern part of Zanzibar (Figure 11). Intermediate numbers of particles were observed between Mafia and Zanzibar. Lower number of particles were recorded south of Mafia.

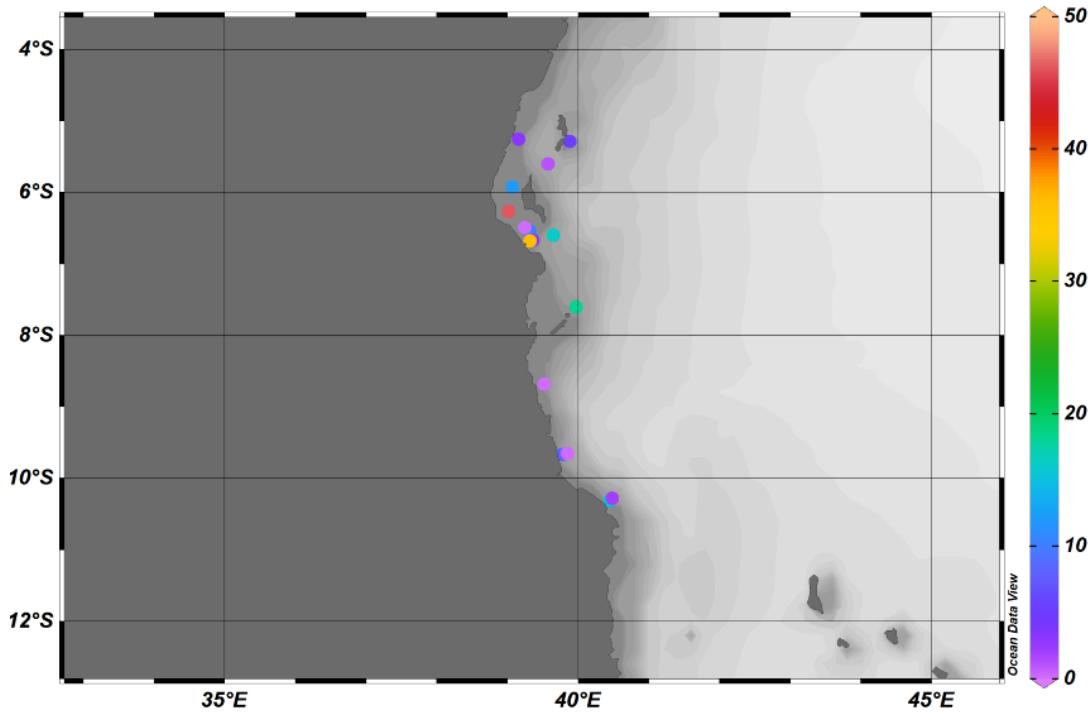


Figure 11. Microplastics collected at “super stations”. The figure gives number of observed microplastic particles in the samples.

3.3 Fish larvae and eggs

Samples for fish eggs and larvae were collected from 100 m, or bottom depth if shallower, to 0 m using a multinet with a mesh size of 305 µm. The highest numbers of fish larvae and eggs were observed from Dar es Salaam, though the Zanzibar Channel, and all the way up to Pemba (Figure 12). In addition, relatively high number of fish larvae were observed at one station on “super transect 3”, south of Mafia. In the southern part and at stations in open waters there were few observations of fish larvae and eggs.

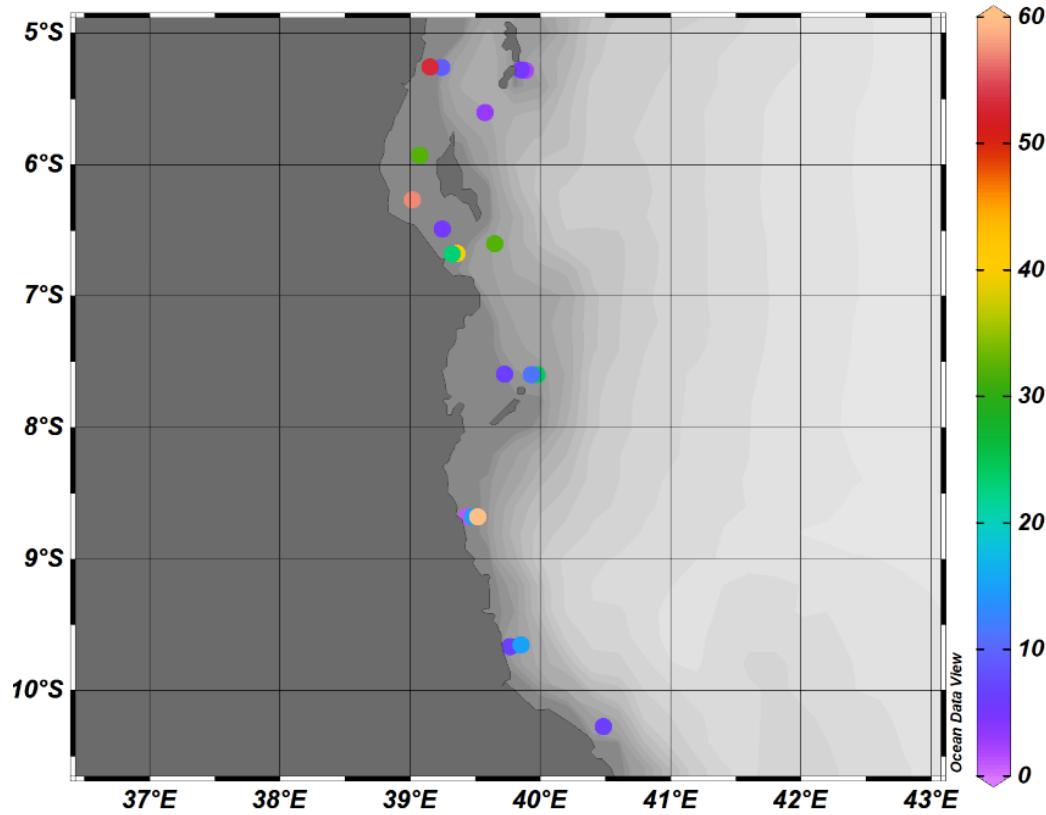


Figure 12. Numbers of fish larvae observed in the up part of the water column (<100 m).

3.3.1 Nutrients

Nutrient samples are undergoing analysis in the IMR laboratory in Norway, and data distributed once these analyses have been completed.

3.4 Plankton

3.4.1 Phytoplankton

Phytoplankton samples have not yet been analysed.

3.4.2 Chl α

This parameter is undergoing analysis at IMR in Norway, and the data distributed once these analyses have been completed.

3.4.3 Zooplankton

Zooplankton samples are undergoing analysis at IMR in Norway, and the data will be shared once these analyses have been completed.

3.4.4 Food safety and parasites

Samples for food safety and parasites are undergoing analysis at IMR in Norway and the data distributed once these analyses have been completed. Preliminary results of the parasite studies are presented in Table 4.

3.5 Acoustic abundance estimates and distribution

The acoustic survey covered the shelf and slope from roughly 20 m (but see 1.4 Narrative) to 1 000 m bottom depth. Acoustic distribution and abundance were estimated for two groups of species, Pelagic 1 and Pelagic 2. Pelagic 1 consists of pelagic fish of the families Clupeidae and Engraulididae, while the Pelagic 2 species consist of the families Carangidae, Scombridae, barracuda and hairtails. Table 5 gives an overview of the most common species belonging to each of these groups. The Pelagic 1 species are typically separated from the Pelagic 2 species based on the presence of the two groups in the trawl catches, and on the acoustic signal as seen during the scrutinizing process, e.g. the fact that the Clupeidae and Engraulididae have a much stronger backscattering signal than the Carangidae and other Pelagic 2 species.

Pelagic 1 species were observed in the Mafia area and the Zanzibar Channel (Figure 13), with estimated abundances of 516 and 6532 metric tons, respectively (Table 7). Both bottom and pelagic trawl samples suggested that the shorthead anchovy (*Enchrasicholina heteroloba*) was the most abundant species (Table 9 and 10). It should be noted that the survey did not cover more shallow areas where there is indication of substantially higher densities of Pelagic 1 species than over deeper waters.

Table 7. Abundance estimates (tons) of Pelagic 1 species in various areas.

The Southern area	Mafia	Zanzibar	Pemba	Total
0	516	6 532	0	7 048

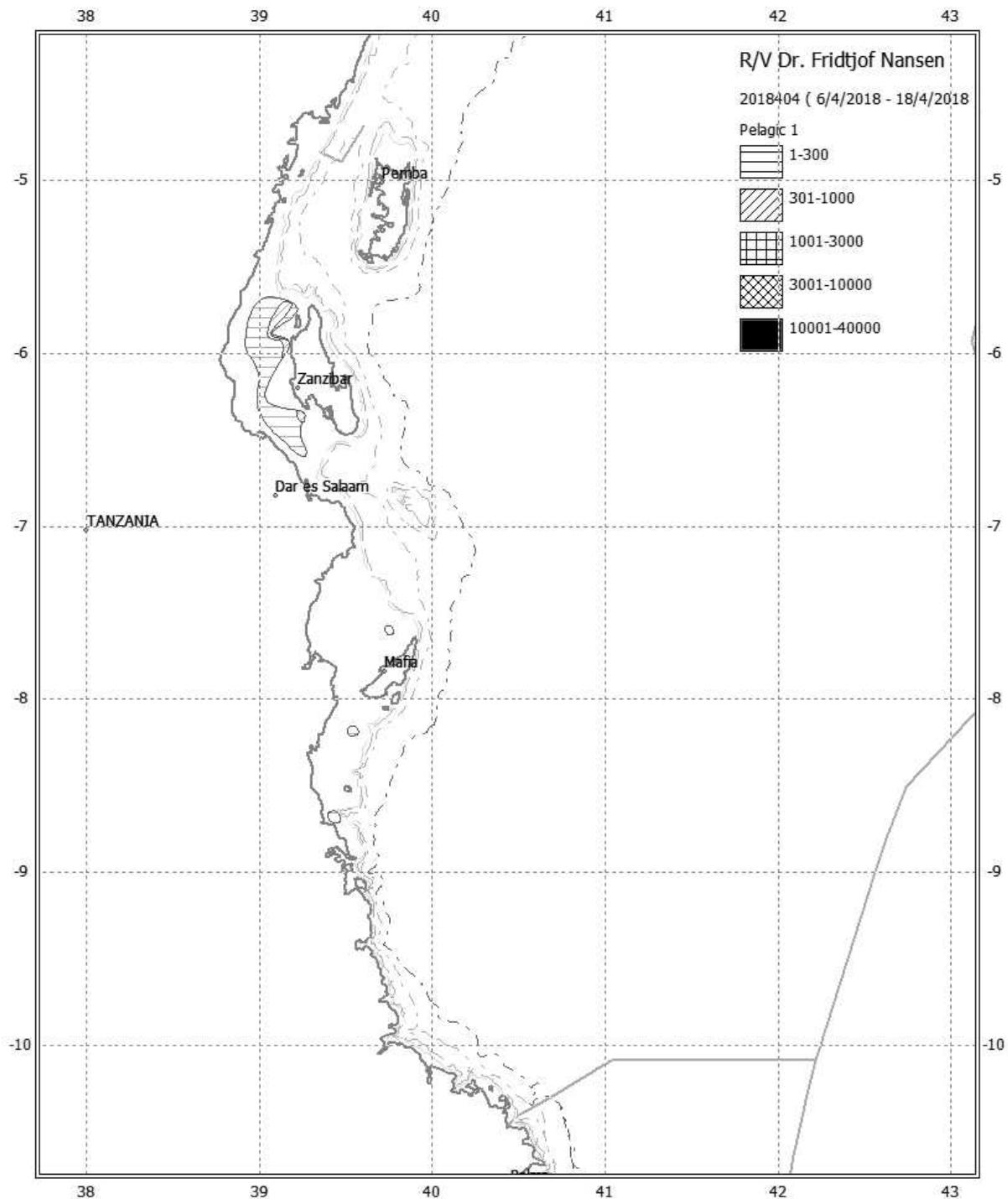


Figure 13. Distribution and density of Pelagic 1 species. Depth contours.

Pelagic 2 species were found in a narrow belt in the Southern area in relatively low concentrations, in two smaller areas in near the Mafia island and the highest abundances in the Zanzibar Channel (Figure 14 and Table 8). In the bottom trawl, the largehead hairtail (*T. lepturus*) was the most abundant Pelagic 2 species (Table 9).

Table 8. Abundance estimates (tons) of Pelagic 2 species in various areas.

The Southern area	Mafia	Zanzibar	Pemba	Total
4 112	2 197	18 816	0	25 125

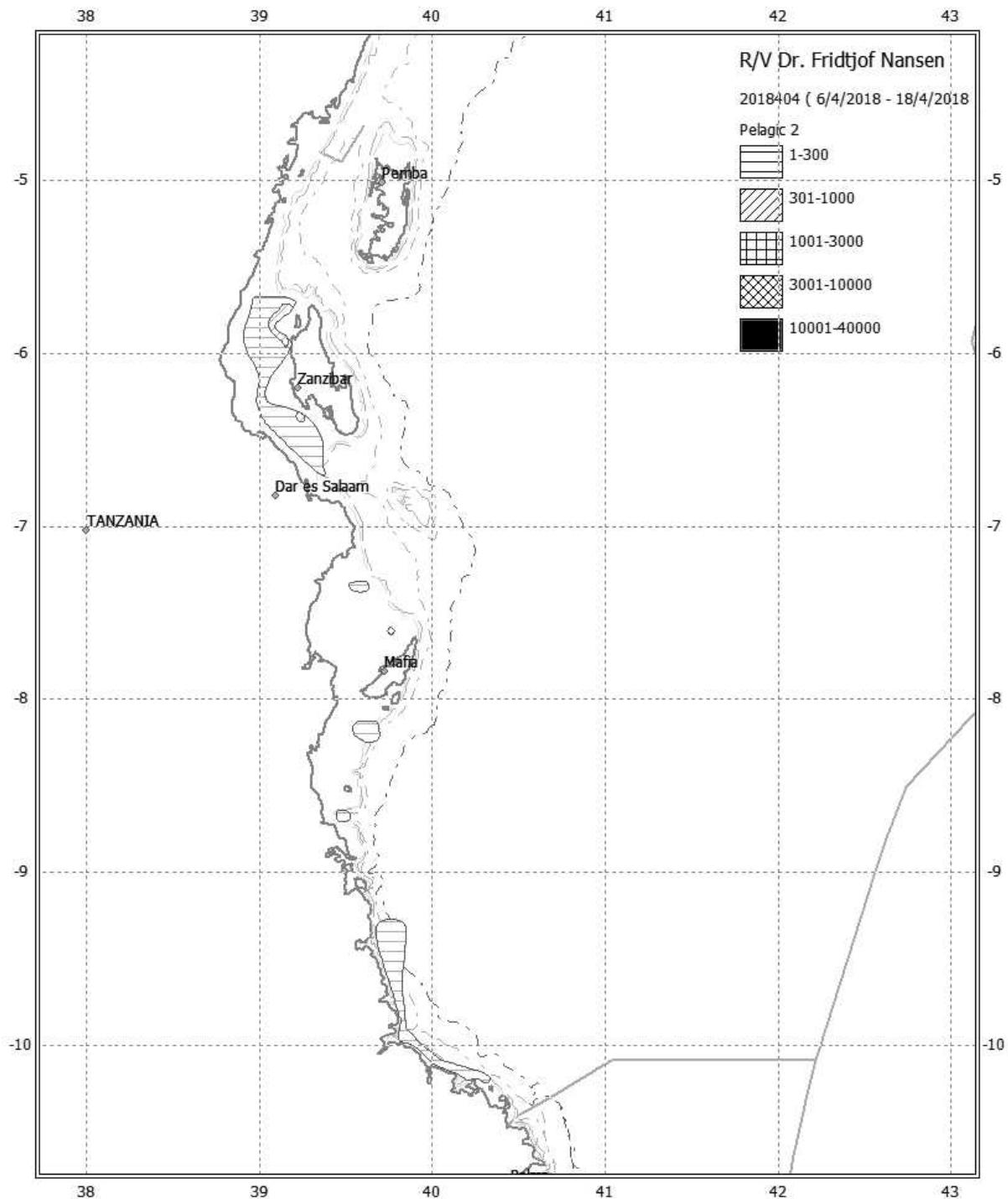


Figure 14. Distribution and density of Pelagic 2 species.

3.6 Swept area abundance and distributions

Because of the narrow shelf and “hilly” bottom along the Tanzanian coast, we were not able to carry out the planned swept area survey and have therefore not given estimates of bottom fish biomass. Instead we focused on aspects of fish diversity.

3.6.1 Bottom trawl catches

Table 9 shows the summarised catches in terms of weight and numbers, and no. of hauls with catch of the 25 most abundant species (in terms of weight) in 30 bottom trawl hauls along the coast of Tanzania. The abundance of all species caught in the bottom trawl is presented in Annex V. All hauls were standardised to one hour of trawling (actual time was ~30 minutes) Mean catch per hour can thus be estimated by dividing the catch by 30 hours.

The catch varied between 79 and 1 539 kg/h, with an average of 335 kg/h. There was no clear pattern between catch and depth (Figure 15a, the highest value in shallow waters was from a haul where a school of *E. heteroloba* was targeted). Number of species per haul varied between 13 and 45, with an average of 27.2. There was no relationship between number of species and depth (Figure 15b).

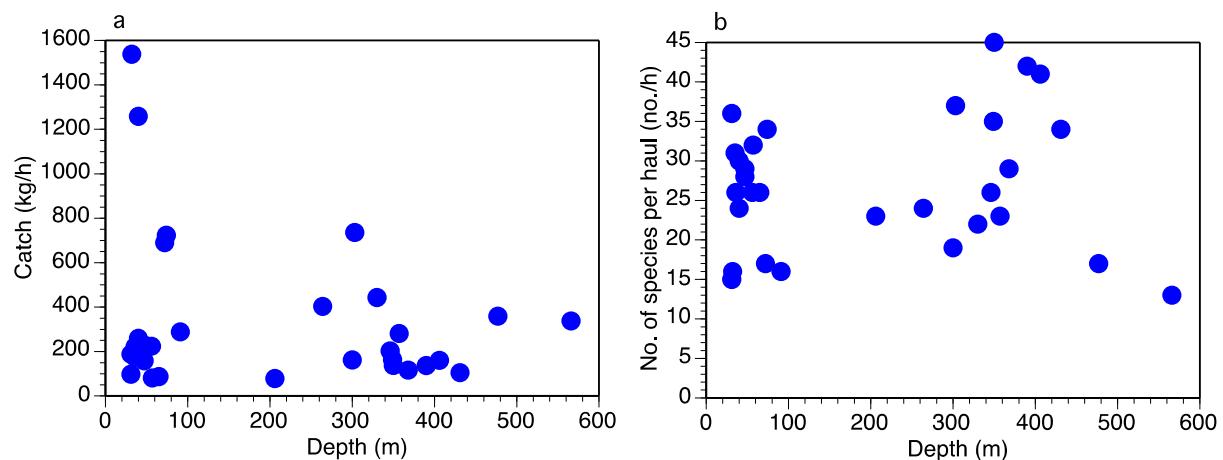


Figure 15. a) Catch vs. depth and b) no. of species per haul vs. depth.

Table 9. Summarized weight, number of fish and number of hauls with catch of the most abundant species (in terms of weight) in 30 bottom trawl hauls.

Species	Catch (kg)	No. of fish	No. of hauls
<i>Encrasicholina heteroloba</i>	1 397.02	49 9078	2
<i>Trichiurus lepturus</i>	697.1	1 227	4
<i>Upeneus taeniopterus</i>	620.3	24 584	6
<i>Decapterus macrosoma</i>	559.4	18 840	8
<i>Saurida undosquamis</i>	374.08	3 036	25
<i>Rexea prometheoides</i>	353.37	4 583	8
<i>Secutor insidiator</i>	307.58	19 823	3
<i>Carangoides malabaricus</i>	278.95	6 178	8
<i>Argentina euchi</i> s	231.72	4 025	8
<i>Leiognathus elongatus</i>	192.99	16 418	7
<i>MYCTOPHIDAE</i>	173.76	29 088	8
<i>Nettastoma parviceps</i>	165.59	1 957	2
<i>Polymixia berndti</i>	153.42	6 497	11
<i>Centrophorus granulosus</i>	136.42	67	3
<i>Zenion sp.</i>	124.4	1 232	1
<i>Aluterus monoceros</i>	115.41	46	2
<i>Upeneus moluccensis</i>	113.28	3 530	5
<i>Gazza minuta</i>	105.32	5 570	3
<i>Decapterus russelli</i>	103.31	1 462	8
<i>Polysteganus coeruleopunctatus</i>	102.53	97	4
<i>Abalistes stellatus</i>	93.09	148	9
<i>Himantura jenkinsii</i>	89.86	8	2
<i>Scomberomorus commerson</i>	85.44	54	4
<i>Leiognathus berbis</i>	84.48	41 814	5

A total of 335 species/groups were caught in bottom trawl during the survey, mainly fish but also some invertebrates (Annex V). Shorthead anchovy (*E. heteroloba*, Pelagic 1 species) contributed to the highest catches (Table 9). However, this species was only caught in two hauls, of which one was targeting a school. From the high no. of fish (column “No. of fish” in Table 9) it is evident that Shorthead anchovy is very small species (mean length was ~9 cm). The second highest catches consisted of largeheaded hairtail (*T. lepturus*, Pealgc 2 species), but this species too was caught in only a few hauls (4). On the other hand, most of the catch was taken at station 21 and 22 (Fig 3a), just south of Dar es Salaam at 270-300 m depth, with 453 and 185 kg/h, respectively. Hence, even though the overall abundance of largeheaded hairtail appears to be relatively low (avg. 23.2 kg/h), there are potentially local concentrations that may sustain a profitable fishery. This may be the case with other species as well.

On average, the 335 species/groups appeared in 2.4 trawl hauls (mean of values in the column “No. of hauls” in Annex V), which corresponds to 8% of the hauls. Hence, the majority of species appear to be relatively rare and have low overall abundance. Brushtooth lizardfish (*S. undosquamis*), on the other hand, was widely distributed as it was present in 25 of 30 hauls, but abundance was relatively low (12.5 kg/h).

CHAPTER 4. OVERVIEW AND SUMMARY OF RESULTS

The survey was carried out with 29 scientific personnel, coming from eight different countries. Everybody spoke English, and neither linguistic nor cultural differences posed any limits on collaboration or friendship among the participants. All work was carried out enthusiastically and with emphasis for high quality output, and the collaboration with the crew on board *Dr Fridtjof Nansen* was excellent.

Data collection during this survey along the coast of Tanzania was extensive, covering a wide range of scientific fields related to marine biology and ecology: hydrography, meteorology, microplastics, phytoplankton, zooplankton, jellyfish, fishery biology including eggs and larvae, genetics and fish abundance. Much of the data has not yet been analysed. Hence, this survey report only provides an overview of what has been done during the survey and presents some preliminary results, without the discussion of the results.

The hydrographic conditions were quite similar along the coast of Tanzania with high temperature in the upper water column (0 to ~60 m) ranging between 25 and 30°C. Surface water salinity was quite uniform, except for lower salinities close to the mouth of River Ruvuma on the southern border of Tanzania and River Rufiji near Mafia. In the upper well mixed waters the oxygen concentration was >4 ml/l (0 to ~65 m), in intermediate waters (60 m to 560 m) between 3 and 4 ml/l, whereas in deep waters (> 650 m) low oxygen concentrations were measured (2.5 – 1.5 ml/l).

Fluorescence measurements indicated low algal concentrations throughout the area, but with slightly elevated values north of Dar es Salaam, in the Zanzibar Channel, inside Mafia and near the mouth of River Ruvuma in the south. pH values were highest in surface and decreased with depth.

Highest numbers of plastic particles were observed outside Dar es Salaam and northwards between Zanzibar and the main land up to the northern part of Zanzibar.

The highest numbers of fish larvae and eggs were observed from Dar es Salaam, through the Zanzibar Channel, and up to Pemba.

Acoustic measurements of pelagic fish abundances were recorded throughout the survey. In general, the abundances were low, in particular over deeper waters outside the continental shelf. Pelagic 1 species were observed in the Mafia area and the Zanzibar Channel, with estimated abundances of 516 and 6 532 metric tons, respectively. Both bottom and pelagic trawl samples suggested that the shorthead ancovy (*E. heteroloba*) was the most abundant species. It should be noted that the survey did not cover more shallow areas where there is indication of substantially higher densities of Pelagic 1 species than over deeper waters. Also, the survey did not include lager fish, tuna and tuna-like fish in offshore waters.

Pelagic 2 species were found in a narrow belt in the Southern area in relatively low concentrations, in two smaller areas in near Mafia island and the highest abundances in the

Zanzibar Channel. Total biomass was estimated to ~25 kt. In the bottom trawl catches, the largehead hairtail (*T. lepturus*) was the most abundant Pelagic 2 species.

Because of the narrow shelf and “hilly” bottom along the Tanzanian coast, we were not able to carry out the planned swept area survey. A total of 335 different species/groups were caught in 30 bottom trawl hauls, mostly fish but also some invertebrates. On average, the 335 species/groups appeared in 2.4 trawl hauls, which corresponds to 8% of the hauls. Hence, the majority of species appear to be relatively rare and have low overall abundance. However, relatively high catches of largeheaded hairtail (*T. lepturus*, Pelagic 2 species) were taken in two hauls south of Dar es Salaam (453 and 185 kg/h). Hence, even though the overall abundance of largeheaded hairtail appears to be relatively low (avg. 23.2 kg/h), there are potentially local concentrations which may sustain a profitable fishery. This may be the case with other species as well.

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ANNEX I.

DESCRIPTION OF INSTRUMENTS AND FISHING GEAR

Acoustic instruments

The Simrad EK80/18, 38, 70,120, 200 and 333 kHz scientific sounder was run during the survey. Scrutinizing was done in LSSS using the data from the 38 kHz transducer. Last standard sphere calibrations was checked on the 23.01.2017 in Sandviksflaket, Bergen, Norway using Cu-64 for the 18 kHz, Cu-60 for the 38 kHz, WC-38.1 for the 70, 120 and 200 kHz, and the WC-22 for the 333 kHz. The details of the settings for the 38 kHz echo sounder were as follows:

Transceiver-2 menu (38 kHz)	
Transducer depth	5 - 8 m
Absorption coeff.	8.3 dB/km
Pulse duration	medium (1,024ms)
Bandwidth	2.43 kHz
Max power	2000 Watt
2-way beam angle	-20,6dB
gain	26,95 dB
SA correction	0.03 dB
Angle sensitivity	21.9
3 dB beamwidth	6.22° along ship 6.28 athwart ship
Alongship offset	-0.10°
Athwardship offset	-0.06°

Bottom detection menu Minimum level -50 dB

Fishing gear

The vessel has one small four-panel 'Åkrahamn' pelagic trawl (Figure A1), one MultPelt 624 trawl (Figure A2) and one 'Gisund super bottom trawl' (Figure A3). All trawls were used during the survey. The smallest pelagic trawl has 10-12 m vertical opening under normal operation, whereas the MultPelt 624 trawl has 30-40 m opening.

The bottom trawl has a 31 m headline and a 47 m footrope fitted with a 12" rubber bobbins gear. The codend has 20 mm meshes and has an inner net with 10 mm mesh size. The vertical opening is about 5.5 m. The distance between the wing tips is about 18 m during towing. The sweeps are 40 m long. The trawl doors are 'Thyborøen' combi, 8 m² and weigh 2000 kg. The door spreading is about 45 m when using restraining rope. Trawling was conducted for species identification only and no restraining rope was therefore used during the survey.

The SCANMAR system was used during all trawl hauls. This equipment consists of sensors, a hydrophone, a receiver, a display unit and a battery charger. Communication between sensors and ship is based on acoustic transmission. The doors are fitted with sensors to provide information on their inter-distance and angle, while a height sensor is fitted on the bottom trawl to measure the trawl opening and provide information on clearance and bottom contact.

The pelagic trawls are equipped with a trawl eye that provides information about the trawl opening and the distance of the footrope to the bottom. A pressure sensor is used to show the depth on the headline.

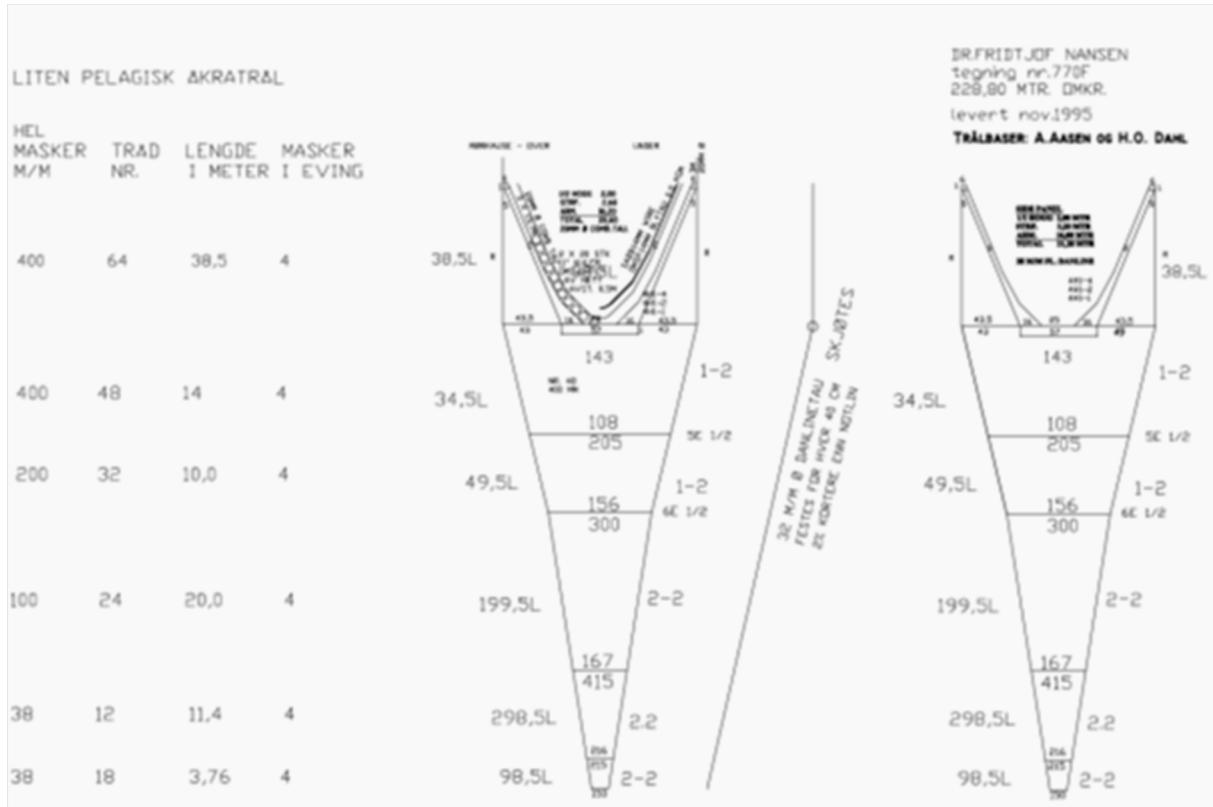


Figure A1. Schematic drawing of the small pelagic Åkratrawl.

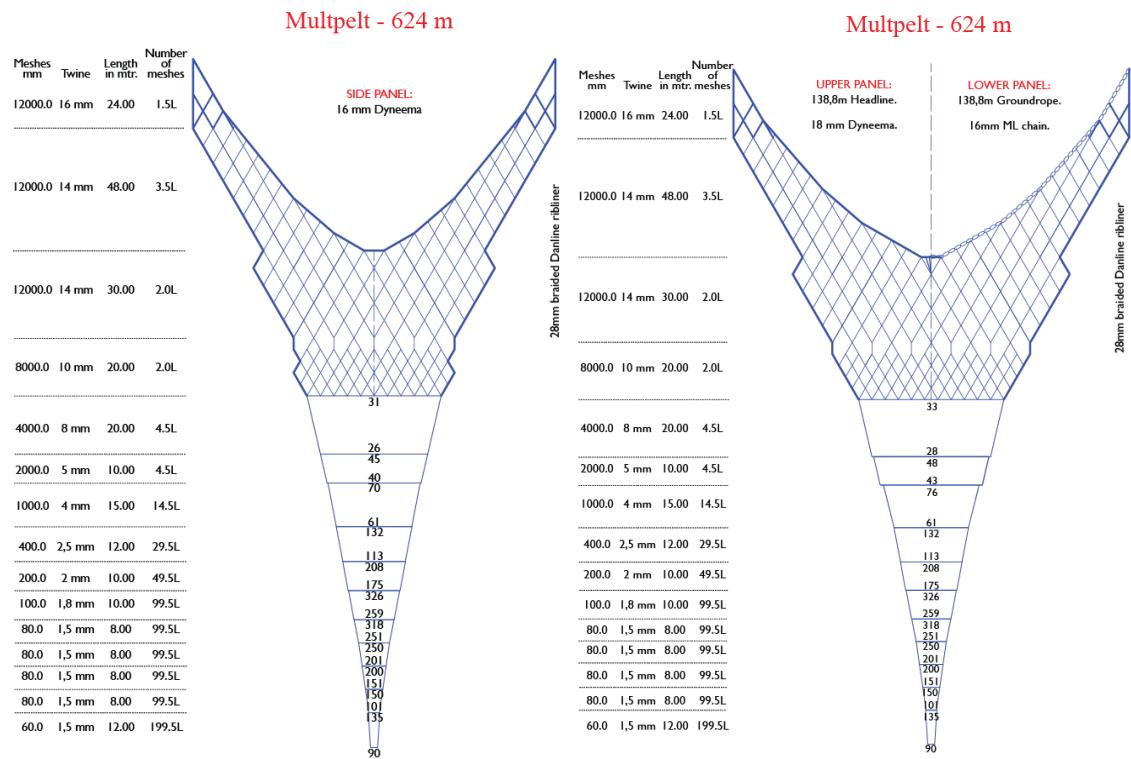


Figure A2. Schematic drawing of the MultPelt 624.

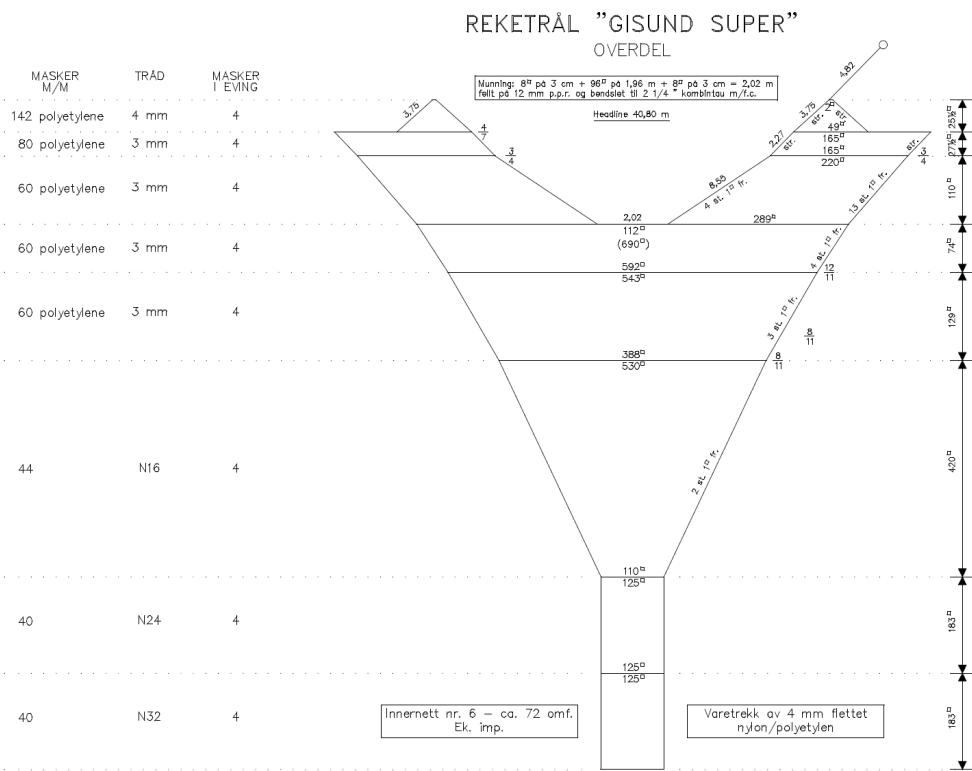
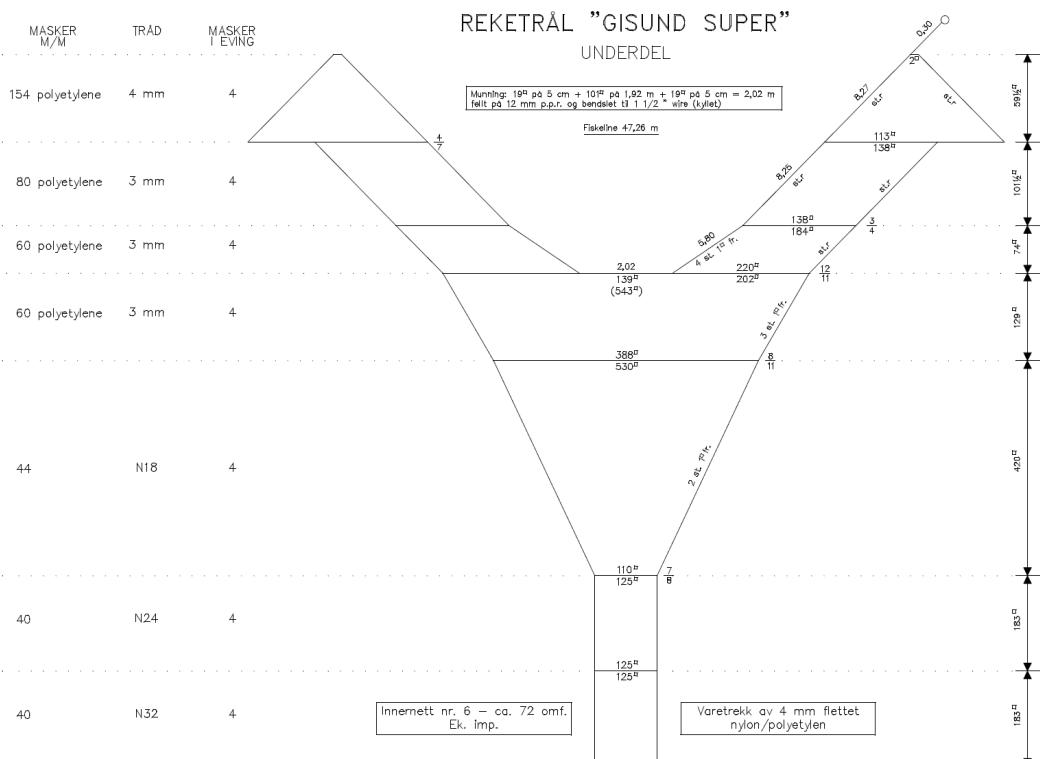


Figure A3. Schematic drawing of the Super Gisund bottom trawl.

ANNEX II.

LIST OF PRIORITY SPECIES

Main Groups	Main Families	Typical Species	70 spp		16 spp	10 spp	15 spp
			Length and weight	Sex and maturation	Stomach	Genetics	Contaminants
Demersal fish	Mullidae	<i>Upeneus taenopterus</i>	x	x	x	x	x
		<i>Upeneus baissi</i>	x	x			
		<i>Upeneus sulfureus</i>	x	x	x	x	x
		<i>Upeneus vittatus</i>	x	x			
	Nemipteridae	<i>Nemipterus japonicus</i>	x	x			
		<i>Nemipterus bipunctatus</i>	x	x			
	Leiognathidae	<i>Leiognathus equulus</i>	x	x			
	Synodontidae	<i>Saurida undosquamis</i>	x	x	x		x
	Merlucciidae	<i>Merluccius paradoxus</i>	x	x	x		
	Scianidae	<i>Johnius dussumieri</i>	x	x	x		
		<i>Johnius amblycephalus</i>	x	x			
		<i>Otolithes ruber</i>	x	x	x		x
	Sparidae	<i>Pagellus natalensis</i>	x	x			
		<i>Polysteganus coeruleopunctatus</i>	x	x			
	*Ariidae (catfish)	<i>Arius dussumieri</i>	x	x			
	Polynemidae (Threadfins)	<i>Polydactylus sextarius</i>	x	x			
		<i>Poly nemus sextarius</i>	x	x			
	Haemulidae	<i>Pomadasys maculatus</i>	x	x	x		x
		<i>Pomadasys kaakan</i>	x	x	x		
Pelagic fish	Clupeidae	<i>Amblygaster sirm</i>	x	x			
		<i>Dussumieria acuta</i>	x	x			
		<i>Herklotischthys quadrimaculat.</i>	x	x			
		<i>Hilsa kelee</i>	x	x	x	x	x
		<i>Pellona ditchela</i>	x	x	x		
		<i>Sardinella gibbosa</i>	x	x			
		<i>Sardinella albelia</i>	x	x	x	x	x
		<i>Sardinops ocellatus</i>	x	x			
	Engraulidae	<i>Engrasicholina punctifer</i>	x	x			
		<i>Thryssa vitrirostris</i>	x	x			
		<i>Thryssa setirostris</i>	x	x	x		x
	Carangidae	<i>Alectis indicus</i>	x	x			
		<i>Alepes djedaba</i>	x	x			
		<i>Caranoides malabaricus</i>	x	x			
		<i>Decapterus tabl</i>	x	x			
		<i>Decapterus macarellus</i>	x	x			
		<i>Decapterus macrosoma</i>	x	x			
		<i>Decapterus kurroides</i>	x	x			
	Scombridae	<i>Decapterus russelli</i>	x	x	x	x	x
		<i>Selar crumenophthalmus</i>	x	x	x	x	x
		<i>Rastrelliger kanagurta</i>	x	x			
		<i>Scomber japonicus</i>	x	x			
	Trichiuridae	<i>Scomberomorus commerson</i>	x	x			
		<i>Scomberomorus plurilineatus</i>	x	x			
		<i>Trichiurus lepturus</i>	x	x	x		x
		<i>Sphyraena forsteri</i>	x	x			
		<i>Sphyraena jello</i>	x	x			
Shrimps	Penaeidae	<i>Sphyraena putnamiae</i>	x	x			
		<i>Sphyraena qenie</i>	x	x			
		<i>Sphyraena barracuda</i>	x	x	x		x
		<i>Penaeus monodon</i>	x	x			
		<i>Penaeus indicus</i>	x	x			x
		<i>Penaeus japonicus</i>	x	x			
		<i>Penaeus semisulcatus</i>	x	x			
	Aristeidae	<i>Penaeus latisulcatus</i>	x	x			
		<i>Penaeopsis balssi</i>	x	x			
		<i>Metapenaeus monoceros</i>	x	x			
	Pandalidae	<i>Metapenaeus stebbingi</i>	x	x			
		<i>Aristeus antennatus</i>	x	x			
		<i>Aristaeomorpha foliacea</i>	x	x			
		<i>Plesiopenaeus edwardsianus</i>	x	x			
Lobsters	Palurinidae	<i>Plesionika martia</i>	x	x			
Crayfish	Nephropidae	<i>Haliporoides triarthrus</i>	x	x			x
Slipper lobster	Scyllaridae	<i>Ibacus novemdentatus</i>	x	x			
Crabs	Portunidae	<i>Scyllarides elisabethae</i>	x	x			
		<i>Portunus sanguinolentus</i>	x	x		x	
	Portunidae	<i>Portunus pelagicus</i>	x	x		x	
	Geryonidae	<i>Chaceon macpherson</i>	x	x		x	

ANNEX III.

RECORDS OF FISHING STATIONS

R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 1	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 5				
DATE :07/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°16.71 Lon E 40°29.90	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°5.77 Lon E 40°3.56				
TIME :19:52:05	20:12:56	20.9 (min)	Purpose : 3	Purpose : 3					
LOG : 1537.08	1538.12	1.0	Region : 7300	Region : 7300					
FDEPTH: 561	573		Gear cond.: 0	Gear cond.: 0					
BDEPTH: 561	573		Validity : 0	Validity : 0					
Towing dir: 0°	Wire out : 1345 m	Speed : 3.0 kn	Towing dir: 0°	Wire out : 530 m	Speed : 3.0 kn				
Sorted : 24	Total catch: 117.44	Catch/hour: 337.95	Sorted : 38	Total catch: 38.47	Catch/hour: 78.51				
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP		
Nettastoma parviceps	164.09	1940	48.55	8	Polysteganus coeruleopunctatus	23.88	29	30.41	36
Centrophorus granulosus	107.57	58	31.83	1	Rhinobatos holochrysnchus	19.51	16	34.85	43
S H R I M P S	14.71	331	4.35	6	Heterodontus ramaleira	11.43	2	14.56	34
Chlorophthalmus agassizii	12.85	83	3.80	10	Satyridichthys adenai	3.82	8	4.86	40
Ventrifossa mystax	9.95	299	2.94	9	Urotrygonchus archionema	3.01	10	3.83	44
Torpedo sp.	9.78	3	2.90		Urotrygonchus duvaucelii	2.71	51	3.46	55
Malacocephalus laevis	5.29	63	1.57	2	Tyliurus spinosissimus	2.63	8	3.35	38
DIODONTIDAE	4.03	3	1.19	7	Etelis carbunculus	2.41	2	3.07	37
Haliporoides triarthrus	3.91	92	1.16	11	Saurida undosquamis	2.12	12	2.70	35
Aristeus antennatus	2.10	135	0.62	12	Pistularia petimba	1.61	2	2.05	42
Rostroraja alba	1.55	3	0.46	4	Ibacus novemdentatus	1.49	8	1.89	50
Polymetme corythaeola	1.41	63	0.42	3	Harpiosquilla harpax	1.22	2	1.56	48
Cruriraja parcomaculata	0.72	3	0.21	5	Champsodon sp.	0.73	118	0.94	53
Total	337.95		100.00	Monocentris japonica	0.53	4	0.68	54	
				Decapterus russelli	0.42	4	0.54	47	
				Rexea prometheoides	0.33	16	0.42	39	
				SALPS	0.27	12	0.34		
				Penaeus marginatus	0.14	2	0.18	49	
				Lepidotrigla alcocki	0.08	8	0.13	45	
				Citharichthys sp.	0.06	8	0.08	56	
				Sepia hieronim	0.03	8	0.04	52	
				Pontinus nigerrimus	0.03	2	0.04	46	
				Antigonion capros	0.02	2	0.02	41	
				Total	78.51		100.00		
R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 2	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 6				
DATE :08/04/18	GEAR TYPE: PT NO: 4	POSITION:Lat S 10°4.12 Lon E 40°19.08	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°2.36 Lon E 40°6.70				
TIME :00:09:30	00:39:22	29.9 (min)	Purpose : 3	Purpose : 3					
LOG : 1562.37	1563.76	1.4	Region : 7300	Region : 7300					
FDEPTH: 0	0		Gear cond.: 0	Gear cond.: 0					
BDEPTH: 851	771		Validity : 0	Validity : 0					
Towing dir: 0°	Wire out : 150 m	Speed : 2.8 kn	Towing dir: 0°	Wire out : 970 m	Speed : 3.3 kn				
Sorted : 12	Total catch: 49.70	Catch/hour: 99.83	Sorted : 13	Total catch: 140.66	Catch/hour: 281.32				
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 6			
Rastrelliger kanagurta	52.43	6	52.52	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°2.36 Lon E 40°6.70			
Cubiceps pauciradiatus	37.50	0	37.57	TIME :09:29:46	09:59:46	30.0 (min)			
MYCTOPHIDAE	6.41	1270	6.42	LOG : 1611.40	1613.03	1.6			
Onychotethus sp.	3.50	123	3.50	FDEPTH: 366	348				
Total	99.83		100.00	BDEPTH: 366	348				
				Towing dir: 0°	Wire out : 970 m	Speed : 3.3 kn			
				Sorted : 13	Total catch: 140.66	Catch/hour: 281.32			
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 6			
Leiognathus berbis	28.11	0	79.71	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°2.36 Lon E 40°6.70			
Decapterus kurroides	4.81	280	13.64	TIME :09:29:46	09:59:46	30.0 (min)			
Polymetme corythaeola	1.72	477	4.88	LOG : 1611.40	1613.03	1.6			
Cheilopogon nigricans	0.27	6	0.78	Purpose : 3	Purpose : 3				
Urotrygonchus duvaucelii	0.20	16	0.55	Region : 7300	Region : 7300				
Lestrolepis intermedia	0.16	2	0.44	Gear cond.: 0	Gear cond.: 0				
Total	35.27		100.00	Validity : 0	Validity : 0				
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 6			
Leiognathus berbis	28.11	0	79.71	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°2.36 Lon E 40°6.70			
Decapterus kurroides	4.81	280	13.64	TIME :09:29:46	09:59:46	30.0 (min)			
Polymetme corythaeola	1.72	477	4.88	LOG : 1611.40	1613.03	1.6			
Cheilopogon nigricans	0.27	6	0.78	Purpose : 3	Purpose : 3				
Urotrygonchus duvaucelii	0.20	16	0.55	Region : 7300	Region : 7300				
Lestrolepis intermedia	0.16	2	0.44	Gear cond.: 0	Gear cond.: 0				
Total	35.27		100.00	Validity : 0	Validity : 0				
R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 3	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 6				
DATE :08/04/18	GEAR TYPE: PT NO: 4	POSITION:Lat S 10°11.25	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°2.36				
TIME :02:13:26	02:44:08	30.7 (min)	Purpose : 3	Purpose : 3					
LOG : 1574.47	1575.97	1.5	Region : 7300	Region : 7300					
FDEPTH: 0	0		Gear cond.: 0	Gear cond.: 0					
BDEPTH: 393	279		Validity : 0	Validity : 0					
Towing dir: 0°	Wire out : 150 m	Speed : 2.9 kn	Towing dir: 0°	Wire out : 970 m	Speed : 3.3 kn				
Sorted : 0	Total catch: 18.04	Catch/hour: 35.27	Sorted : 13	Total catch: 74.11	Catch/hour: 281.32				
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 6			
Leiognathus berbis	28.11	0	79.71	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°2.36 Lon E 40°6.70			
Decapterus kurroides	4.81	280	13.64	TIME :09:29:46	09:59:46	30.0 (min)			
Polymetme corythaeola	1.72	477	4.88	LOG : 1611.40	1613.03	1.6			
Cheilopogon nigricans	0.27	6	0.78	Purpose : 3	Purpose : 3				
Urotrygonchus duvaucelii	0.20	16	0.55	Region : 7300	Region : 7300				
Lestrolepis intermedia	0.16	2	0.44	Gear cond.: 0	Gear cond.: 0				
Total	35.27		100.00	Validity : 0	Validity : 0				
R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 4	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 7				
DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 10°6.58	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 9°54.04				
TIME :05:18:02	05:32:25	14.4 (min)	Purpose : 3	Purpose : 3					
LOG : 1590.20	1591.03	0.8	Region : 7300	Region : 7300					
FDEPTH: 88	100		Gear cond.: 0	Gear cond.: 0					
BDEPTH: 88	100		Validity : 0	Validity : 0					
Towing dir: 0°	Wire out : 280 m	Speed : 3.4 kn	Towing dir: 0°	Wire out : 700 m	Speed : 3.0 kn				
Sorted : 66	Total catch: 69.17	Catch/hour: 288.43	Sorted : 37	Total catch: 74.11	Catch/hour: 163.55				
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 7			
Aluterus monoceros	108.49	42	37.62	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 9°54.04			
Leiognathus elongatus**	100.57	0	34.87	TIME :04:13:33	15:07:45	27.2 (min)			
Gnathanodon speciosus	25.85	4	8.96	LOG : 1650.54	1651.89	1.4			
Carangoides dinema	12.93	4	4.48	Purpose : 3	Purpose : 3				
Carangoides sp.	11.51	4	3.99	Region : 7300	Region : 7300				
Carangoides chrysophrys	9.67	25	3.35	Gear cond.: 0	Gear cond.: 0				
Decapterus macrostoma	9.55	1539	3.31	Validity : 0	Validity : 0				
Total	288.42		100.00	Towing dir: 0°	Wire out : 700 m	Speed : 3.0 kn			
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 7			
Aluterus monoceros	108.49	42	37.62	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 9°54.04			
Leiognathus elongatus**	100.57	0	34.87	TIME :04:13:33	15:07:45	27.2 (min)			
Gnathanodon speciosus	25.85	4	8.96	LOG : 1650.54	1651.89	1.4			
Carangoides dinema	12.93	4	4.48	Purpose : 3	Purpose : 3				
Carangoides sp.	11.51	4	3.99	Region : 7300	Region : 7300				
Carangoides chrysophrys	9.67	25	3.35	Gear cond.: 0	Gear cond.: 0				
Decapterus macrostoma	9.55	1539	3.31	Validity : 0	Validity : 0				
Seriola rivoliana	3.84	0	1.33	Towing dir: 0°	Wire out : 700 m	Speed : 3.0 kn			
Urotrygonchus duvaucelii	2.59	271	0.90	Sorted : 37	Total catch: 74.11	Catch/hour: 163.55			
Nettastoma parviceps	1.60	17	0.52						
Carangoides sp.	1.25	142	0.43						
Malacocephalus laevis	0.42	17	0.14						
Nemipterus sp.	0.18	0	0.06						
Coelorinchus denticulatus	0.04	4	0.02						
Cynoglossus lida	0.04	4	0.01						
Waste General	0.00	4	0.00						
Total	288.42		100.00						
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 7			
Aluterus monoceros	108.49	42	37.62	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 9°54.04			
Leiognathus elongatus**	100.57	0	34.87	TIME :04:13:33	15:07:45	27.2 (min)			
Gnathanodon speciosus	25.85	4	8.96	LOG : 1650.54	1651.89	1.4			
Carangoides dinema	12.93	4	4.48	Purpose : 3	Purpose : 3				
Carangoides sp.	11.51	4	3.99	Region : 7300	Region : 7300				
Carangoides chrysophrys	9.67	25	3.35	Gear cond.: 0	Gear cond.: 0				
Decapterus macrostoma	9.55	1539	3.31	Validity : 0	Validity : 0				
Seriola rivoliana	3.84	0	1.33	Towing dir: 0°	Wire out : 700 m	Speed : 3.0 kn			
Urotrygonchus duvaucelii	2.59	271	0.90	Sorted : 37	Total catch: 74.11	Catch/hour: 163.55			
Nettastoma parviceps	1.60	17	0.52						
Carangoides sp.	1.25	142	0.43						
Malacocephalus laevis	0.42	17	0.14						
Nemipterus sp.	0.18	0	0.06						
Coelorinchus denticulatus	0.04	4	0.02						
Cynoglossus lida	0.04	4	0.01						
Waste General	0.00	4	0.00						
Total	288.42		100.00						
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 7			
Aluterus monoceros	108.49	42	37.62	DATE :08/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat S 9°54.04			
Leiognathus elongatus**	100.57	0	34.87	TIME :04:13:33	15:07:45	27.2 (min)			
Gnathanodon speciosus	25.85	4	8.96	LOG : 1650.54	1651.89	1.4			
Carangoides dinema	12.93	4	4.48	Purpose : 3	Purpose : 3				
Carangoides sp.	11.51	4	3.99	Region : 7300	Region : 7300				
Carangoides chrysophrys	9.67	25	3.35	Gear cond.: 0	Gear cond.: 0				
Decapterus macrostoma	9.55	1539	3.31	Validity : 0	Validity : 0				
Seriola rivoliana	3.84	0	1.33	Towing dir: 0°	Wire out : 700 m	Speed : 3.0 kn			
Urotrygonchus duvaucelii	2.59	271	0.90	Sorted : 37	Total catch: 74.11	Catch/hour: 163.55			
Nettastoma parviceps	1.60	17	0.52						
Carangoides sp.	1.25	142	0.43						
Malacocephalus laevis	0.42	17	0.14						
Nemipterus sp.	0.18	0	0.06						
Coelorinchus denticulatus	0.04	4	0.02						
Cynoglossus lida	0.04	4	0.01						
Waste General	0.00	4	0.00						
Total	288.42		100.00						
SPECIES	CATCH/HOUR weight numbers	% OF TOT. C	SAMP	R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION: 7			
Aluterus monoceros	108.49	42	37.62	DATE :					

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 8
 DATE :08/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 8°40.38
 start stop duration Lon E 39°49.14
 TIME :21:12:20 21:39:18 26.8 (min) Purpose : 3
 LOG : 1686.69 1687.99 1.3 Region : 7300
 FDEPTH: 358 378 Gear cond.: 0
 BDEPTH: 358 378 Validity : 0
 Towing dir: 0° Wire out : 850 m Speed : 2.9 kn
 Sorted : 12 Total catch: 52.02 Catch/hour: 116.46

SPECIES

	CATCH/HOUR	% OF TOT.	C	SAMP
weight numbers				
Centrophorus molluccensis	17.51	4	15.03	118
Polytmus coryphaeola	14.40	513	12.36	106
Saurida undosquamis	9.40	40	8.07	105
Polyipnus indicus	8.57	0	7.36	
Bythaelurus lutarius	8.26	83	7.09	133
Chaunax sp.	7.50	22	6.44	132
Malacocephalus laevis	6.27	67	5.38	111
Chlorophthalmus agassizii	5.66	99	4.86	126
Himantura sp.	5.17	22	4.44	115
Neobrychites kenyaeensis	5.13	22	4.40	116
Aristea antennatus	4.75	237	4.08	125
Argentina euchus	3.83	83	3.29	119
Polytmixia berndti	3.09	11	2.65	129
Rexea prometheoides	2.91	29	2.50	113
Etmopterus sentosus	2.69	25	2.31	117
Linuparus somniosus	2.15	7	1.55	
Urotrygon duvauceillii	1.99	13	1.71	130
PENAEIDAE	1.23	11	1.06	127
Polytmixia sp.	1.12	11	0.96	128
Neobrychites cf. somaliaensis	0.92	13	0.79	121
Coelorinchus trunovii	0.90	38	0.77	112
Zenion hololepis	0.76	83	0.65	122
Physiculus natalensis	0.76	7	0.65	120
Atrobucca nibe	0.54	7	0.46	131
Beryx splendens	0.31	7	0.27	124
Tydemania navigatoris	0.31	7	0.27	110
Lestrolepis intermedia	0.16	7	0.13	123
Antigonius rubescens	0.11	7	0.10	108
Rastrelliger kanagurta	0.07	13	0.06	109
Total	116.46		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 9
 DATE :09/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 8°40.72
 start stop duration Lon E 39°25.93
 TIME :12:18:12 12:47:27 29.3 (min) Purpose : 3
 LOG : 1791.63 1793.34 1.7 Region : 7300
 FDEPTH: 22 41 Gear cond.: 0
 BDEPTH: 22 41 Validity : 0
 Towing dir: 0° Wire out : 140 m Speed : 3.5 kn
 Sorted : 0 Total catch: 750.28 Catch/hour: 1538.52

SPECIES

	CATCH/HOUR	% OF TOT.	C	SAMP
weight numbers				
Encrasicholina heteroloba	1396.75	498845	90.79	147
Scomberomorus commerson	36.71	21	2.39	134
Himantura jenkinsii	30.64	4	1.99	149
Balistes capriscus	29.20	23	1.90	
Urotrygon duvauceillii	16.67	416	1.08	146
Leiognathus berbis	12.24	504	0.80	135
OSTRACIIDAE	5.58	12	0.36	144
Arlothrinus stellatus	5.09	2	0.33	143
Carangoides malabaricus	1.90	4	0.12	136
Cruriraja paracomulata	1.49	23	0.10	145
Fistularia petimba	0.88	23	0.06	111
Sardinella gibosa	0.44	66	0.03	140
Leiognathus elongatus**	0.14	66	0.03	139
Decapterus kurroides	0.29	23	0.02	138
Canthigaster sp.	0.17	23	0.01	148
Torquigenes hypselogenion	0.04	2	0.00	142
Total	1538.52		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 10
 DATE :09/04/18 GEAR TYPE: PT NO: 1 POSITION:Lat S 8°40.48
 start stop duration Lon E 39°32.70
 TIME :17:37:21 18:08:20 31.0 (min) Purpose : 3
 LOG : 1805.65 1807.19 1.5 Region : 7300
 FDEPTH: 65 55 Gear cond.: 0
 BDEPTH: 526 397 Validity : 0
 Towing dir: 0° Wire out : 155 m Speed : 3.0 kn
 Sorted : 2 Total catch: 12.58 Catch/hour: 24.35

SPECIES

	CATCH/HOUR	% OF TOT.	C	SAMP
weight numbers				
MYCTOPHIDAE	23.14	1936	95.02	
Muraenesox bagio	0.70	12	2.87	
Leptoccephalus	0.40	35	1.65	
Lestrolepis intermedia	0.11	23	0.47	
Total	24.35		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 11
 DATE :09/04/18 GEAR TYPE: PT NO: 4 POSITION:Lat S 8°24.71
 start stop duration Lon E 39°42.60
 TIME :23:58:36 00:28:59 30.4 (min) Purpose : 3
 LOG : 1847.69 1849.03 1.3 Region : 7300
 FDEPTH: 779 663 Gear cond.: 0
 BDEPTH: 0 0 Validity : 0
 Towing dir: 0° Wire out : 145 m Speed : 2.7 kn
 Sorted : 3 Total catch: 2.79 Catch/hour: 5.50

SPECIES

	CATCH/HOUR	% OF TOT.	C	SAMP
weight numbers				
MYCTOPHIDAE	2.39	2393	43.45	
Urotrygon duvauceillii	1.78	219	32.32	151
Leptoccephalus	0.81	296	14.79	
Seriola rivoliana	0.43	2	7.79	152
Scomber japonicus	0.04	4	0.65	155
Lestrolepis intermedia	0.03	6	0.61	150
Decapterus tabi	0.01	2	0.22	154
Zanclus cornutus	0.01	2	0.11	153
Hemiramphus sp.	0.00	2	0.07	156
Total	5.50		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 12
 DATE :10/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 8°10.88
 start stop duration Lon E 39°33.10
 TIME :05:51:05 06:17:30 26.4 (min) Purpose : 3
 LOG : 1880.53 1882.20 1.7 Region : 7300
 FDEPTH: 27 35 Gear cond.: 0
 BDEPTH: 27 35 Validity : 0
 Towing dir: 0° Wire out : 130 m Speed : 3.8 kn
 Sorted : 0 Total catch: 43.04 Catch/hour: 97.71

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 13
 DATE :10/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 8°10.00
 start stop duration Lon E 39°42.92
 TIME :08:50:03 09:20:45 30.7 (min) Purpose : 3
 LOG : 1896.58 1898.16 1.6 Region : 7300
 FDEPTH: 349 310 Gear cond.: 0
 BDEPTH: 349 310 Validity : 0
 Towing dir: 0° Wire out : 850 m Speed : 3.1 kn
 Sorted : 26 Total catch: 226.77 Catch/hour: 443.20

Total 97.71 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 14
 DATE :10:16:12 22:46:58 30.8 (min) Purpose : 3
 LOG : 1990.95 1992.41 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 443.20 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 15
 DATE :11:04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 7°32.72
 start stop duration Lon E 39°58.67
 TIME :22:16:12 22:45:58 30.8 (min) Purpose : 3
 LOG : 1991.58 1992.41 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 16
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1992.41 1993.29 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 17
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1993.29 1994.17 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 18
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1994.17 1995.05 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 19
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1995.05 1996.93 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 20
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1996.93 1997.81 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 21
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1997.81 1998.69 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 22
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1998.69 1999.57 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 23
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 1999.57 2000.45 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 24
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2000.45 2001.33 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 25
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2001.33 2002.21 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 26
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2002.21 2003.09 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 27
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2003.09 2003.97 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 28
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2003.97 2004.85 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 29
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2004.85 2005.73 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 30
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2005.73 2006.61 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 31
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2006.61 2007.49 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch/hour: 359.11

Total 359.11 100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 32
 DATE :11:23:12 22:52:00 30.8 (min) Purpose : 3
 LOG : 2007.49 2008.37 1.5 Region : 7300
 FDEPTH: 486 468 Gear cond.: 0
 BDEPTH: 486 468 Validity : 0
 Towing dir: 0° Wire out : 1150 m Speed : 2.8 kn
 Sorted : 26 Total catch: 184.17 Catch

R/V Dr. Fridtjof Nansen		SURVEY:2018404		STATION: 19						
DATE :11/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat	S 7°33.75	POSITION:Lat	S 7°21.64	Lon	E 39°44.03			
TIME : 08:52:03	09:22:28	30.4 (min)	Purpose : 3	start	stop	duration				
LOG : 2025.56	2027.35	1.8	Region : 7300	2076.38	2077.68	1.3				
FDEPTH: 46	33		Gear cond.: 0	FDEPTH:	55	65	Purpose : 3			
BDEPTH: 46	33		Validity : 0	BDEPTH:	193	233	Region : 7300			
Towing dir: 0°	Wire out : 145 m		Speed : 3.5 kn	Towing dir:	0°	Wire out : 155 m	Gear cond.: 0			
Sorted : 33	Total catch: 638.24		Catch/hour: 1258.86	Sorted :	1	Total catch: 9.34	Validity : 0			
SPECIES										
		CATCH/HOUR	% OF TOT. C	SAMP	SPECIES					
Upeneus taeniopterus		weight numbers			MYCTOPHIDAE					
Secutor insidiator	457.14	17862	36.31	247	Trichirurus lepturus	17.03	14005			
Carangoides malabaricus	291.21	17862	23.13	248	Leiognathus elongatus**	2.94	5			
Gazza minuta	148.57	3258	11.80	239	Lestrolepis intermedia	0.86	109			
Dussumieriacauta	55.45	1525	4.40	240	Decapterus tabl	0.86	109			
Chirocentrusdorab	27.51	85	2.19	255	Apogon sp.	0.71	10			
Leiognathus elongatus**	25.40	550	2.02	236	Dipterygonotus balteatus	0.22	8			
Leiognathus berbis	25.40	381	2.02	241	Bregmacerosp. sp.	0.22	13			
Gerres filamentosus	24.55	0	1.95	237	LOLIGINIDAE	0.22	218			
Thenus orientalis	17.78	85	1.41		SEPIIDAE	0.22	23			
Decapterus russelli	14.39	126	1.14	252	J E L L Y F I S H	0.17	23			
Terapontheraps	11.85	254	0.94	243	CARANGIDAE	0.00	66			
Diagramma centurio	8.05	2	0.64	256	TETRAODONTIDAE	0.00	43			
Saurida undosquamis	6.96	57	0.55		Total	23.67	100.00			
Laevitrygoninermis	5.52	2								
Nemipterusjaponicus	5.37	85	0.43	242	R/V Dr. Fridtjof Nansen	SURVEY:2018404				
Ariusaficanus	5.21	2	0.41	257	STATION: 20	STATION: 20				
Leiognathus berbis	5.08	37968	0.40		DATE :11/04/18	GEAR TYPE: BT NO: 1				
Rastrelliger kanagurta	5.08	211	0.40	251	POSITION:Lat	S 7°19.51	POSITION:Lat	S 7°19.51		
Sphyraenaflavicauda	5.08	126	0.40	244	LOG : 2086.36	2087.94	1.6	LOG : 2086.36	2087.94	1.6
Upeneusvittatus	4.87	41	0.39	245	FDEPTH: 347	345	Gear cond.: 0	FDEPTH: 347	345	Gear cond.: 0
Apogon sp.	4.23	170	0.34	253	BDEPTH: 347	345	Validity : 0	BDEPTH: 347	345	Validity : 0
Upeneusmoluccensis	2.58	85	0.21	246	Towing dir: 0°	Wire out : 810 m	Speed : 3.1 kn	Towing dir: 0°	Wire out : 810 m	Speed : 3.1 kn
Loligo duvauceli	2.12	254	0.17	254	Sorted : 32	Total catch: 105.06	Catch/hour: 202.88	Sorted : 32	Total catch: 105.06	Catch/hour: 202.88
Total		1258.86		100.00						
SPECIES								SPECIES		
		CATCH/HOUR	% OF TOT. C	SAMP				CATCH/HOUR	% OF TOT. C	SAMP
Upeneustaeiophterus	22.72	1636	14.31	274	MYCTOPHIDAE	86.22	11755	42.50	332	
Carangoides malabaricus	21.36	915	13.45	281	Dalatias licha	20.12	2	9.92		
Saurida undosquamis	18.56	839	11.69	271	Peristedion weberi	17.67	282	8.71	326	
Epinephelusareolatus	11.82	4	7.44	260	Penaeopsis balssi	14.60	133	7.20		
Dussumieriacauta	11.42	501	7.19	270	Polymixia berndti	13.71	218	6.76	324	
Drepane sp.	7.37	67	4.64	261	Argentinita euchnus	9.54	234	4.70	318	
Scomberomorus commerson	5.54	9	3.49	265	PROCESSIDAE	7.20	114	3.55		
Rastrelliger kanagurta	4.65	67	2.93	279	Decapterus macrosoma	7.20	106	3.55	331	
Loligo duvauceli	3.83	154	2.41	268	Saurida undosquamis	5.12	27	2.52	317	
Penaeussemisulcatus	3.38	85	2.13		Cynoglossus capensis	4.31	42	2.12	335	
Apogon sp.	3.20	610	2.02	277	Bembrops sp.	3.11	21	1.53	327	
Abalistesstellatus	2.96	2	1.86	269	Lestrolepis intermedia	2.97	183	1.46	323	
Secutorinsidiator	2.87	398	1.81	276	Bembrops platyrhynchus	1.81	35	0.89	341	
Laevitrygoninermis	2.54	51	1.60	267	Polymixia sp.	1.77	446	0.87		
Chirocentrusdorab	2.23	9	1.40	266	Citharoides macrolepis	1.70	56	0.84	336	
Pistularia petimba	1.74	67	1.10	264	Tydemania navigatoris	0.99	35	0.49	328	
Metapenaeusmonoceros	1.19	118	0.75	275	Otolithes ruber	0.78	8	0.38	319	
Leiognathus equulus	1.07	24	0.67	272	Macrorhamphosus sp.	0.71	8	0.35	338	
Pteroisrusselli	0.76	9	0.48	282	Malacocephalus laevis	0.71	8	0.35	320	
Upeneusvittatus	0.75	9	0.47	278	Chauanax sp.	0.67	14	0.33		
Sphyraenabotus	0.73	18	0.46	263	Colorinches sp.	0.57	35	0.28	321	
Nemipterusjaponicus	0.63	9	0.39	259	Neobrythites kenyensis	0.49	56	0.24	337	
Mene maculata	0.53	9	0.34		Spatelloides sp.	0.35	21	0.17	329	
Sphyraenabarracuda	0.39	9	0.25	262	Decapterus russelli	0.28	8	0.14	330	
Leiognathus splendens	0.31	9	0.20	273	Arnoglossus sp.	0.14	29	0.07	334	
Total		158.82		100.00	Loligo duvauceli	0.14	8	0.07	333	
R/V Dr. Fridtjof Nansen								R/V Dr. Fridtjof Nansen		
		CATCH/HOUR	% OF TOT. C	SAMP				CATCH/HOUR	% OF TOT. C	SAMP
Upeneustaeiophterus	22.72	1636	14.31	274	Trichirurus lepturus	453.36	828	61.61	342	
Carangoides malabaricus	21.36	915	13.45	281	Polymixia berndti	35.85	4671	4.87	343	
Saurida undosquamis	18.56	839	11.69	271	Saurida undosquamis	29.16	85	3.96	355	
Epinephelusareolatus	11.82	4	7.44	260	Uranoscopus archionema	19.12	101	2.60	357	
Dussumieriacauta	11.42	501	7.19	270	Cynoglossus capensis	18.78	327	2.55	351	
Drepane sp.	7.37	67	4.64	261	LOLIGENIDAE	17.99	149	2.44	349	
Scomberomorus commerson	5.54	9	3.49	265	Peristedion weberi	17.79	36	2.42	363	
Rastrelligerkanagurta	4.65	67	2.93	279	Squalus megalops	17.71	24	2.41	364	
Loligo duvauceli	3.83	154	2.41	268	Pteryxylonates boyvinus	17.35	2	2.26		
Penaeussemisulcatus	3.38	85	2.13		Loligo duvauceli	16.26	168	2.21	373	
Apogon sp.	3.20	610	2.02	277	Rexea prometheoides	14.28	164	1.94	366	
Abalistesstellatus	2.96	2	1.86	269	Citharoides macrolepis	10.06	93	1.37	344	
Secutorinsidiator	2.87	398	1.81	276	Scorpaena scrofa	8.74	16	1.19	345	
Laevitrygoninermis	2.54	51	1.60	267	Spatelloides sp.	8.08	277	1.10	350	
Chirocentrusdorab	2.23	9	1.40	266	Holohalaelurus punctatus	7.94	4	1.08		
Pistularia petimba	1.74	67	1.10	264	Projassus parkeri	4.57	293	0.62		
Metapenaeusmonoceros	1.19	118	0.75	275	Parabembrops sp.	4.33	20	0.59	376	
Leiognathus equulus	1.07	24	0.67	272	Unidentified fish	3.55	0	0.48	378	
Epinephelusmalabaricus	38.26	10	5.29		Tydemania navigatoris	3.17	145	0.43	348	
Decapterusmacrocosma	36.28	843	5.11	291	Chauanax sp.	2.89	8	0.39	370	
Ariusvenosus	27.09	6	3.74	284	Sepia hieronims	2.73	52	0.37	353	
Nemipterusmetopias	27.00	309	3.73	303	Raja sp.	2.30	4	0.31	352	
Sauridaundosquamis	22.08	703	3.05	304	Neopinnula orientalis	2.26	184	0.31	361	
Pomadasysmaculatus	21.94	168	3.03	296	CREPIDULIDAE (CALYPTERAIDAE)	2.22	16	0.30	371	
Leiognathusequulus	19.69	252	2.72	289	Atelolepis natalensis	2.15	4	0.29	374	
Psettodeserumei	12.66	4	1.75	286	Peristedion weberi	1.91	52	0.26	356	
Pomadasyskaakan	7.57	2	1.05	285	Macrourididae	1.92	12	0.26	367	
Loligo duvauceli	6.01	589	0.83	310	Halaelurus sp.	1.56	16	0.21		
Unidentified crustacean	5.06	591	0.70		Bythaelurus lutarius	1.56	16	0.21	362	
Dussumieriacauta	3.94	112	0.54	307	Neoscopelus sp.	1.31	8	0.18	377	
Apogon sp.	3.93	843	0.54	301	Lepidotethis sp.	1.29	12	0.17	372	
Leiognathusleuciscus**	3.37	168	0.47	299	Pliotrema warreni	1.21	8	0.16	358	
Gazza minuta	2.75	112	0.38	300	Branchiostegus doliatius	1.09	8	0.15	346	
Oxyurichthyspetersii	2.25	168	0.31	315	Antigonion capros	0.74	16	0.10	369	
Thryssa vitrirostris	2.25	112	0.31	306	Total	735.80	100.00			
Plastic	2.00	0	0.28							
Polyneussextarius**	1.69	56	0.23	302	Sepiopsetta zanzibarensis**	0.43	8	0.06	354	
Sepiaprashadi	1.12	28	0.16	311	Poecilopsetta zanzibarensis**	0.08	4	0.01	360	
Terapontheraps	0.56	28	0.08	298	Macracocephalus laevis	0.08	4	0.01		
Trypauchenmicrocephalus	0.33	55	0.05	316	Macrourhamphosus gracilis	0.08	8	0.01	359	
Octopuscyanus	0.25	28	0.03							
Citharoidesmacrolepis	0.20	28	0.03	313						
Laeopsnatalensis	0.17	28	0.02	314						
Citharichthyssp.	0.14	28	0.02	312						

SPECIES	CATCH/HOUR		% OF TOT.	C	SAMP
	weight	numbers			
<i>Trichiurus lepturus</i>	185.45	295	45.95		381
<i>Polysteganus coeruleopunctatus</i>	34.79	27	8.62		395
<i>Saurida undosquamis</i>	30.95	102	7.67		379
<i>Apogon</i> sp.	29.93	1287	7.42		
<i>Uranoscopus archionema</i>	18.18	84	4.50		
<i>Satyrichthys adeni</i>	17.34	33	4.30		392
<i>Lophiodes mutilus</i>	16.48	8	4.08		
<i>Rexea prometheoides</i>	14.70	135	3.64		382
<i>Citharus</i> sp.	14.70	178	3.64		387
<i>Sepia hieronis</i>	9.29	195	2.30		383
<i>Squalus megalops</i>	8.28	8	2.05		
<i>Spratelloides</i> sp.	5.66	271	1.40		384
<i>Loligo duvauceillii</i>	4.55	43	1.13		398
<i>Linuparitus somniosus</i>	3.38	8	0.84		
UNIDENTIFIED FISH					
<i>Parabembas</i> sp.	2.77	297	0.69		
<i>Cynoglossus lida</i>	2.36	18	0.59		393
A B S					
<i>Muraenocelphalus laevis</i>	2.11	43	0.52		389
<i>Polymixia bernardi</i>	0.74	18	0.18		
<i>Legidopterigla multispinosa</i>	0.59	16	0.15		
<i>Tydemania navigatoris</i>	0.23	8	0.06		396
<i>Uroconger</i> sp.	0.23	18	0.06		
<i>Parapenaeus fissurus</i>	0.08	8	0.02		
Total		402.62			100.00

R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION:	23
DATE :12/04/18	GEAR TYPE: BT NO: 1	POSITION:	Lat S 6°32.26 Lon E 39°36.83
start stop duration			
TIME :22:17:28 22:48:21	30.9 (min)	Purpose :	3
LOG : 2262.88	2264.30	Region :	7300
FDEPTH: 404	410	Gear cond.:	0
BDEPTH: 404	410	Validity :	0
Towing dir: 0°	Wire out : 940 m	Speed :	2.7 kn
Sorted : 27	Total catch: 82.66	Catch/hour:	160.61

SPECIES		CATCH/HOUR	% OF TOT.	C	SAMP
		weight numbers			
<i>Argentina euchus</i>		10.80 909	25.41		409
CARIIDA		15.93 1168	9.92		
<i>Linuparatus sonniosus</i>		13.35 .27	8.31	0	
MYCTOPHIDAE		10.78 2217	6.71		
<i>Octopus sp.</i>		9.85 16	6.13	434	
<i>Polyipnops indicus</i>		9.69 1426	6.03	421	
<i>Otolithes ruber</i>		8.23 6	5.13	425	
<i>Squalus megalops</i>		5.75 6	3.58	399	
<i>Malacocephalus laevis</i>		4.95 85	3.08	410	
Peristedion weberi		4.95 155	3.08	426	
<i>Polymetme corythaeola</i>		4.53 350	2.82	411	
<i>Tydemania navigatoris</i>		4.04 97	2.51	415	
<i>Lepidopus caudatus</i>		3.34 6	2.08	438	
<i>Cocciella sp.</i>		2.58 16	1.61	430	
<i>Raja sp.</i>		2.53 12	1.57	405	
<i>Neopinnula orientalis</i>		2.31 6	1.44	428	
<i>Cynoglossus tenuis</i>		1.72 37	1.07	418	
<i>Serranichthys adami</i>		1.67 6	1.04		
<i>Saurida undosquamis</i>		1.35 12	0.84	424	
<i>Physiculus natalensis</i>		1.26 12	0.79	404	
<i>Bathycongrus wallacei</i>		1.18 16	0.74		
<i>Uranoscopus archionema</i>		1.13 6	0.70	437	
<i>Sepia sp.</i>		0.97 91	0.60	401	
<i>Uroteuthis duvauzelii</i>		0.91 6	0.57		
<i>Caelorinchus trunovi</i>		0.91 6	0.57	412	
<i>Satyrichthys investigatoris</i>		0.75 12	0.47	435	
<i>Sepia hieronis</i>		0.65 12	0.40	419	
<i>Neobrythites kenyensis</i>		0.51 21	0.32	408	
<i>Polymixia bernardi</i>		0.48 6	0.30	414	
<i>Bythaelurus lutarius</i>		0.48 6	0.30	429	
<i>Spratelloides sp.</i>		0.48 323	0.30	427	
<i>Citharoides macrolepis</i>		0.43 12	0.27		
<i>Acropomae japonicum</i>		0.43 6	0.27	422	
<i>Epidotrigla sp.</i>		0.38 6	0.23	431	
<i>Etmopterus sentosus</i>		0.32 6	0.20	420	
<i>Apogon sp.</i>		0.21 6	0.13	423	
<i>Xenolepidichthys dagleishi</i>		0.17 12	0.11	403	
<i>Macrorhamphosus scolopax</i>		0.16 12	0.10	413	
<i>Chaulax sp.</i>		0.16 12	0.10	433	
Unidentified fish		0.16 12	0.10	402	
<i>Stomias boa boa</i>		0.11 12	0.07	436	

R/V Dr. Fridtjof Nansen	SURVEY:2018404	STATION:	24
DATE :13/04/18	GEAR TYPE: BT NO: 1	POSITION:Lat	S 6°39.57
	start stop duration	Lon	E 39°19.37
TIME : 05:45:24 06:12:00	26.6 (min)	Purpose	: 3
LOG : 2294.40	2295.62	Region	: 7300
BDEPTH: 67	60	Gear cond.	: 0
BDEPTH: 67	60	Validity	: 0
Towing dir: 0°	Wire out : 170 m	Speed	: 2.8 kn

SPECIES		CATCH/HOUR	% OF	TOT.	C	SAMP.
	weight	numbers				
<i>Sepia vermiculata</i>	28.69	72	32.79		459	
<i>Acroteriobatus zanzibarensis</i>	16.06	9	18.36		465	
<i>Urothechites duvaucelii</i>	12.59	519	14.39		439	
<i>Tetrosomus concatenatus</i>	5.41	14	6.19		450	
<i>Pterois miles</i>	4.24	9	4.85		440	
<i>Gymnocranius griseus</i>	3.92	29	4.49		456	
<i>Saurida undosquamis</i>	3.38	52	3.87		441	
<i>Lactoria diaphana</i>	3.29	14	3.76		449	
<i>Nemipterus zyprion</i>	3.20	59	3.66		442	
<i>Decapterus macromosoma</i>	2.21	7	2.53		451	
<i>Paratriacanthodes retrospinis</i>	0.86	29	0.98		454	
<i>Lagocephalus guentheri</i>	0.63	18	0.72		447	
<i>Alabistes stellatus</i>	0.54	2	0.62		446	
<i>Caranxoides macrobaricus</i>	0.54	2	0.62		457	
<i>Diplodus guttatus</i>	0.51	2	0.58		445	
<i>Parupeneus heptacanthus</i>	0.34	2	0.39		443	
<i>Peristedion weberi</i>	0.23	9	0.26		448	
<i>Nemipterus bipunctatus</i>	0.22	2	0.25		444	
<i>Leiognathus elongatus**</i>	0.18	108	0.21		453	
<i>Fistularia petimba</i>	0.14	2	0.15		452	
<i>Polytmus corythaeola</i>	0.09	9	0.10		462	
<i>Bothus pantherinus</i>	0.09	2	0.10		460	
<i>Apogonidae - juvenile</i>	0.05	23	0.05		458	
<i>Polyipnus indicus</i>	0.05	7	0.05		461	
UNIDENTIFIED FISH	0.03	5	0.03			
MYCTOPHIDAE	0.02	5	0.02		455	

R/V Dr. Fridtjof Nansen SURVEY:2018A04 SURVEY:2018A04 STATION: 25
 DATE :13/04/18 GEAR TYPE: PT NO: 5 POSITION:Lat S 6°34.01
 start stop duration Lon E 39°15.30
 TIME : 08:00:08 08:27:33 27.4 (min) Purpose : 3
 LOG : 2308.18 2310.30 2.1 Region : 7300
 FDDEPTH: 0 30 Gear cond.: 0
 BDEPTH: 58 64 Validity : 0
 Towing dir: 0° Wire.out : 200 m Speed : 4.7 kn
 Towing dir: 0° Wire.out : 200 m Speed : 4.7 kn

SPECIES	CATCH/HOUR weight	% OF TOT. numbers	C	SA
<i>Engrasicholina punctifer</i>	211.51	88144	100.00	4

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Total           211.51          100.00

R/V Dr. Fridtjof Nansen      SURVEY:2018404      STATION: 26
DATE :14/04/18      GEAR TYPE:BT NO: 1      POSITION:Lat S 6°30'.61
                           start    stop   duration
                           TIME      : 12:28:40 12:56:25 27.8 (min)
                           LOG       : 2346.66 2348.98 1.5
                           FDEPTH:   60      54
                           BDEPTH:  60      54
                           Towing dir: 0° Wire out : 170 m Speed : 3.3 kn
                           Sorted : 38 Total catch: 37.70 Catch/hour: 81.52

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SPECIES	CATCH/HOUR	% OF TOT. C	SA
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	Weight	Number	Mean	Range
Saurida undosquamis	17.56	223	21.54	4
Gymnocranius griseus	12.50	28	15.33	4
Upeneus bensasi	9.38	763	11.51	4
Abalistes stellatus	7.87	11	9.65	4
Ostracion cubicus	5.77	6	7.08	4
Carcharhinus limbatus	4.71	2	5.78	4
Tetrosomus concatenatus	4.43	17	5.44	4
Sepia prashadi	3.59	2	4.40	4
Loligo duvauceli	3.16	50	3.87	4
Acroteriobatus zanzibarensis	2.85	2	3.50	4
Arothron hispidus	2.29	2	2.81	4
Nemipterus japonicus	1.66	24	2.04	4
Paramonacanthus sp.	0.99	108	1.22	4
Psettodes erumei	0.86	4	1.06	4
Lagocephalus gentheri	0.76	48	0.93	4
Fistularia petimba	0.56	19	0.69	4
Sphoeroides annulatus	0.53	2	0.64	4
Choerodon cyanogenys	0.45	9	0.56	4
Sepia hieronimi	0.26	4	0.32	4
Epinephelus areolatus	0.24	2	0.29	4
UNIDENTIFIED FISH	0.22	13	0.27	4
Lethrinus microdon	0.19	2	0.24	4
Leiognathus equinus	0.13	2	0.16	4
Upeneus moluccensis	0.13	4	0.16	4
Oxycheilinus bimaculatus	0.13	15	0.16	4
Thennus orientalis	0.09	4	0.11	4
Caranxoides malabaricus	0.09	2	0.11	4
Teixeirichthys jordani	0.06	4	0.08	4
Apogon sp.	0.04	9	0.05	4
Apogon quadripectus**	0.01	6	0.01	4
Invertebrate	0.00	4	0.00	4
SEA URCHINS	0.00	2	0.00	4
Total	101.56	1000	100.06	4

R/V Dr. Fridtjof Nansen	SURVEY:20180404	STATION: 27
DATE : 14/04/18	GEAR TYPE: BT NO: 1	POSITION: Lat S 6°19.39' Lon E 39°13.31'
start	stop	duration
TIME : 14:27:23	14:56:01	28.6 (min)
LOG : 2359.86	2361.70	1.8
DEPTH : 33	37	
BDEPTH : 33	37	
Towing dir: : 0°	Wire out : 130 m	Speed : 3.9 kn
Sorted : 23	Total catch: 85.02	Catch/hour: 178.11

SPECIES		CATCH/HOUR weight numbers	% OF TOT.	C	SA
Rachycentron canadum		69.76	31	39.17	
Caranxoides malabaricus		26.19	505	14.70	5
Saurida undosquamis		18.12	247	10.17	5
Lagocephalus sceleratus		9.76	233	5.48	4
Sepla trigonina		5.89	63	3.31	5
Leiognathus leuciscus**		5.57	419	3.13	4
Upeneus densirostris		5.56	557	2.95	5
Theraps ocellatus		4.94	23	2.78	5
Sepla verniculata		4.48	8	2.52	5
Cyclichthys sp.		4.34	8	2.43	4
Caranxoides ferdau		4.02	86	2.26	5
Nemipterus sp.		3.39	31	1.91	5
Gazza minuta		3.10	40	1.74	5
Pterois miles		2.16	8	1.21	5
Parupeneus sp.		2.16	31	1.21	5
Decapterus macrosoma		1.55	54	0.87	5
Caranx tillæ		1.24	23	0.69	5
Gerres filamentosus		1.24	40	0.69	4
Amanses scopas		0.78	69	0.44	5
Caranxoides armatus		0.78	54	0.44	5
CARANGIDAE		0.5	15	0.4	5
Decapterus russelli		0.61	8	0.34	5
Fistularia petimba		0.46	23	0.26	5
DACTYLOPTERIDAE		0.46	8	0.26	5
Uroctenus duvaucllei		0.38	15	0.21	5
Engrasiicholina heteroloba		0.27	233	0.15	5
Bothus myriaster		0.23	8	0.13	5
Teixeirichthys jordani		0.15	15	0.08	5
Plotosus lineatus		0.13	54	0.07	5
Apogon quadrifasciatus**		0.08	8	0.05	5

Citharichthys sp. 0.02 8 0.01

Total		178.11	100.00
R/V Dr. Fridtjof Nansen	SURVEY:2018A404	STATION: 28	
DATE : 14/04/18	GEAR TYPE: PT NO: 1	POSITION:Lat S 6°22.26	Lon E 39°12.44
TIME : 16:00:23	start stop	duration	Purpose : 3
LOG : 2367.01	2368.23	20.4 (min)	Gear/second : 1300
FDEPTH : 25	30		Validity : 0
BDEPTH : 41	39		Speed : 3.6 kn
Towing dir: 0°	Wire out :	100 m	Catch/hour: 48.87
Sorted : 15	Total catch:	16.62	

SPECIES		CATCH/HOUR	% OF TOT.	C	SA
	weight	numbers			
<i>Spratelloides gracilis</i>	17.37	10859	35.55	5	
<i>Engraulis encrasicolus heterolebba</i>	13.58	7644	27.79	5	
<i>Decapterus macrosoma</i>	10.55	1479	22.20	5	
<i>Amblygaster sirm</i>	5.17	676	10.59	5	
<i>Rastrelliger kanagurta</i>	0.72	68	1.47	5	
UNIDENTIFIED FISH	0.53	185	1.08	5	
<i>Sphyraena obtusata</i>	0.32	15	0.66	5	
<i>Loligo duvaucelii</i>	0.21	79	0.44	5	
<i>Samaris cristatus**</i>	0.08	15	0.16	5	
<i>Apogon</i> sp.	0.03	9	0.05	5	
Total		48.87		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 29
 DATE :14/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 6°17.87
 start stop duration Lon E 39°1.25
 TIME :20:40:12 21:10:37 30.4 (min) Purpose : 3
 LOG : 2394.39 2396.26 1.9 Region : 7300
 FDEPTH: 32 30 Gear cond.: 0
 BDEPTH: 32 30 Validity : 0
 Towing dir: 0° Wire out : 130 m Speed : 3.7 kn
 Sorted : 22 Total catch: 95.54 Catch/hour: 188.38

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Stephanolepis auratus	36.34	6974	19.29	562
Saurida undosquamis	23.29	39	12.36	536
Upeneus bensasi	21.06	1782	11.18	539
Pseudorhombus elevatus ***	14.71	1530	7.81	548
SOFT SPONGES	11.22	0	5.96	
Lagocephalus sceleratus	10.61	244	5.63	
Paramonacanthus pusillus	10.45	300	5.55	561
Decapterus russelli	6.49	158	3.44	538
Thenus orientalis	5.70	87	3.02	
Paracallionymus costatus	4.93	244	2.62	570
Lutjanus fulvus	4.67	578	2.48	557
Lethrinus microdon	3.94	47	2.09	546
Aristeus carinatus	3.81	237	2.02	560
Teixeirichthys jordani	3.73	260	1.98	545
Nemipterus bipunctatus	3.57	39	1.89	541
SYNOIDAE	3.17	32	1.69	565
Rastrelliger kanagurta	2.94	39	1.56	537
Unidentified crustacean	2.54	103	1.35	
Torquigenere hypselogenion	2.44	79	1.30	552
Bothus myriaster	1.97	71	1.05	547
Lethrinus nebulosus	1.81	32	0.96	544
Upeneus taeniopterus	1.81	79	0.96	540
Sepia hieronis	1.34	16	0.71	554
Herklotischthys quadrimaculatus	0.95	16	0.50	563
Gymnocranius griseus	0.79	16	0.42	558
Apogon sp.	0.63	71	0.33	556
Choerodon gymnotensis	0.63	16	0.33	559
Loligo duvauceli	0.55	16	0.29	555
Sphyraena guachancho	0.47	8	0.25	
Gerres cyrena	0.47	8	0.25	566
Choerodon sp.	0.45	24	0.24	568
Plotosus lineatus	0.39	260	0.21	549
Samaris cristatus**	0.24	16	0.13	553
UNIDENTIFIED FISH	0.10	32	0.05	569
Hoplichthys acanthophorus	0.08	8	0.04	564
Chauanax sp.	0.08	8	0.04	
Total	188.38		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 30
 DATE :15/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 6°7.46
 start stop duration Lon E 38°59.46
 TIME :00:53:42 01:17:47 24.1 (min) Purpose : 3
 LOG : 2415.69 2417.07 1.4 Region : 7300
 FDEPTH: 35 37 Gear cond.: 0
 BDEPTH: 35 37 Validity : 0
 Towing dir: 0° Wire out : 140 m Speed : 3.4 kn
 Sorted : 0 Total catch: 89.04 Catch/hour: 221.76

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Upeneus bensasi	42.14	3078	19.00	579
Lethrinus variegatus	36.01	598	16.24	571
Saurida undosquamis	23.51	224	10.78	586
Scolopsis bimaculata	20.47	1494	9.23	578
Teixeirichthys jordani	18.93	1494	8.54	
Parapercis sp.	17.78	329	8.02	
Thenus orientalis	10.01	45	4.51	583
Lagocephalus sceleratus	9.71	164	4.38	575
Stephanolepis auratus	5.98	0	2.70	587
Pterois miles	5.23	30	2.36	585
Bothus sp.	4.87	45	2.20	590
Paramonacanthus pusillus	4.78	269	2.16	588
Aristeus carinatus	4.48	254	2.02	580
Abalistes stellatus	3.74	15	1.68	573
Parupeneus macronemus	2.99	60	1.35	572
Plotosus lineatus	2.39	194	1.08	581
Sepia latimanus	1.94	75	0.88	582
Siganus surtor	1.79	45	0.81	584
Nemipterus bipunctatus	1.34	15	0.61	593
Rastrelliger kanagurta	0.90	15	0.40	589
Gymnocranius griseus	0.45	60	0.20	574
Dussumieriaca	0.45	15	0.20	595
Choerodon gymnotensis	0.45	15	0.20	577
Synodus sp.	0.45	30	0.20	576
Lethrinus sp.	0.38	15	0.17	592
BOTHIDAE	0.17	30	0.08	591
Total	221.76		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 31
 DATE :15/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°56.01
 start stop duration Lon E 39°8.95
 TIME :05:37:48 06:08:57 31.1 (min) Purpose : 3
 LOG : 2444.31 2445.92 1.6 Region : 7300
 FDEPTH: 45 50 Gear cond.: 0
 BDEPTH: 45 50 Validity : 0
 Towing dir: 0° Wire out : 150 m Speed : 3.1 kn
 Sorted : 15 Total catch: 119.04 Catch/hour: 229.29

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Himantura jenkinsii	59.22	4	25.83	617
Upeneus taeniopterus	27.83	1285	12.14	613
Ariopsis	23.58	0	10.28	624
Ariommabeanii	23.46	6	10.23	625
Leiognathus leuciscus**	14.99	917	6.54	612
Upeneus sulphureus	14.68	466	4.40	603
Saurida undosquamis	13.16	299	5.74	597
Carangoides malabaricus	10.56	206	4.60	610
Stolephorus waitei	10.40	4956	4.54	623
Arotrochus stellatus	5.82	2	2.54	621
Scomberomorus commerson	3.74	8	1.63	616
Fsettoidea erumei	3.66	2	1.60	620
Abalistes stellatus	3.22	15	1.40	600
Upeneus moluccensis	2.29	214	1.00	601
Fistularia petimba	1.83	46	0.80	598
Thenus orientalis	1.27	8	0.55	604
Penaeus semisulcatus	1.23	31	0.54	
Dussumieriaca acuta ***	1.23	62	0.54	619
Gerres filamentosus	1.23	15	0.54	607
Nemipterus metopias	1.08	8	0.47	618
Epinephelus malabaricus	1.08	15	0.47	608
Sepia vermiculata	1.07	5	0.47	599
Paramonacanthus pusillus	0.77	100	0.14	615
Apogon quadrifasciatus**	0.46	15	0.20	605
Mene maculata	0.46	15	0.20	611
Terapon thompsoni	0.44	8	0.19	622
Sphyraena putnamiae	0.31	8	0.13	609
Rastrelliger kanagurta	0.15	39	0.07	614
Plotosus lineatus	0.08	23	0.03	606
Total	229.29		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 32
 DATE :15/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°44.79
 start stop duration Lon E 39°5.89
 TIME :10:24:15 10:53:43 29.5 (min) Purpose : 3
 LOG : 2474.63 2476.06 1.4 Region : 7300
 FDEPTH: 71 74 Gear cond.: 0
 BDEPTH: 71 74 Validity : 0
 Towing dir: 0° Wire out : 200 m Speed : 2.9 kn
 Sorted : 57 Total catch: 338.98 Catch/hour: 690.15

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Decapterus macrosoma	465.48	9799	67.45	629
Carangoides malabaricus	72.54	1362	10.51	635
Upeneus moluccensis	49.78	1568	7.21	636
Rastrelliger kanagurta	21.19	43	3.07	643
Leiognathus berbis	19.04	1325	2.76	633
Abalistes stellatus	12.54	49	1.82	630
Upeneus taeniopterus	8.67	0	1.26	642
Nemipterus japonicus	8.43	120	1.22	627
Saurida undosquamis	7.72	240	1.12	628
CARCHARHINIDAE	7.57	2	1.10	641
Apogon sp.	7.47	12	1.08	638
Loligo duvauceli	3.14	96	0.45	639
Stolephorus indicus	3.01	96	0.44	634
Sphyraena putnamiae	3.57	12	0.23	
Dussumieriaca acuta	1.26	24	0.18	632
Paramonacanthus sp.	0.49	71	0.07	
Fistularia petimba	0.24	12	0.04	631
Total	690.15		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 33
 DATE :15/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°26.80
 start stop duration Lon E 39°5.28
 TIME :15:10:15 15:45:40 30.5 (min) Purpose : 3
 LOG : 2511.29 2512.84 1.6 Region : 7300
 FDEPTH: 65 51 Gear cond.: 0
 BDEPTH: 65 51 Validity : 0
 Towing dir: 0° Wire out : 189 m Speed : 3.1 kn
 Sorted : 5 Total catch: 113.97 Catch/hour: 224.12

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Leiognathus elongatus**	48.14	10637	21.48	661
Epinephelus coloides	40.55	2	18.09	648
Scomberomorus commerson	39.45	16	17.60	645
Lutjanus sanganeus	19.15	4	8.56	662
Lutjanus gibbus	17.45	2	7.87	663
Gymnocranius griseus	9.99	65	4.46	660
Sphyraena putnamiae	7.87	2	3.51	669
Loligo duvauceli	7.83	865	3.49	664
Aluterus monoceros	6.92	4	3.09	665
Nemipterus zyron	6.06	104	2.70	655
Decapterus kurroides	2.44	47	1.09	663
Stolephorus indicus	2.37	94	1.06	647
Saurida tumbil	2.16	6	0.97	
Thenus orientalis	2.01	8	0.90	651
Lagocephalus sceleratus	1.85	2	0.82	658
Diodon hystrix	1.85	2	0.82	659
Parupeneus heptacanthus	1.61	12	0.72	657
Trachinops myops	1.53	47	0.68	667
Lactoria cornuta	1.26	4	0.56	649
Saurida undosquamis	1.10	6	0.49	652
Tetrosomus concatenatus	1.06	2	0.47	644
Sepia vermiculata	0.55	2	0.25	646
Rogadius asper	0.37	12	0.17	650
Apogon quadrifasciatus**	0.24	12	0.11	666
Saurida gracilis	0.04	8	0.02	653
Fistularia petimba	0.04	2	0.02	656
Total	224.12		100.00	

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 34
 DATE :15/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°27.60
 start stop duration Lon E 39°11.44
 TIME :17:44:19 18:14:57 30.6 (min) Purpose : 3
 LOG : 2523.17 2524.81 1.6 Region : 7300
 FDEPTH: 356 340 Gear cond.: 0
 BDEPTH: 356 340 Validity : 0
 Towing dir: 0° Wire out : 845 m Speed : 3.2 kn
 Sorted : 21 Total catch: 70.13 Catch/hour: 137.33

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Centrophorus granulosus	19.9	6	14.12	682
Pristiophorus nancyae	11.51	82	8.98	690
Dipturus stenorhynchus	9.20	94	6.70	676
Etmopterus sentosus	4.39	49	3.19	688
Cocciella sp.**	4.33	16	3.15	711
Sepia hieronis	4.17	55	3.04	703
Palinurus sp.	3.51	27	2.55	714
Loligo duvauceli	3.41	27	2.48	704
Cubiceps whiteleggi	3.23	76	2.35	670
Dysomma anguillare	3.17	27	2.31	674
Polyipnus indicus	3.11	1073	2.27	677
Neobythites kenyensis	2.36	16	1.72	683
Paralutjanus hamrur	2.35	12	1.71	678
Physiculus natalensis	2.35	16	1.71	698
Bythaelurus uritus	2.25	49	1.64	687
Parapercis weberi	1.98	82	1.44	675
Locinoides megalurus	1.80	6</		

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 35
 DATE :16/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°15.57
 start stop duration Lon E 39°12.19
 TIME :19:07:57 19:29:15 21.3 (min) Purpose : 3
 LOG : 2701.19 2702.33 1.1 Region : 7300
 FDEPTH: 390 390 Gear cond.: 0
 BDEPTH: 390 390 Validity : 0
 Towing dir: 0° Wire out : 970 m Speed : 3.2 kn
 Sorted : 13 Total catch: 48.71 Catch/hour: 137.22

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Saurida undosquamis	36.37	189	26.51	716
MYCTOPHIDAE	20.42	5104	14.88	
Chlorophthalmus agassizii	9.60	242	7.00	
Centrophorus granulosus	9.46	3	6.90	
Parapeneus sp.	8.11	1245	5.91	
Peristedion weberi	6.76	39	4.93	
Chaunax sp.	5.81	39	4.24	738
Ornithotheuthis volatilis	4.29	54	3.13	748
Tydemania navigatoris	4.26	6	3.10	740
Metanephrops mozambicus	4.12	48	3.01	
Atrobucca nibe	2.59	3	1.89	735
Solenocera agoensis	2.57	54	1.87	
Malacocephalus laevis	2.37	20	1.72	742
Polymixia berndti	2.37	28	1.72	741
Aristeus antennatus	2.16	68	1.58	
Polyipnus indicus	1.76	527	1.28	
Rutjanus pretiosus	1.69	14	1.23	
Velodona costata	1.57	6	1.15	753
Phosichthys argenteus	1.49	101	1.08	
Spratelloides sp.	1.15	39	0.84	
Fristiophorus nancyae	1.13	11	0.82	
Satyrichthys investigatoris	1.08	48	0.79	732
Thyrsoptoides marleyi	1.01	14	0.74	718
Stomias boa boa	0.81	155	0.59	752
Lestrolepis intermedia	0.64	34	0.47	725
Cynoglossus marleyi	0.44	6	0.32	726
Gymnoscopelus sp.	0.41	130	0.30	751
Bathycongrus wallacei	0.41	3	0.30	
Macrorhamphosus scolopax	0.41	54	0.30	717
Malacocephalus laevis	0.27	87	0.20	
Poecilopsetta zanzibarensis	0.21	20	0.15	727
Coelorinchus denticulatus	0.20	6	0.15	730
Etmopterus sentosus	0.17	3	0.12	
Cynoglossus lida	0.17	6	0.12	728
Eridacnis radcliffei	0.14	3	0.10	
Bythaelurus lutarius	0.14	3	0.10	
Bembrops platyrhynchus	0.14	6	0.10	755
Physosteges sp.	0.14	6	0.10	739
Lepidotrigla fawleri	0.14	6	0.10	747
Paratriacanthodes retrospinis	0.11	0	0.08	754
Parabathynomus natalensis	0.10	6	0.07	749
Lacops nigromaculatus	0.03	6	0.02	734
Total	137.22	100.00		

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 36
 DATE :16/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°3.46
 start stop duration Lon E 39°16.74

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Argentina euchus	14.88	293	14.15	769
Polymetme coryphaeola	9.77	319	9.29	
Parapenaeus investigatoris	9.24	800	8.79	
Parascolopsis sp.	8.30	266	7.90	
Peristedion weberi	7.28	115	6.93	770
Saurida undosquamis	5.95	27	5.66	759
Neobythites cf somaliaensis	4.97	5	4.73	
Gymnoscopelus sp.	4.88	1017	4.65	787
MYCTOPHIDAE	4.44	1292	4.22	766
Sepia sp.	3.86	5	3.68	
Malacocephalus laevis	3.73	124	3.55	771
Lestrolepis intermedia	3.64	160	3.46	768
Scorpaenidae	3.20	41	3.04	767
Chaunax sp.	2.84	41	2.70	788
Tydemania navigatoris	2.80	27	2.66	
Polyipnus indicus	2.71	848	2.58	786
Sepia megalops	2.45	4	2.33	
BAJIDAe	2.04	41	1.94	
Uroconger lepturus	1.60	9	1.52	762
Sepia hieronis	1.15	14	1.10	
Spratelloides sp.	0.76	32	0.72	
Tydemania navigatoris	0.72	27	0.69	
Rhinochimaera atlantica	0.71	18	0.68	
Lophiodes sp.	0.49	9	0.46	
Stomias boa boa	0.49	98	0.46	763
Cynoglossus lida	0.40	14	0.38	
Polymixia berndti	0.31	5	0.30	773
Avocettina sp.	0.31	14	0.30	765
Neobythites analis	0.31	18	0.30	764
Plastic	0.30	14	0.28	
Unidentified demersal fish	0.18	14	0.17	
Unidentified demersal fish	0.15	14	0.14	789
Macrorhamphosus scolopax	0.13	14	0.13	
Dussumieria acuta	0.13	5	0.13	761
Total		105.13		100.00

R/V Dr. Fridtjof Nansen SURVEY:2018404 STATION: 37
 DATE :17/04/18 GEAR TYPE: BT NO: 1 POSITION:Lat S 5°51.88
 start stop duration Lon E 39°26.30

SPECIES	CATCH/HOUR	% OF TOT.	C	SAMP
	weight	numbers		
Rexea prometheoides	91.64	1453	56.53	790
Zeus capensis	15.69	6	9.68	801
Peristedion weberi	15.17	57	9.36	800
Squalus megalops	8.69	4	5.36	
Plastic	7.58	10	4.68	
Saurida undosquamis	7.48	27	4.61	793
Dendrocerus russelli	3.26	10	2.01	792
Loligo duvaucellii	2.69	20	1.66	798
Apogon sp.	2.11	191	1.30	794
Lepidotrigla sp.	1.47	31	0.91	803
Antigonion rubescens	1.42	64	0.88	797
Citharoides macrolepis	1.26	10	0.78	796
Polymixia berndti	1.21	53	0.75	795
Spratelloides sp.	1.00	53	0.62	791
Atelopus sp.	0.42	6	0.26	805
Sepia hieronis	0.32	6	0.19	799
Synagrops sp.	0.26	6	0.16	804
Eridacnis radcliffei	0.21	6	0.13	
Bembrops sp.	0.21	6	0.13	802
Total		162.10		100.00

ANNEX IV. OVERVIEW OF BIOLOGICAL SAMPLES PER SPECIES

Length – length measurements only, Length/weight – both and length and weight, Biology – sex, maturity and stomach fullness, Stomach – frozen stomach samples, Otoliths – no. of fish where otoliths were collected, Fin clips – no. of individuals sampled for genetical analysis, Frozen – no. of individuals frozen, Preserved – no. of individuals preserved in formalin.

Species/groups	Length	Length/ Weight	Biology	Liver	Stomach	Otoliths	Fin clips	Frozen	Preserved
<i>Chaunax</i> sp.	32								
<i>Abalistes stellatus</i>	27	11					12		
<i>Acropoma japonicum</i>	1								
<i>Acroteriobatus zanzibarensis</i>	1	4							
<i>Aluterus monoceros</i>	5								
<i>Aluterus monoceros</i>		2							
<i>Amanses scopas</i>		9							
<i>Amblygaster leiogaster</i>	1								
<i>Amblygaster sirm</i>	50								
<i>Antigonia capros</i>	8								
<i>Antigonia rubescens</i>	13								
<i>Apistus carinatus</i>	31								
<i>Apogon quadrifasciatus</i>	10	10							
<i>Apogon</i> sp.	201	10							
<i>Argentina euchus</i>	427	24							
<i>Aristaeus antennatus</i>	70		13						
<i>Arius africanus</i>		3							
<i>Arius venosus</i>	2								
<i>Arnoglossus</i> sp.	9								
<i>Arothron hispidus</i>	1								
<i>Arothron stellatus</i>	1	4							
<i>Ateleopus natalensis</i>	1								
<i>Ateleopus</i> sp.	1								
<i>Atlantoraja cyclophora</i>	1								
<i>Atrobucca nibe</i>	1	1							
<i>Atrobucca nibe</i>	11								
<i>Avocettina</i> sp.	6								
<i>Bathycongrus wallacei</i>		1							
<i>Bembrops platyrhynchus</i>	5	1							
<i>Bembrops</i> sp	10								
<i>Beryx splendes</i>	2								
<i>Bothidae</i>		2							
<i>Bothus myriaster</i>	9	1							
<i>Bothus pantherinus</i>	1								
<i>Bothus</i> sp.	1	3							
<i>Branchiostegus doliatus</i>	2	2							
<i>Bythaelurus lutarius</i>	1	21							
<i>Carcharhinus limbatus</i>	1								
<i>Coelorhincus trunovi</i>	1								
<i>Caelorinchus trunovi</i>	1								
<i>Callionymus marleyi</i>	4								
<i>Canthigaster</i> sp.		1							
<i>Carangoides armatus</i>		7							
<i>Carangoides chrysophrys</i>	6								
<i>Carangoides dinema</i>	1								
<i>Carangoides ferdau</i>		11							
<i>Carangoides malabaricus</i>	121	132	31						
<i>Carangoides</i> sp.	39								
<i>Caranx tille</i>	1								
<i>Carcharhinus</i> sp.	1								
<i>Carcharinidae</i>	1								
<i>Caridea</i> sp.	30								
<i>Centrophorus granulosus</i>		1							
<i>Centrophorus moluccensis</i>	2								

Species/groups	Length	Length/	Biology	Liver	Stomach	Otoliths	Fin	Frozen	Preserved
		Weight					clips		
<i>Centrophorus</i> sp.		20							
<i>Champsodon</i> sp.	64								
<i>Chascanopsetta lugubris</i>	2								
<i>Cheilopogon nigricans</i>		3							
<i>Chirocentrus dorab</i>	3								
<i>CHLOROPHTHALMIDAE</i>	1								
<i>Chlorophthalmus agassizi</i>	12	44							
<i>Choerodon gymnogenys</i>	7								
<i>Choerodon</i> sp.	3								3
<i>Citharichthys</i> sp.	6								
<i>Citharoides macrolepis</i>	47								
<i>Citharus</i>	35								
<i>Cociella</i>	8								
<i>Coelorhincus denticulatus</i>	1								
<i>Coelorhincus turnovi</i>	5								
<i>Coelorinchus denticulatus</i>		1							
<i>Crepidulidae</i>	4								
<i>Cruriraja parcomaculata</i>		1							
<i>Cubiceps whiteleggi</i>		14							
<i>Cylichthys</i> sp.		1							
<i>Cynoglossus capensis</i>	90								
<i>Cynoglossus lida</i>	17	4							
<i>Cynoglossus marleyi</i>		1							
<i>Dactylopteridae</i>		1							
<i>Decapterus kurroides</i>	5	133						30	
<i>Decapterus macarellus</i>	19								
<i>Decapterus macrosoma</i>	174	247	37		30			120	
<i>Decapterus russelli</i>	25	57	54		13			58	
<i>Decapterus tabl</i>	1								
<i>Diagramma centurio</i>		5	5						
<i>Diaphus knappi</i>		100							
<i>Diodon hystrix</i>	5								
<i>Diodontidae</i> sp		1							
<i>Dipturus stenorhynchus</i>	1								
<i>Dussumeria acuta</i>	139	1							
<i>Dussumeria acuta</i>		8							
<i>Dysomma anguillare</i>		5							
<i>Echeneis naucrates</i>	1								
<i>Engrasicholina punctifer</i>		100							
<i>Engrasicholina heteroloba</i>		100							
<i>Epigonus robustus</i>	2								
<i>Epinephelus aerolatus</i>	3								
<i>Epinephelus coioides</i>		1							
<i>Epinephelus malabaricus</i>		2							
<i>Eridacnis radcliffei</i>	3	20							
<i>Etelis carbunculus</i>	1								
<i>Etmopterus molleri</i>		8							
<i>Etmopterus sentosus</i>	36	10							
<i>Fistularia petimba</i>	1								
<i>Fistularia petimba</i>	14	11							
<i>Gazza minuta</i>	132	5							
<i>Gerres filamentosus</i>	20	7							
<i>Gerres oyena</i>	4								
<i>Gnathanodon speciosus</i>	1								
<i>Gobius</i> spp.	16								
<i>Gymnocranius griseus</i>	6	61	15					15	
<i>Gymnoscopelus</i> sp.	50								
<i>Halaclurus lutarius</i>	5								
<i>Halaclurus</i> sp.	5								
<i>Haliporoides triarthrus</i>		9							
<i>Harpisquilla harpax</i>	1								
<i>Hemiramphus</i> sp.	1								
<i>Heptanchias perlo</i>	5								
<i>Herklotischthys quadrimaculat</i>	2								

Species/groups	Length	Length/ Weight	Biology	Liver	Stomach	Otoliths	Fin	Frozen	Preserved
							clips		
<i>Heterodontus ramalheria</i>	1								
<i>Himantura gerradi</i>	4								
<i>Himantura jenkinsii</i>		4							
<i>Holohaelurus regani</i>	1								
<i>Holohaelurus punctatus</i>	1								
<i>Hoplichthys acanthopleurus</i>	1								
<i>Ibacus novemdentatus</i>	4								
<i>Lactoria cornuta</i>		2							
<i>Lactoria</i> sp.	6								
<i>Laeops nataliensis</i>	1								
<i>Laeops nigromaculatus</i>		1							
<i>Laeops pectoralis</i>		15							
<i>Lagocephalus guentheri</i>	30								
<i>Lagocephalus inermis</i>	7								
<i>Lagocephalus sceleratus</i>	44	10							
<i>Leiognathus berbis</i>	211	23							
<i>Leiognathus elongatus</i>	254	109							
<i>Leiognathus equulus</i>	4	18	18						
<i>Leiognathus leuciscus</i>	55	56							
<i>Lepidopus caudatus</i>	1								
<i>Lepidoteuthis</i> sp.	3								
<i>Lepidotrigla alcocki</i>	4								
<i>Lepidotrigla fawleri</i>	14	17							
<i>Lepidotrigla multispinosa</i>	1								
<i>Lepidotrigla</i> sp.	7								
<i>Lestrolepis intermedia</i>	67	6							
<i>Lethrinus crocineus</i>	1								
<i>Lethrinus mahsena</i>	1								
<i>Lethrinus nebulosus</i>	6								
<i>Lethrinus</i> sp.	25								
<i>Lethrinus variegatus</i>	40								
<i>Linuparus somniosus</i>	5								
<i>Linuparus</i> sp.	2								
<i>Loligo duvaceli</i>	320	8							
<i>Lophiodes mutilus</i>	6	1							
<i>Lophius</i> sp.	1								
<i>Lutjanidae</i>	1								
<i>Lutjanus fulvus</i>	73								
<i>Lutjanus rivulatus</i>	1								
<i>Lutjanus sanguineus</i>		2					2		
<i>Macrorhamphosidae</i>	7								
<i>Macrorhamphosus gracilis</i>	2								
<i>Macrorhamphosus scolopax</i>	6	12							
<i>Malacocephalus</i>	1								
<i>Malacocephalus laevis</i>	91	21							
<i>Mene maculata</i>		2							
<i>Metanephrops mozambicus</i>									
<i>Monocentris japonica</i>	2								
<i>Myctophidae</i>	132								
<i>Narcine rierai</i>	11								
<i>Narcine</i> sp.	3								
<i>Nemipterus bipunctatus</i>	1								
<i>Nemipterus bleekeri</i>	10	4							
<i>Nemipterus japonicus</i>	15								
<i>Nemipterus metopias</i>	12	1							
<i>Nemipterus zyron</i>	26	53							
<i>Neobythites analis</i>	4								
<i>Neobythites kenyensis</i>	16	3							
<i>Neobythites somaliaensis</i>	3								
<i>Neopinnula orientalis</i>	10								
<i>Neopinnula orientalis</i>	47								
<i>Neoscopelus</i> sp.	2								
<i>Nettastoma parviceps</i>	101								
<i>Nettostoma parviceps</i>	6								

Species/groups	Length	Length/	Biology	Liver	Stomach	Otoliths	Fin	Frozen	Preserved
		Weight					clips		
<i>Octopus sp</i>	3								
<i>Octopus cyaneus</i>	1								
<i>Ornithoteuthis volatilis</i>	8								
<i>Ostraciidae</i>	6								
<i>Ostracion cubicus</i>	5	2							
<i>Otolithes ruber</i>	5	1	1		1		1		
<i>Oxycheilinus bimaculatus</i>	6								
<i>Palinurus sp.</i>	5								
<i>Parabathynomus</i>	1								
<i>Parabembras sp.</i>	7								
<i>Paracallionymus costatus</i>	31								
<i>Paramonacanthus pusillus</i>	38	31							
<i>Paramonacanthus sp.</i>	6	51							
<i>Parapenaeus sp.</i>									
<i>Paratriacanthodes retrospinis</i>	1	4							
<i>Parupeneus heptacanthus</i>	10								
<i>Parupeneus macronemus</i>	4								
<i>Parupeneus sp.</i>	3	4							
<i>Penaeidae</i>	5								
<i>Penaeus marginatus</i>	1								
<i>Penaeus semisulcatus</i>		4							
<i>Peristedion sp.</i>	9								
<i>Peristedion weberi</i>	128	25							
<i>Physiculus natalensis</i>	7								
<i>Physiculus nateliensis</i>	1								
<i>Pliotrema warreni</i>	2								
<i>Plotosus lineatus</i>	78	9							
<i>Poecilopsetta zanzibarensis</i>	5	3							
<i>Polyipnus indicus</i>	51	227							
<i>Polymetme corythaeola</i>	296	30							
<i>Polymixia berndti</i>	191								
<i>Polymixia sp.</i>	5	40							
<i>Polynemus sextarius</i>	2								
<i>Polysteganus caeruleopunctatus</i>	23	13	13		13		13		
<i>Pomadasys kaakan</i>	1								
<i>Pomadasys maculatus</i>	6								
<i>Pontinus nigerimum</i>	3								
<i>Priacanthus hamrur</i>		3							
<i>Pristiophorus nancyae</i>	15								
<i>Projassus parkeri</i>	9								
<i>Psettodes erumei</i>	3	1							
<i>Pseudobalistes flavimarginatus</i>		2							
<i>Pseudobalistes fuscus</i>	1								
<i>Pseudorhombus elevatus</i>	104								
<i>Pterios miles</i>	4								
<i>Pterois miles</i>	2	1							
<i>Pterois russelii</i>	1								
<i>Pteromylaeus bovinus</i>	1								
<i>Raja sp.</i>	5			1					
<i>Rajidae</i>	11								1
<i>Rastrelliger kanagurta</i>	6	65	19				5		
<i>Rastroraja alba</i>		1							
<i>Rexea prometheoides</i>	291								
<i>Rhinobatos</i>	8								
<i>Rhinochimaera atlantica</i>	4								
<i>Rogadius asper</i>	1								
<i>Ruvettus pretiosus</i>	3								
<i>Samaris cristatus</i>	3								
<i>Sardinella gibbosa</i>		3	3				3		
<i>Satyrichthys adeni</i>	20								
<i>Satyrichthys investigatoris</i>		7							
<i>Satyrinchus investigatoris</i>	2								
<i>Saurida gracilis</i>	5								
<i>Saurida tumbil</i>	3								

Species/groups	Length	Length/	Biology	Liver	Stomach	Otoliths	Fin	Frozen	Preserved
		Weight					clips		
<i>Saurida undosquamis</i>	102	300	31				12		
<i>Sauridia undosquamis</i>		106	36		42		57		
<i>Scolopsis bimaculata</i>	40								
<i>Scomber japonicus</i>	2								
<i>Scomberomorus commerson</i>	1	16	20				20		
<i>Scombrops boops</i>		9							
<i>Scorpaena scorfa</i>	6								
<i>Scorpaenopsis sp</i>	1								
<i>Secutor insidiator</i>	47								
<i>Sepia hieronis</i>	57								
<i>Sepia latimanus</i>	5								
<i>Sepia prashadi</i>	5								
<i>Sepia sp.</i>	19								
<i>Sepia trygonina</i>		9							
<i>Sepia vermiculata</i>		35							
<i>Seriola rivoliana</i>	2								
<i>Seriolina nigrofasciata</i>	1								
Shrimps		20							
<i>Siganus sutor</i>	3								
<i>Sinodus sp</i>	2								
<i>Solenocera agoensis</i>	8								
<i>Sparidae</i>	18						15		
<i>Sphyraena barracuda</i>	1								
<i>Sphyraena flavicauda</i>	3								
<i>Sphyraena forsteri</i>	1								
<i>Sphyraena jello</i>	10								
<i>Sphyraena obtusa</i>	2	50							
<i>Sphyraena obtusta</i>	2	1							
<i>Sphyraena putnamae</i>	2	22							
<i>Spratelloides</i>	122								
<i>Spratelloides gracilis</i>		200			15		63		
<i>Squalus megalops</i>	8	1							
<i>Squatina sp.</i>									
<i>Stephanolepis auratus</i>	103	3							
<i>Stolephorus heterolobus</i>	29	100					30		
<i>Stolephorus indicus</i>	13	8							
<i>Stormias boa boa</i>	47								
SYNODONTIDAE	4								
<i>Teixeirichthys jordani</i>	2	102							
<i>Terapon theraps</i>	13	1							
Tetraodontidae	4								
<i>Tetrosomus concatentus</i>	14								
<i>Thenus orientalis</i>	15	14							
<i>Thryssa vitrirostris</i>	4								
<i>Thunus orientalis</i>	2								
<i>Thyrsitoides marleyi</i>		39							
<i>Torquigener hypselogenion</i>	10	1							
<i>Trachinocephalus myops</i>		4							
<i>Trichiurus lepturus</i>	14	115	60		60		30		
<i>Tydemania navigatoris</i>	74								
<i>Tylerius spinosissimus</i>	4								
Unidentified fish	20	10						4	
<i>Upeneus bensasi</i>	220	32							
<i>Upeneus guttatus</i>	4								
<i>Upeneus moluccensis</i>	163	28							
<i>Upeneus sp.</i>	116								
<i>Upeneus sulphureus</i>		61			30		30		
<i>Upeneus taeniopterus</i>	40	192	30		30		30		
<i>Upeneus vittatus</i>	2								
<i>Uranoscopus archionema</i>	32								
<i>Uroconger lepturus</i>	2								
<i>Uroconger sp.</i>	1								
<i>Uroteuthis duvadelli</i>	100	32							
<i>Uroteuthis sp</i>	132								

Species/groups	Length	Length/	Biology	Liver	Stomach	Otoliths	Fin	Frozen	Preserved
		Weight					clips		
<i>Velodona togata</i>	1								
<i>Ventrifossa mystax</i>		29							
<i>Xenolepidichthys dagleishi</i>	22								
<i>Zanclus cornutus</i>	1								
<i>Zenion</i> sp.	97								
<i>Zeus capensis</i>	1								
<i>Zeus faber</i>	2								
Total	7 083	3 664	395	1	234		531	15	8

ANNEX V. SPECIES/GROUPS CAUGHT IN 30 BOTTOM TRAWL HAULS ALONG THE COAST OF TANZANIA

Each haul was standardised to 60 minutes of towing. Weight -summarised catch in all hauls, No. of fish – summarised no of fish, No. of hauls – no. of hauls with catch.

<i>Species/groups</i>	Weight (kg)	No. of fish	No. of hauls
<i>Engrasicholina heteroloba</i>	1 397.0	499 078	2
<i>Trichiurus lepturus</i>	697.1	1 227	4
<i>Upeneus taeniopterus</i>	620.3	24 584	6
<i>Decapterus macrosoma</i>	559.4	18 840	8
<i>Saurida undosquamis</i>	374.1	3 036	25
<i>Rexea prometheoides</i>	353.4	4 583	8
<i>Secutor insidiator</i>	307.6	19 823	3
<i>Carangoides malabaricus</i>	279.0	6 178	8
<i>Argentina euchi</i> s	231.7	4 025	8
<i>Leiognathus elongatus</i>	193.0	16 418	7
MYCTOPHIDAE	173.8	29 088	8
<i>Nettastoma parviceps</i>	165.6	1 957	2
<i>Polymixia berndti</i>	153.4	6 497	11
<i>Centrophorus granulosus</i>	136.4	67	3
<i>Zenion</i> sp.	124.4	1 232	1
<i>Aluterus monoceros</i>	115.4	46	2
<i>Upeneus moluccensis</i>	113.3	3 530	5
<i>Gazza minuta</i>	105.3	5 570	3
<i>Decapterus russelli</i>	103.3	1 462	8
<i>Polysteganus coeruleopunctatus</i>	102.5	97	4
<i>Abalistes stellatus</i>	93.1	148	9
<i>Himantura jenkinsii</i>	89.9	8	2
<i>Scomberomorus commerson</i>	85.4	54	4
<i>Leiognathus berbis</i>	84.5	41 814	5
<i>Upeneus bensasi</i>	82.4	6 823	6
<i>Peristedion weberi</i>	80.7	943	11
<i>Rastrelliger kanagurta</i>	77.2	1 270	8
<i>Decapterus macarellus</i>	74.0	1 434	1
<i>Dussumieria acuta</i>	72.7	2 182	6
<i>Loligo duvauceli</i>	72.1	2 462	14
<i>Rachycentron canadum</i>	69.8	31	1
<i>Sphyraena obtusata</i>	67.1	1 424	2
<i>Malacocephalus laevis</i>	63.8	1 047	13
<i>Sphyraena jello</i>	55.3	102	1
<i>Apogon</i> sp.	52.9	3 601	11
<i>Lepidotrigla faueri</i>	47.7	230	4
<i>Diagramma centurio</i>	47.6	14	2
<i>Muraenesox bagio</i>	45.0	14	1
<i>Thenus orientalis</i>	44.4	275	8
<i>Polymetme corythaëola</i>	43.8	2 189	8
<i>Urotrychis duvaucelii</i>	43.2	1 373	11
<i>Squalus megalops</i>	43.2	48	6
<i>Stephanolepis auratus</i>	42.4	6 990	3
<i>Uranoscopus archionema</i>	41.4	201	4
<i>Epinephelus coioides</i>	40.6	2	1
<i>Epinephelus malabaricus</i>	39.3	25	2
<i>Lethrinus variegatus</i>	36.0	598	1
<i>Gerres filamentosus</i>	34.9	173	4
<i>Narcine riera</i>	34.9	125	3
<i>Sepia vermiculata</i>	34.8	90	4
<i>Lagocephalus sceleratus</i>	34.2	645	5
<i>Centrophorus moluccensis</i>	34.1	14	2
<i>Lophiodes mutilus</i>	31.2	26	4
<i>Chirocentrus dorab</i>	29.7	94	2
<i>Satyrichthys adeni</i>	29.5	71	4
<i>Balistes capriscus</i>	29.2	23	1
<i>Arius africanus</i>	28.7	8	2

<i>Species/groups</i>	Weight (kg)	No. of fish	No. of hauls
<i>Arothron stellatus</i>	28.6	11	4
<i>Nemipterus metopias</i>	28.2	327	3
<i>Chlorophthalmus agassizi</i>	28.1	424	3
<i>Gymnocranius griseus</i>	27.7	198	5
<i>Chaunax</i> sp.	27.4	209	10
<i>Etomopterus sentosus</i>	27.2	399	7
<i>Arius venosus</i>	27.1	6	1
<i>Polyipnus indicus</i>	26.0	3 910	7
<i>Gnathanodon speciosus</i>	25.9	4	1
<i>Leiognathus leuciscus</i>	23.9	1 504	3
<i>Ariopsis</i> sp.	23.6	10	1
<i>Cynoglossus capensis</i>	23.1	369	2
<i>Teixeirichthys jordani</i>	22.9	1 773	4
<i>Sepia hieronis</i>	22.6	404	12
<i>Pomadasys maculatus</i>	21.9	168	1
<i>Upeneus</i> sp.	21.0	1 317	1
<i>Leiognathus equulus</i>	20.9	278	3
SHRIMPS	20.5	989	2
<i>Scolopsis bimaculata</i>	20.5	1 494	1
<i>Dalatias licha</i>	20.1	2	1
<i>Citharoides macrolepis</i>	19.9	249	7
<i>Parupeneus</i> sp.	19.9	360	2
<i>Rhinobatos holcorhynchus</i>	19.5	16	1
<i>Tydemania navigatoris</i>	19.2	481	11
<i>Lutjanus sanguineus</i>	19.2	4	1
<i>Acroteriobatus zanzibarensis</i>	18.9	11	2
<i>Linuparus somniosus</i>	18.9	42	3
<i>Squatina</i> sp.	18.3	2	1
<i>Peristedion</i> sp.	17.8	36	1
<i>Carangooides fulvoguttatus</i>	17.7	2	1
<i>Lutjanus rivulatus</i>	17.6	2	1
<i>Ostracion cubicus</i>	17.6	16	3
RAJIDAE	17.5	108	3
<i>Spratelloides</i> sp.	17.5	1 016	7
<i>Pteromylaeus bovinus</i>	17.4	2	1
<i>Psettodes erumei</i>	17.2	10	3
<i>Nemipterus japonicus</i>	16.1	238	4
<i>Pseudobalistes flavidorsalis</i>	16.1	5	1
<i>Priacanthus hamrur</i>	16.1	142	2
<i>Paramonacanthus pusillus</i>	16.0	669	3
CARIDAE	15.9	1 168	1
<i>Chascanopsetta lugubris</i>	15.9	30	2
<i>Zeus capensis</i>	15.7	6	1
<i>Pseudorhombus elevatus</i>	14.7	1 530	1
<i>Citharus</i> sp.	14.7	178	1
<i>Upeneus sulphureus</i>	14.7	466	1
<i>Penaeopsis balssi</i>	14.6	133	1
<i>Terapon theraps</i>	14.4	341	4
<i>Carangooides</i> sp.	13.8	191	3
<i>Atrobucca nibe</i>	13.7	42	4
<i>Diodon hystrix</i>	13.1	11	2
<i>Carangooides dinema</i>	12.9	4	1
<i>Bythaelurus lutarius</i>	12.7	159	6
<i>Polymixia</i> sp.	12.6	545	3
<i>Pristiphorus nancyae</i>	12.6	93	2
<i>Lethrinus</i> sp.	12.5	242	3
<i>Epinephelus areolatus</i>	12.1	6	2
<i>Lethrinus crocineus</i>	11.9	2	1
<i>Pterois miles</i>	11.6	47	3
<i>Heterodontus ramalheira</i>	11.4	2	1
<i>Spratelloides gracilis</i>	11.4	5 115	1
SOFT SPONGES	11.2	0	1
<i>Lestrolepis intermedia</i>	10.9	442	6
<i>Tetrosomus concatenate</i>	10.9	33	3
<i>Stolephorus waitei</i>	10.4	4 956	1

Species/groups	Weight (kg)	No. of fish	No. of hauls
<i>Ventrifossa mystax</i>	10.0	299	1
<i>Octopus</i> sp.	9.9	16	1
<i>Torpedo</i> sp.	9.8	3	1
<i>Sphyraena putnamae</i>	9.8	22	3
<i>Carangoides chrysophrys</i>	9.7	25	1
<i>Nemipterus zysron</i>	9.3	163	2
<i>Parapenaeus investigatoris</i>	9.2	800	1
<i>Aristeus antennatus</i>	9.0	440	3
<i>Otolithes ruber</i>	9.0	29	2
<i>Scorpaena scrofa</i>	8.7	16	1
<i>Neobythites kenyensis</i>	8.5	117	5
<i>Parascolopsis</i> sp.	8.3	266	1
<i>Apistus carinatus</i>	8.3	491	2
<i>Xenolepidichthys dagleishi</i>	8.2	315	2
<i>Parapenaeus</i> sp.	8.1	1 245	1
<i>Lagocephalus inermis</i>	8.1	53	2
Unidentified fish	8.0	386	7
<i>Holohaelurus punctatus</i>	7.9	4	1
<i>Neoepinnula orientalis</i>	7.8	231	3
Unidentified crustacean	7.6	694	2
CARCHARHINIDAE	7.6	2	1
<i>Pomadasys kaakan</i>	7.6	2	1
PENAEIDAE	7.5	377	3
<i>Fistularia petimba</i>	7.5	196	9
<i>Drepane</i> sp.	7.4	67	1
PROCESSIDAE	7.2	114	1
<i>Cubiceps</i> sp.	6.8	86	1
<i>Parabembras</i> sp.	6.7	38	2
<i>Raja</i> sp.	6.6	45	3
<i>Zeus faber</i>	6.5	7	1
<i>Stolephorus indicus</i>	6.4	251	3
<i>Eridacnis radcliffei</i>	6.3	123	4
<i>Sepia</i> sp.	6.1	134	3
<i>Neobythites cf somaliaensis</i>	5.9	18	2
<i>Sepia trigonina</i>	5.9	63	1
<i>Seriolina nigrofasciata</i>	5.8	7	2
<i>Thyrsitoides marleyi</i>	5.8	129	2
<i>Upeneus vittatus</i>	5.6	50	2
OSTRACIIDAE	5.6	12	1
<i>Carcharhinus</i> sp.	5.5	9	1
<i>Bothus</i> sp.	5.3	55	2
<i>Gymnoscopelus</i> sp.	5.3	1 147	2
<i>Himantura</i> sp.	5.2	22	1
<i>Atlantoraja cyclophora</i>	5.2	16	1
<i>Nemipterus bipunctatus</i>	5.1	56	3
<i>Sphyraena flavicauda</i>	5.1	126	1
<i>Cynoglossus lida</i>	5.0	122	7
<i>Dipturus stenorhynchus</i>	5.0	6	1
<i>Decapterus kurroides</i>	5.0	346	3
<i>Paracallionymus costatus</i>	4.9	244	1
<i>Carcharhinus limbatus</i>	4.7	2	1
<i>Sepia prashadi</i>	4.7	30	2
<i>Physiculus natalensis</i>	4.7	43	5
<i>Lutjanus fulvus</i>	4.7	578	1
<i>Penaeus semisulcatus</i>	4.6	116	2
<i>Projasus parkeri</i>	4.6	293	1
<i>Pseudobalistes fuscus</i>	4.4	2	1
<i>Cylichthys</i> sp.	4.3	8	1
<i>Cociella</i> sp.	4.3	16	1
<i>Ornithoteuthis volatilis</i>	4.3	54	1
<i>Lethrinus microdon</i>	4.1	49	2
<i>Ruvettus pretiosus</i>	4.1	28	2
<i>Metanephrops mozambicus</i>	4.1	48	1
DIODONTIDAE	4.0	3	1
<i>Carangoides ferau</i>	4.0	86	1

Species/groups	Weight (kg)	No. of fish	No. of hauls
<i>Heptanchias perlo</i>	4.0	6	1
<i>Haliporoides triarthrus</i>	3.9	92	1
<i>Seriola rivoliana</i>	3.8	0	1
<i>Nemipterus</i> sp.	3.6	31	2
<i>Palinurus</i> sp.	3.5	27	1
<i>Lepidopus caudatus</i>	3.3	6	1
<i>Bembrops</i> sp.	3.3	27	2
<i>Lactoria diaphana</i>	3.3	14	1
ARGENTINIDAE	3.3	26	1
<i>Cubiceps whiteleggi</i>	3.2	76	1
<i>Dysomma anguillare</i>	3.2	27	1
SYNODONTIDAE	3.2	32	1
SPARIDAE	3.1	204	1
<i>Parupeneus macronemus</i>	3.0	60	1
<i>Plotosus lineatus</i>	3.0	531	4
<i>Diaphus knappi</i>	2.9	0	1
<i>Scomberops boops</i>	2.8	41	1
OPHIDIIDAE	2.8	52	1
<i>Parupeneus heptacanthus</i>	2.7	48	3
<i>Tylerius spinosissimus</i>	2.6	8	1
<i>Cociella</i> sp.	2.6	16	1
<i>Solenocera agoensis</i>	2.6	54	1
<i>Torquigener hypselogenion</i>	2.5	81	2
<i>Etelis carbunculus</i>	2.4	2	1
<i>Callionymus marleyi</i>	2.4	0	1
<i>Branchiostegus doliatus</i>	2.3	15	2
<i>Arothron hispidus</i>	2.3	2	1
<i>Oxyurichthys petersii</i>	2.3	168	1
<i>Thryssa vitrirostris</i>	2.3	112	1
CREPIDULIDAE	2.2	16	1
<i>Velodona togata</i>	2.2	8	2
<i>Cruriraja parcomaculata</i>	2.2	26	2
<i>Bothus myriaster</i>	2.2	79	2
<i>Leucoraja wallacei</i>	2.2	20	1
<i>Saurida tumbil</i>	2.2	6	1
<i>Ateleopus natalensis</i>	2.2	4	1
<i>Bembrops platyrhynchus</i>	2.1	47	3
<i>Sepia latimanus</i>	1.9	75	1
<i>Narcine</i> sp.	1.9	12	1
<i>Lepidotrigla</i> sp.	1.9	37	2
<i>Satyrichthys investigatoris</i>	1.8	60	2
<i>Lethrinus nebulosus</i>	1.8	32	1
<i>Avocettina</i> sp.	1.8	26	2
<i>Siganus sutor</i>	1.8	45	1
<i>Polynemus sextarius</i>	1.7	56	1
<i>Lophius</i> sp.	1.7	16	1
<i>Paratriacanthodes retrospinis</i>	1.6	51	3
<i>Uroconger lepturus</i>	1.6	9	1
<i>Bathycongrus wallacei</i>	1.6	19	2
<i>Halaelurus</i> sp.	1.6	16	1
<i>Rostroraja alba</i>	1.6	3	1
<i>Antigonia rubescens</i>	1.5	71	2
<i>Choerodon gymnocephalus</i>	1.5	40	3
<i>Trachinocephalus myops</i>	1.5	47	1
<i>Ibacus novemdentatus</i>	1.5	8	1
<i>Phosichthys argenteus</i>	1.5	101	1
<i>Paramonacanthus</i> sp.	1.5	179	2
<i>Stomias boa</i> <i>boa</i>	1.4	265	3
<i>Antigonia capros</i>	1.4	25	3
<i>Lagocephalus guentheri</i>	1.4	66	2
<i>Neoscopelus</i> sp.	1.3	8	1
<i>Lepidoteuthis</i> sp.	1.3	12	1
<i>Lactoria cornuta</i>	1.3	4	1
<i>Arnoglossus</i> sp.	1.2	56	2
<i>Caranx</i> <i>tille</i>	1.2	23	1

Species/groups	Weight (kg)	No. of fish	No. of hauls
<i>Dussumieri acuta</i>	1.2	62	1
<i>Harpisquilla harpax</i>	1.2	2	1
<i>Macrorhamphosus scolopax</i>	1.2	116	5
<i>Pliotrema warreni</i>	1.2	8	1
<i>Metapenaeus monoceros</i>	1.2	118	1
<i>Etmopterus</i> sp.	1.2	24	1
<i>Caelorinchus trunovi</i>	1.1	12	2
<i>Epigonus robustus</i>	1.1	17	1
<i>Scorpaena</i> sp.	1.0	18	1
<i>Mene maculata</i>	1.0	24	2
<i>Herklotischthys quadrimaculatus</i>	1.0	16	1
<i>Coelorinchus trunovi</i>	0.9	38	1
CHLOROPHTHALMIDAE	0.9	67	1
<i>Apogon quadrifasciatus</i> **	0.8	41	4
<i>Amanses scopas</i>	0.8	69	1
<i>Carangooides armatus</i>	0.8	54	1
<i>Pterois russelii</i>	0.8	9	1
<i>Zenion hololepis</i>	0.8	83	1
CRABS	0.7	18	1
<i>Champsodon capensis</i>	0.7	18	1
<i>Champsodon</i> sp.	0.7	118	1
<i>Macrorhamphosus</i> sp.	0.7	8	1
<i>Rhinochimaera atlantica</i>	0.7	18	1
CARANGIDAE	0.6	15	1
<i>Coelorinchus</i> sp.	0.6	35	1
<i>Holohaelurus regani</i>	0.6	2	1
<i>Monocentris japonica</i>	0.5	4	1
<i>Poecilopsetta zanzibarensis</i>	0.5	29	2
<i>Sphyraena forsteri</i>	0.5	2	1
<i>Upeneus guttatus</i>	0.5	9	1
<i>Thunnus orientalis</i>	0.5	5	1
<i>Lophiodes</i> sp.	0.5	9	1
<i>Gerres oyena</i>	0.5	8	1
<i>Sphyraena guachancho</i>	0.5	8	1
DACTYLOPTERIDAE	0.5	8	1
<i>Choerodon</i> sp.	0.5	24	1
<i>Synodus</i> sp.	0.5	30	1
<i>Cynoglossus marleyi</i>	0.4	6	1
<i>Sardinella gibbosa</i>	0.4	66	1
<i>Acropoma japonicum</i>	0.4	6	1
<i>Poecilopsetta zanzibarensis</i>	0.4	8	1
<i>Ateleopus</i> sp.	0.4	6	1
<i>Pontinus nigerimum</i>	0.4	9	2
<i>Sphyraena barracuda</i>	0.4	9	1
<i>Linuparus</i> sp.	0.4	12	1
<i>Rogadius asper</i>	0.4	12	1
<i>Trypauchen microcephalus</i>	0.3	56	1
Unidentified demersal fish	0.3	28	2
<i>Beryx splendens</i>	0.3	9	2
<i>Leiognathus splendens</i>	0.3	9	1
<i>Neobythites analis</i>	0.3	18	1
SALPS	0.3	12	1
<i>Synagrops</i> sp.	0.3	6	1
<i>Octopus cyaneus</i>	0.3	28	1
<i>Coelorinchus denticulatus</i>	0.2	10	2
<i>Samaris cristatus</i>	0.2	16	1
<i>Lepidotrigla multispinosa</i>	0.2	8	1
<i>Uroconger</i> sp.	0.2	8	1
<i>Citharichthys</i> sp.	0.2	40	3
BOTHIDAE	0.2	30	1
<i>Canthigaster</i> sp.	0.2	23	1
<i>Laeops natalensis</i>	0.2	28	1
<i>Gobius</i> sp.	0.2	49	1
SERRANIDAE	0.2	10	1
<i>Penaeus marginatus</i>	0.1	2	1

<i>Species/groups</i>	Weight (kg)	No. of fish	No. of hauls
<i>Oxycheilinus bimaculatus</i>	0.1	15	1
<i>Lepidotrigla alcocki</i>	0.1	8	1
<i>Parabathynomus natalensis</i>	0.1	6	1
<i>Bothus pantherinus</i>	0.1	2	1
<i>Saurida gracilis</i>	0.1	18	2
<i>Hoplichthys acanthopleurus</i>	0.1	8	1
<i>Macrorhamphosus gracilis</i>	0.1	8	1
<i>Parapenaeus fissurus</i>	0.1	8	1
<i>Apogonidae - juvenile</i>	0.1	23	1
<i>Echeneis naucrates</i>	0.1	10	1
<i>Amblygaster leiogaster</i>	0.0	10	1
<i>Laeops nigromaculatus</i>	0.0	6	1
LUTJANIDAE	0.0	10	1
<i>Fishing gears</i>	0.0	2	1
<i>Glass sponge</i>	0.0	5	1
<i>Invertebrate</i>	0.0	4	1
SEA URCHINS	0.0	2	1

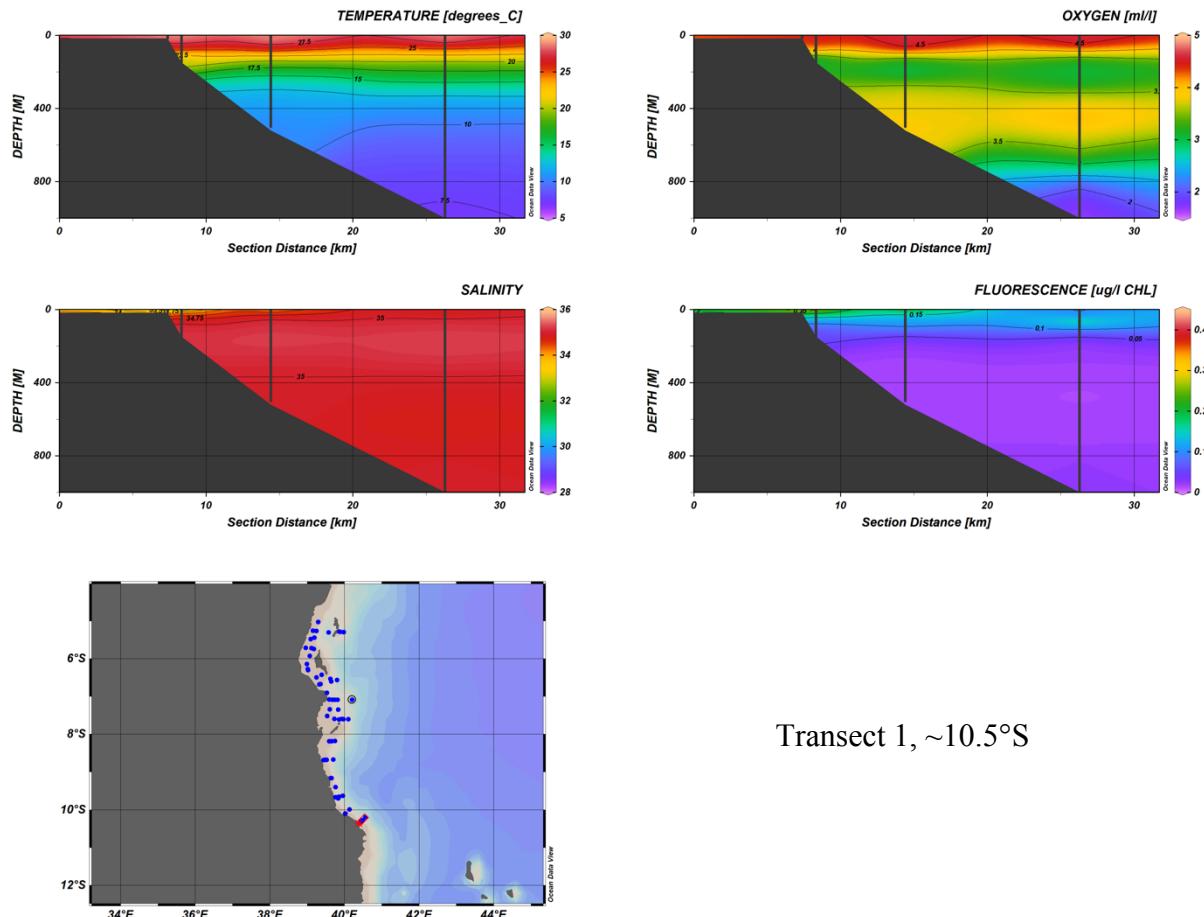
ANNEX VI. SPECIES/GROUPS CAUGHT IN 7 PELAGIC TRAWL HAULS ALONG THE COAST OF TANZANIA

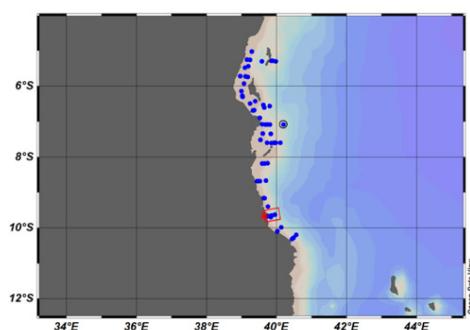
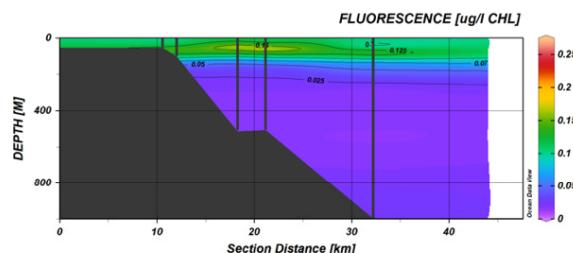
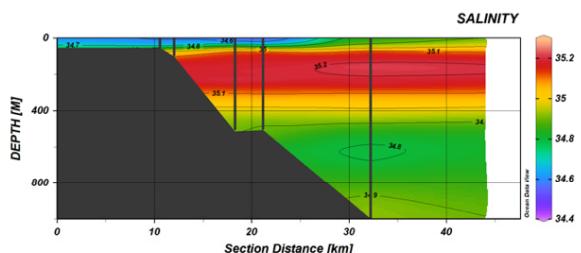
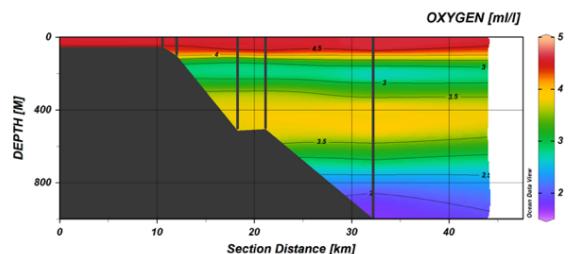
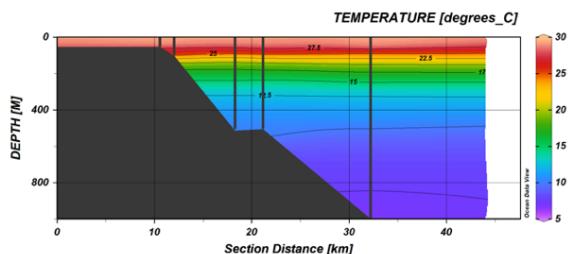
Each haul was standardised to 60 minutes of towing. Weight -summarised catch in all hauls, No. of fish – summarised no of fish, No. of hauls – no. of hauls with catch.

Species/groups	Weight (kg)	No. of fish	No. of hauls
<i>Engraulis encrasicolus punctifer</i>	211.5	88144	1
<i>Rastrelliger kanagurta</i>	53.2	74	2
MYCTOPHIDAE	49.0	19604	4
<i>Cubiceps pauciradiatus</i>	37.5	-	1
<i>Leiognathus berbis</i>	28.1	-	1
<i>Spratelloides gracilis</i>	17.4	10859	1
<i>Engraulis heteroloba</i>	13.6	7684	1
<i>Decapterus macrosoma</i>	10.9	1479	1
<i>Amblygaster sirm</i>	5.2	676	1
<i>Decapterus kurroides</i>	4.8	280	1
<i>Onychoteuthis</i> sp.	3.5	123	1
<i>Trichiurus lepturus</i>	2.9	5	1
<i>Uroteuthis duvaucelii</i>	2.0	235	2
<i>Polytmus coryphaeola</i>	1.7	477	1
<i>Leptocephalus</i>	1.2	331	2
<i>Lestrolepis intermedia</i>	1.2	140	4
<i>Leiognathus elongatus</i>	0.9	109	1
<i>Decapterus tabl</i>	0.7	12	2
<i>Muraenesox bagio</i>	0.7	12	1
UNIDENTIFIED FISH	0.5	185	1
<i>Seriola rivoliana</i>	0.4	2	1
<i>Sphyraena obtusata</i>	0.3	15	1
<i>Cheilopogon nigricans</i>	0.3	6	1
<i>Apogon</i> sp.	0.3	32	2
<i>Bregmaceros</i> sp.	0.2	13	1
<i>Dipterygonotus balteatus</i>	0.2	8	1
LOLIGINIDAE	0.2	218	1
SEPIIIDAE	0.2	23	1
<i>Loligo duvaucelii</i>	0.2	79	1
JELLY FISH	0.2	23	1
<i>Samaris cristatus</i>	0.1	15	1
<i>Scomber japonicus</i>	0.0	4	1
<i>Zanclus cornutus</i>	0.0	2	1
Carangidae	0.0	66	1
<i>Hemiramphus</i> sp.	0.0	2	1
TETRAODONTIDAE	0.0	43	1

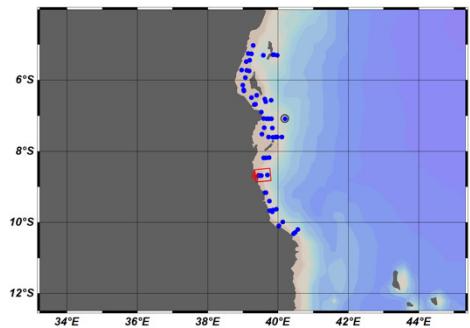
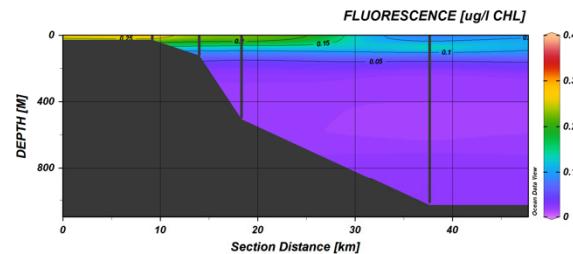
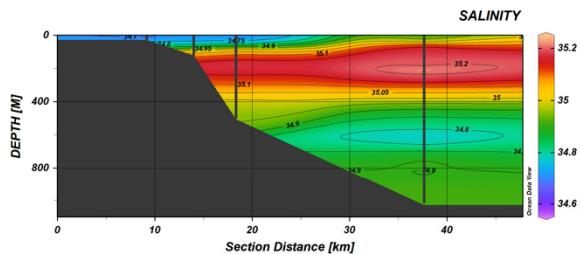
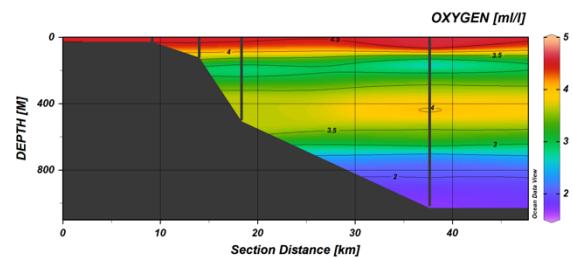
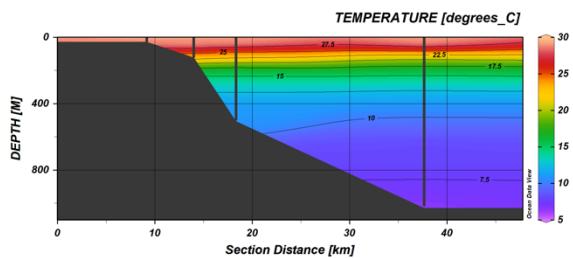
ANNEX VII. HYDROGRAPHICAL TRANSECTS

In this Annex, data obtained at super transects are presented. These transects were carried out perpendicular to the coast line, except for the transect along the Zanzibar Channel. Geographical position of the various transects are indicated in the attached maps. The transects are organised from south to north.

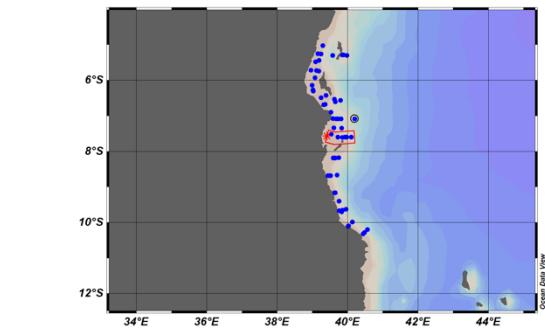
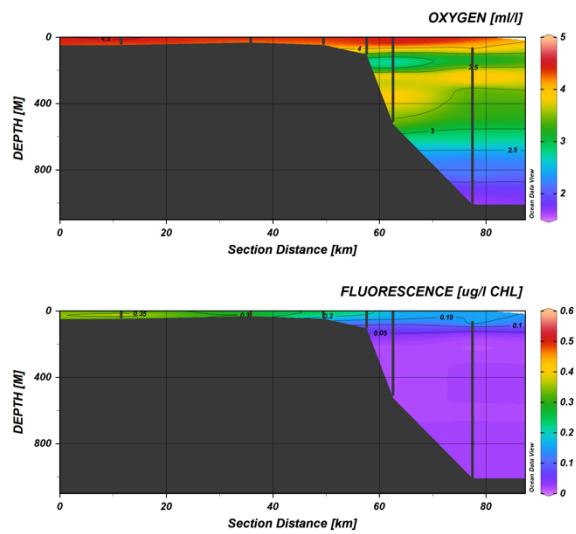
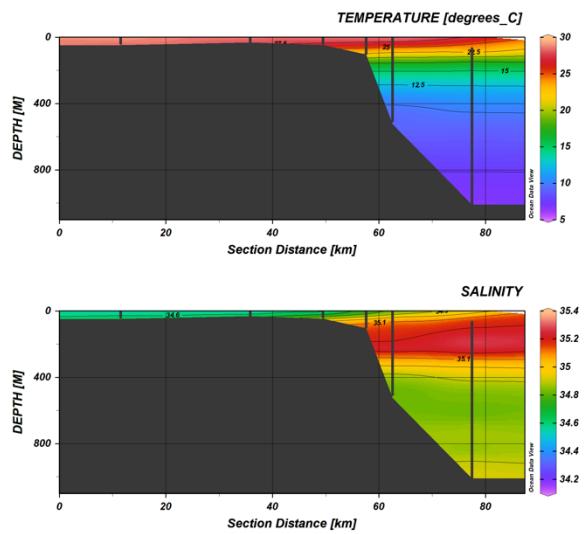




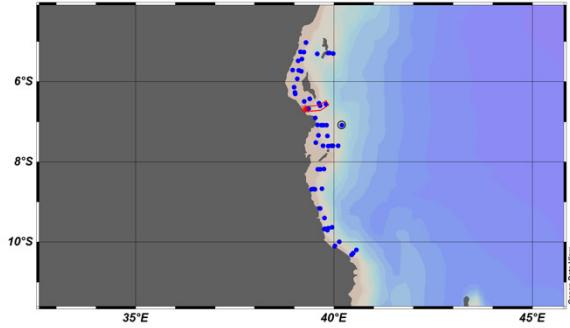
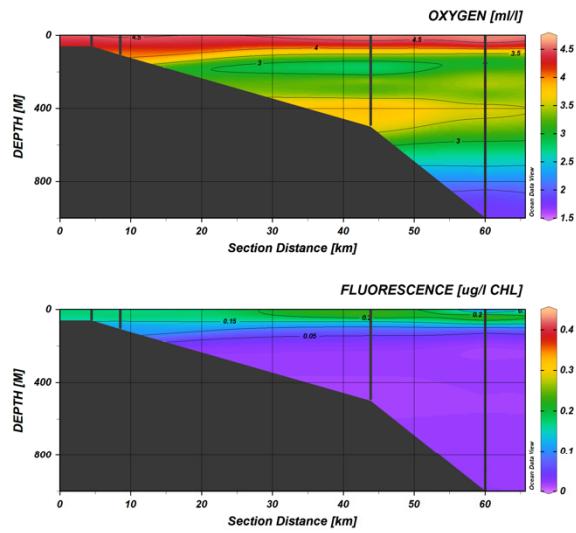
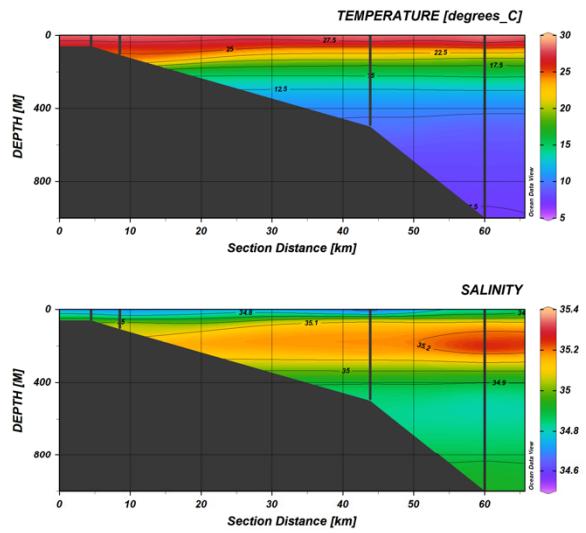
Transect 2, ~9.5°S



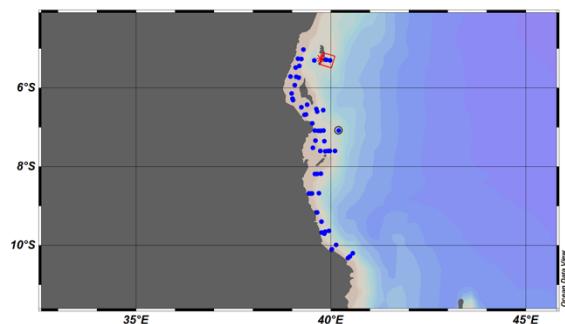
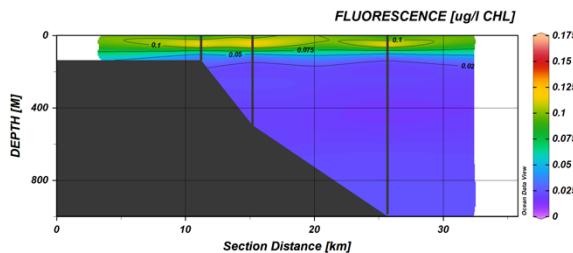
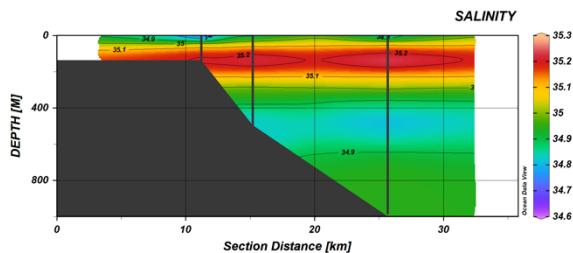
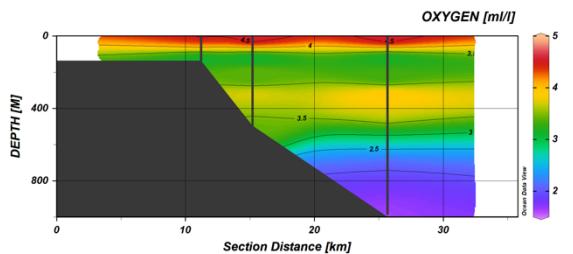
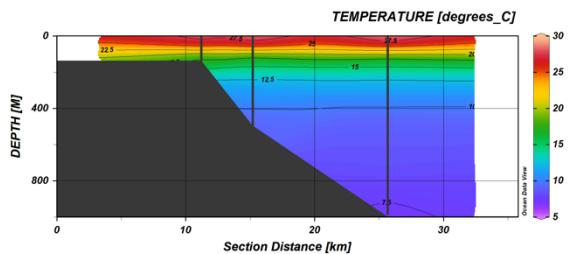
Transect 3, ~10.5°S



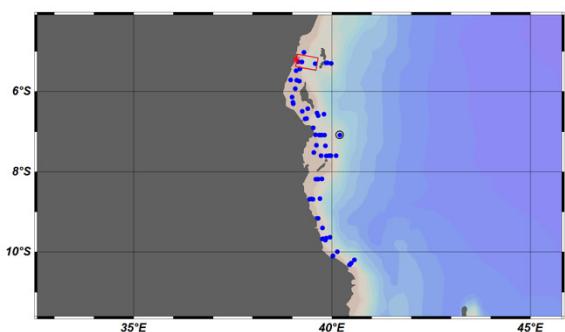
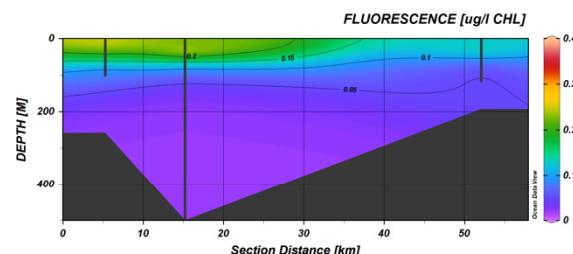
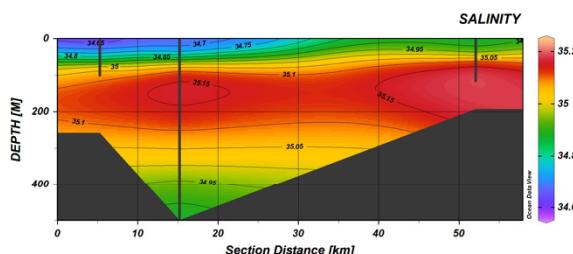
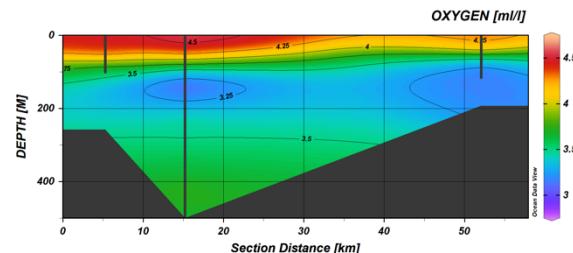
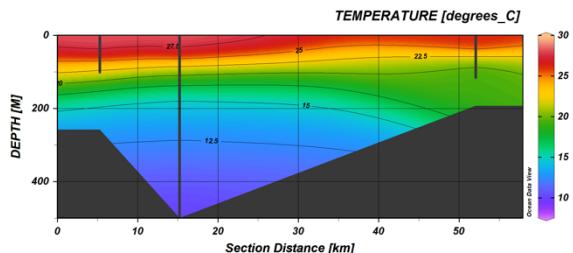
Transect 4, ~7.5°S



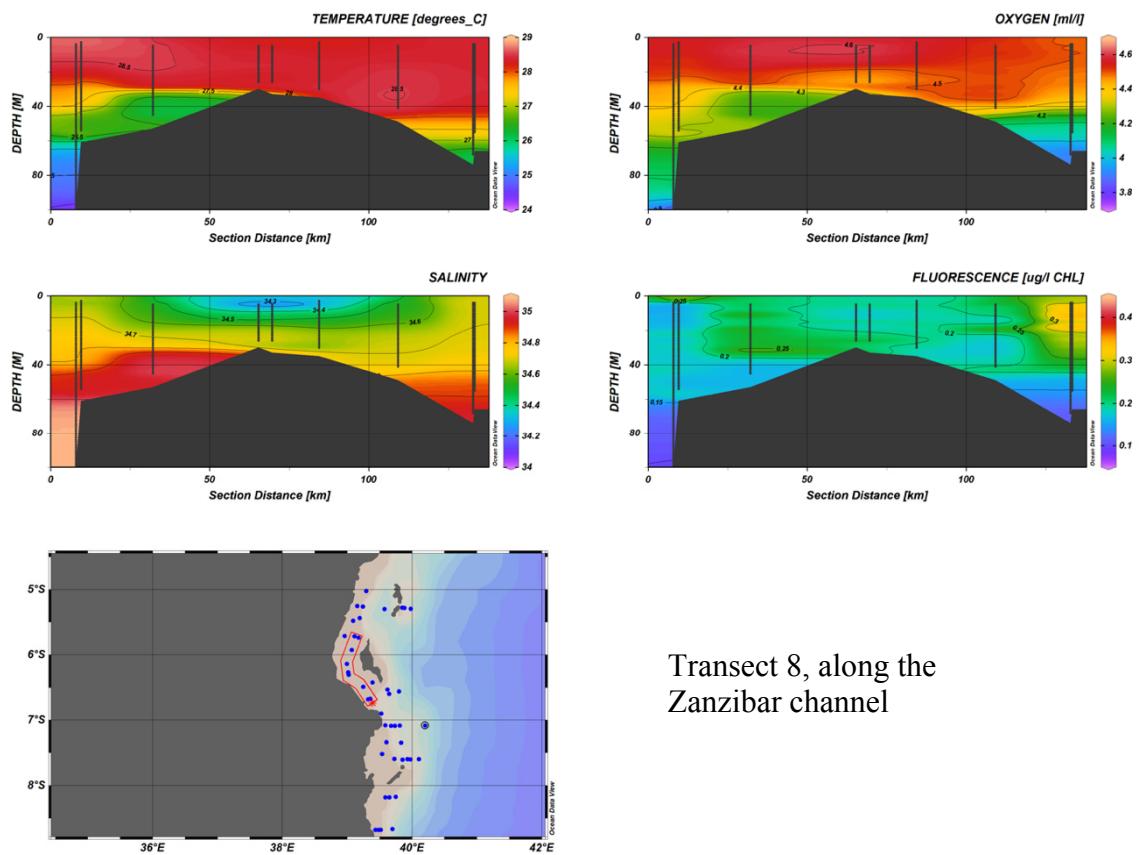
Transect 5, ~6.5°S



Transect 6, $\sim 5.5^{\circ}\text{S}$
outside Pemba



Transect 7, $\sim 5.5^{\circ}\text{S}$ in
the Pemba channel



Transect 8, along the Zanzibar channel

ANNEX VIII.

OVERVIEW OF COLLECTED SAMPLES

Gear/equipment	Samples	Preservation	Port of off Loading	Transport	Institution address	Contact person	Number	Status
Niskin bottles	Nutrients	Chloroform	Norway	Air freight	IMR	Lars Naustvoll		processed
Niskin bottles	Chlorophyll a	Frozen	Norway	Air freight	IMR	Lars Naustvoll		Processed
Algae net	Phytoplankton	Formaldehyde	Dar-es-Salam	To Zanzibar	IMS	M.s. Kyewalyanga	19	Unprocessed
Niskin bottles	DIC/TA	N/A	N/A	N/A	N/A	N/A	N/A	Processed on board
WP2 (180 µm): (1) vertical haul from max 200 m) – ½ sample, (2) vertical haul from 30 m – ½ sample	Zooplankton (biomass)	Dried	IMR	Air freight	IMR	Stamatina Isari	36	Processed
(1) vertical haul from max 200 m)- ½ sample (2) vertical haul from 30 m – ½ sample	Zooplankton identification	Formaldehyde	Dar-es-Salam	Boat	IMS	M.s. Kyewalyanga	36	Unprocessed
MultiNet (Midi, 1 x 180 µm): oblique tow from max 200 m	Fish eggs and larvae	Formaldehyde	Dar-es-Salam	Boat	IMS	M.s. Kyewalyanga	21	Unprocessed
Manta trawl (375 µm): surface tow for 15 mins	Jelly fish	Formaldehyde	South Africa	Air freight	South Africa	Mark Gibbons	Unknown	Unprocessed
Manta trawl (375 µm): surface tow for 15 mins	Microplastics	Photographed and frozen	Norway	Air freight	IMR	Bjørn Einar Grøsvik	21	Initial analysis done on board
Trawl	Unidentified fish samples	Formaldehyde	Norway	Air freight	IMR	Peter Pomasakis	Unknown	Unprocessed
Trawl	Finclips of priority species for genetic analysis	Ethanol	Norway	Air freight	IMR	Erling Kåre Stenvik	531	Unprocessed
Trawl	Standard Nansen food safety sampling	Freeze-dried	Norway	Air freight	IMR	Vidar Fauskanger	142	Unprocessed
Trawl	Parasites in fish	UV-press method; frozen at -80°C (?)	Norway	Air freight	IMR	Lucilla Giulietti	Unknown	Unprocessed
Trawl - cylinder	Sediments grain size/Composition	Frozen	Norway	Air freight	IMR	Bjørn Einar Grøsvik	2x30	Unprocessed