Cruise report

HM2025009032

(HB25-265)

E. Darelius & B. Risebrobakken

1. Cruise overview

The cruise was organized as part of the project FJO2RD (Funded by RCN) and CLIFFORD (funded by the Bjerknes Centre) and we visited Masfjorden, Haugsværfjorden, Lurefjorden and Osterfjorden to do hydrographic work, collect water samples, and collect sediment cores and samples.

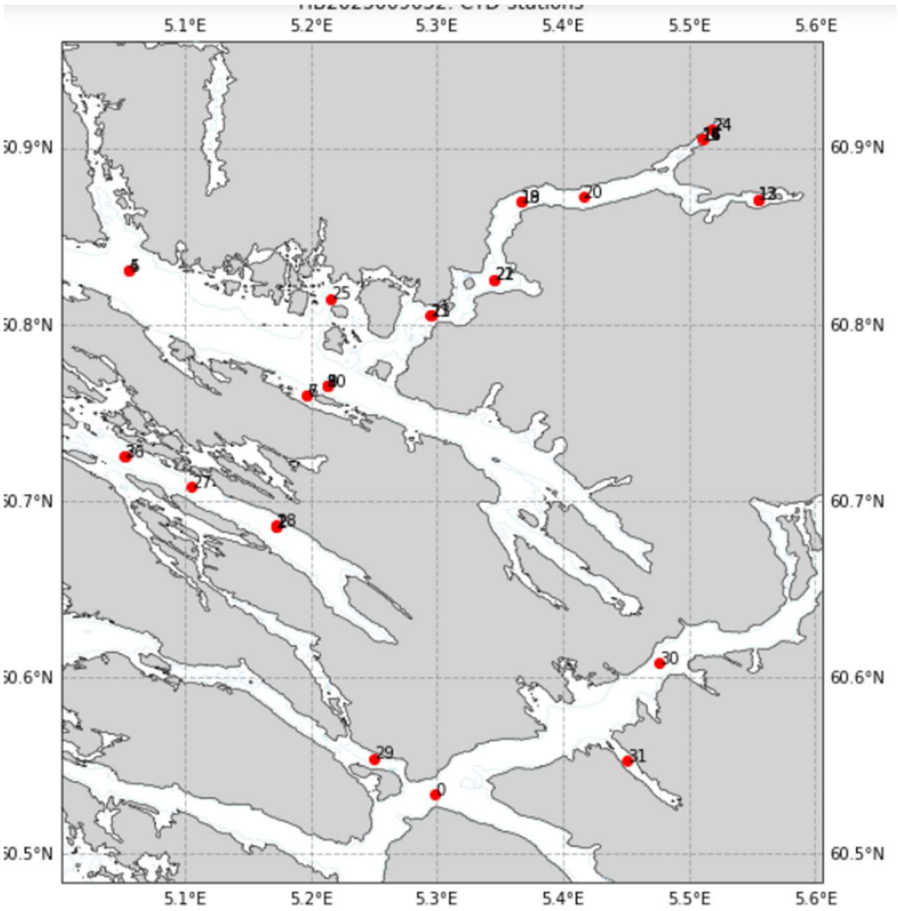

Figure 1: Map over the study area. CTD-stations are indicated by numbered, red dots.

2. Cruise participants

Leg 1: 23-24/6
Elin Darelius
Lea Chiche (observer)

Aurelien Thomas (student)
Monica Jaeger (observer/artist)
Kristin J-. Misje (technician)

Leg 2 : 24-27/6
Bjørg Risebrobakken
Dag-Inge Blindheim (technician)
Monica Jaeger (observer,artist)

3. CTD

We occupied a total of 31 CTD-casts during the cruise using the RBR Maestro sn 205914 available onboard. The conductivity, temperature and pressure sensor were last calibrated in February, 2025.

The stations are listed in

	St.name	Date			UTC	Depth	Latitude/ N		Longitude/ E		
CAST		year	mon	day	hh:mm	m	deg	min	deg	min	
0	O1	2025	6	23	8:13	543	60	32.050	5	17.925	
1	L6	2025	6	23	10:04	420	60	41.101	5	10.383	
2	L6	2025	6	23	10:45	420	60	41.101	5	10.383	
3	L4	2025	6	23	11:16	251	60	43.521	5	3.159	Tracemetals
4	F6	2025	6	23	12:52	539	60	49.813	5	3.344	
5	F6	2025	6	23	12:52	539	60	49.813	5	3.344	
6	F_Bjørg	2025	6	23	14:19	83	60	45.602	5	11.787	Two casts, forgot water
7	F_Bjørg	2025	6	23	14:25	83	60	45.602	5	11.787	
8	M4	2025	6	23	14:37	654	60	45.912	5	12.767	Bottle 5 did not close
9	M4	2025	6	23	15:20	654	60	45.912	5	12.767	Only bottle 5
10	M4	2025	6	23	15:30	654	60	45.912	5	12.767	
11	M12	2025	6	23	16:02	121	60	48.327	5	17.731	No bottles
12	M35	2025	6	24	06:21	179	60	52.258	5	33.331	
13	M35	2025	6	24	06:45	179	60	52.258	5	33.331	
14	HF	2025	6	24	07:15	121	60	54.284	5	30.658	Bottles were not reset
15	HF	2025	6	24	07:25	121	60	54.284	5	30.658	
16	HF	2025	6	24	07:54	121	60	54.284	5	30.658	
17	HF	2025	6	24	08:17	121	60	54.284	5	30.658	
18	MF24	2025	6	24	09:14	469	60	52.178	5	22.026	
19	MF24	2025	6	24	09:52	469	60	52.178	5	22.026	
20	MF26	2025	6	24	11:02	479	60	52.355	5	24.957	
21	M16	2025	6	24	11:43	297	60	49.533	5	20.754	
22	M16	2025	6	24	12:12	297	60	49.533	5	20.754	
23	M12	2025	6	24	12:49	107	60	48.302	5	17.694	
24	HF02	2025	6	25	12:32	43	60	54.604	5	31.0882	S N4, Trace N5
25	F_B2	2025	6	26	06:29	112	60	88.460	5	12.9676	S & Trace 1
26	L04	2025	6	26	11:00	244	60	43.511	5	3.1462	

27	L05	2025	6	26	11:53	390	60	42.481	5	6.3501	
28	L06	2025	6	26	12:53	425	60	41.176	5	10.3734	
29	St3	2025	6	27	05:30	197	60	33.252	5	15.018	
30	St8	2025	6	27	06:58	590	60	36.472	5	28.599	Trace elements
31	St6	2025	6	27	07:54	111	60	33.190	5	27,0576	Trace elements

Table 1.

Several of the stations were occupied two or three times, with one cast to the bottom followed by a shallower cast to collect water samples from the upper part of the water column.

4. Water sampling

Water samples were obtained from the Rosette available onboard during the upcast. As CTD data cannot be viewed live, sample depths had to be chosen beforehand. Sample depths are listed in

Cast	Station	Date & time	Latitude	Longitude	Temp	Samples
32	L03	23/6 11:34	60 43.931 N	5 01.556 E	13C	*Methane, DIC, Nutrients, NH4*
33	F03	23/6 12:25	60 51.184	4 52.511	13C	*Methane, DIC, Nutrients, NH4, Salt*
34	F08	23/6 14:01	60 47.44	5 7.845	13.5C	*Methane, DIC, Nutrients, NH4, Salt*
35	M28	24/6 8:50	60 52.54 N	5 26.33 E	13C	*Methane, DIC, Nutrients, NH4, Salt*
36	M26	24/6 8:59	60 52.37 N	5 24.94 E	13C	*Methane, DIC, Nutrients, NH4, Salt*
37	M22	24/6 11:33	60 51.07 N	5 20.99 E	14C	*Methane, DIC, Nutrients, NH4, Salt*

Table 2, where the depths are the depths given by the pressure sensor of the Rosette. We waited 60 seconds after the CTD stopped before releasing the bottle.

In addition, samples from the surface were collected using a separate water sampler. These samples were collected using a messenger with the sampler about 0.5 m below the surface. The temperature (as measured by the thermometer within the sampler) was noted down on the sampling sheet.

a) Dissolved oxygen

Samples for measuring dissolved oxygen were collected using a tube, ensuring each sample was as bubble-free and exposed to air as little as possible. Draw temp was measured before we added 1 mL MnCl2 and 1 mL NaOH/NaI to the sample and put a cap on the flask. The sample was then shaken for about 20 sec, and stored dark and cool until Winkler titration started.

Winkler titration was carried out by Kristin J. Misje at the Geophysical Institute, Bergen 25/6 using the automated titration system.

b) H2S

We expected anoxic conditions and H2S in the basin waters of Haugsværfjorden. The fjord had however renewed, and displayed an oxygen minimum around 55 m depth. At both 50 m and 40 m, one sample was taken for H2S along with one sample for dissolved oxygen. These were collected as for oxygen but 1 ml of ZnCl2 was added instead of the other chemicals. There was oxygen present at those depths so the samples were not analyzed for H2S.

c) Dissolved Inorganic Carbon / Alkalinity, Ph, and nutrients

Samples for carbon analysis (one bottle for dissolved inorganic carbon and alkalinity, and a separate bottle for pH) were collected using a tube, adding four drops (ca 0.1 ml) of mercuric chloride to the sampled bottles. Samples were kept cool and dark, and brought back to GFI for analysis.

Samples for nutrients were collected by rinsing the flasks three times, then adding a drop of chloroform to the sample. Samples were stored in the fridge, and sent to IMR for analysis.

Samples for NH4 were collected after rinsing the bottle three times, and the samples were placed in the freezer.

d) Salinity

The samples were collected following standard procedures, i.e., the bottles were rinsed three times and then brought back to GFI, where they were analyzed in the lab by K. Jackson-Misje.

Salinity samples were collected both from the CTD (for calibration) and from the surface sampler (for determination of salinity of the surface waters).

e) Methane & NO2

Samples for determination of methane and N2O concentrations were collected following standard procedures: the vials were rinsed three times before they were overfilled and 0.2 ml of $ZnCl_2$ was added using a syringe. The vials were then capped and crimped, and stored upside down in the fridge. The samples were brought to Gothenburg by E. Darelius for analysis by T. Politi.

f) **Trace metals**

The LDPE bottles were labelled and rinsed three times with seawater (together with the lids) before the bottles were filled to the shoulder. 21 drops (1 ml) of 37% HCl was added to the

samples that were then put in double bags. The samples were stored in the fridge and later analyzed by A. Tisserand at NORCE, Bergen.

5. Sediment sampling

We occupied a total of 24 MC casts at 14 stations using the small (2 tubes) Multicorer from the Department of Earth sciences at the University of Bergen. At 9 stations, two casts were done.10 cores have been archived. At all stations, samples have been taken for analyses of living and dead foraminifera (stained) and unstained foraminifera (upper 10 cm) and for aDNA (upper 1cm). At one site the 0-1 cm slice was split in three, for aDNA, stained and unstained foraminifera. All details can be found in Table 4.

a) Foraminiferal assemblage and geochemical analyses

The upper 10 cm of one MC was sampled by extruding the sediment out of the core. Half the 1 cm sediment slices were put in bags and stored in the fridge. The other half was put into boxes and mixed with rose Bengal solution for analyses of living and dead foraminifera.

b) aDNA

At 14 stations the upper 0-1 cm of one of the MC tubes were sampled for aDNA. We avoided the material clos eto the liner.The core liner was cleaned to avoid contamination of surface sediments with other sediment. The upper cm was pushed up using the core extruder and excess water was removed. Clean lab gloves were put on and a sterile spoon was used to transfer the surface sediment into a sterile (labelled) bag. The sediment in contact with the core label was avoided and not included in the sample. The bag was closed and stored at -20°C while onboard. New gloves and spoons were used for each core.

c) Archived MCs

At 3 stations, 2 extra cores were stored for future analyses, at 4 stations 1 core was archived. At one station we got no cores. Here the cores were cleaned, water was tapped, and the sediment surface was secured using oasis, before the cores was capped and marked.

6. Calibration of CTD sensors

a) Salinity

The salinity of the water samples was determined by K. Jackson using a Portasal. 22 samples were collected, six of these were surface samples that were not included in the calibration.

Two samples were flagged as outliers (cast 24 and cast 29)

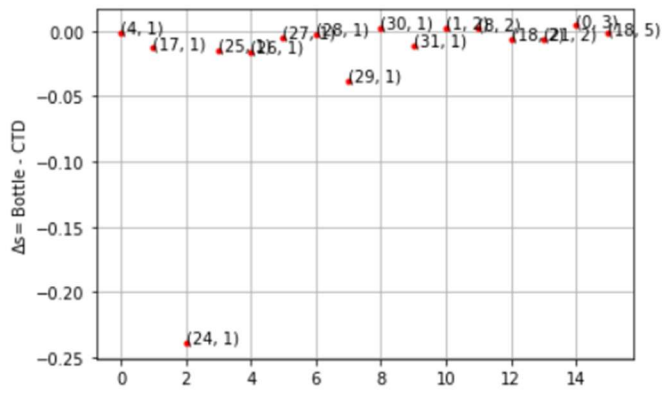

Figure 2: Difference between the measured salinity (Portasal) and that observed by the CTD. Results from station 24 and 29 were flagged as outliers.

Fourteen samples were included in the calibration. The difference between the bottle salinity and the sensor salinity was a function of pressure, and the correction applied was

$CTD_{corrected} = CTD_{uncorrected} - 1.602e\text{-}2 + 3.032e\text{-}5 * CTD_{pressure}$

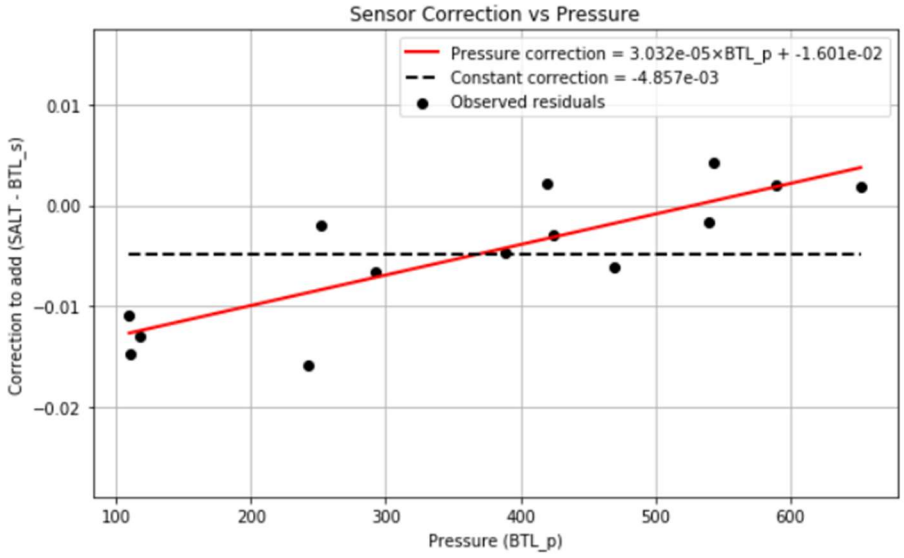

Figure 3: Difference between the salinity measured in the bottles and that observed by the CTD.

```
=== F-test comparing constant vs pressure-dependent correction ===
F-statistic: 28.0418
p-value: 0.0002
✅ Use a pressure-dependent correction.
```

b) Dissolved Oxygen

Oxygen concentrations observed by the CTD and those determined through Winkler titration were converted to [umol/kg] and compared.

No sample was flagged as bad during the analysis. When the difference between doubles (samples taken from the same depth and station, but not necessarily the same Niskin) was lower than 3 umol/kg, the mean value was retained. If the value was higher than 3 umol/kg both samples were removed. No doubles were removed.

We fitted a line to the data using linear regression, and samples with an error larger than 2.5 times the root mean square error were removed. This procedure was repeated until either no more samples were removed, or the root means square error of the remaining samples was smaller than 2 umol/kg.

A total of 19 samples were included in the regression analysis, and 18 samples were included in the final regression (Figure 4).

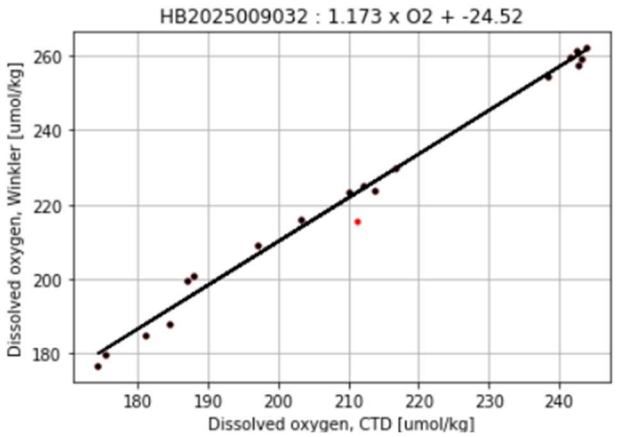

Figure 4: Dissolved oxygen concentration observed with the CTD versus that determined through Winkler titration. The black line shows the regression line used to correct the CTD data, and the black (red) dots are the samples included (not included) in the final regression analysis.

7. Cruise diary (local times)

23 June, 2025

Station O1: Osterfjorden has also been renewed

Station L6: Forgot to take DIC of the surface sample

F_Bjørg: Made two casts

M04, first cast: Bottle 5 didn't close

In general: we had trouble getting the surface sampler to release.

24 June, 2025

HF: The watersampler was not turned on, so we ended up doing four casts. The fjord had been renewed, so the sampling strategy was revised as there was no point sampling for H2S in the basin water.

MF24: Bottle 1 did not close

MF26: The station was added to the program as we had plenty of time.

Still troubles with the surface sampler.

25 June, 2025

We did two MC casts at all stations: at some to secure two good cores, at others to keep one or two to store. Leftover materials were made available for MUJ for artistic purpose.

At each station one core was sampled for aDNA (0-1 cm), another for foraminiferal work (1-10 cm, rose Bengal stained and unstained)..

One extra station was added in Masfjordsen, at a depth close to where the O2 minimum was located from the HF CTD cast (pressure 43). Water was sampled for salinity and trace elements.

26 June, 2025

F_B1 (F_Bjørg): No sediment recovered.

F_B2, added station: CTD, water (salinity and trace elements) at pressure 112, and 2 MC casts. First cast gave two nice cores; second cast came up empty. One MC sampled, one archived. In the one sample, 0-1 cm was split into three (aDNA, stained and unstained foraminifera). The rest of the core was sampled every cm for stained (until 10 cm) and unstained foraminifera (until 22 cm/bottom of core).

F06 – Two good cores at first cast. One sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera, every 1 cm 0-10 cm.

F04 – Two good cores at first cast. Set of towards L04, the F04 MCs fell over, so we had to turn around and go back to F04 for another MC cast. Two good MCs in the second cast. One sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera, every 1 cm 0-10 cm.

L04 – CTD and salinity water sample (pressure 243,5). Two MC casts, four good cores. Two MCs capped and stored, one sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera, every 1 cm 0-10 cm.

L05 – CTD and salinity water sample (pressure 389,5).

L06 – CTD and salinity water sample (pressure 424,5). One MC cast, two full cores. One sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera, every 1 cm 0-10 cm. In the one sampled for stained and unstained foraminifera we found a ca 4*3*2 cm sea urchin between 4 and 8 cm depth, within the softest upper sediment. Coarser/more sturdy sediment from 8-10 cm.

27 June, 2025

St3 – CTD (pressure 197.3) and water for salinity. Not enough for trace elements. One MC cast, 2 good cores. One sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera,

every 1 cm 0-10 cm. No more aDNA gloves available, used gloves for trace elements (this and the following stations).

O01 – One MC cast, 2 good cores. One sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera, every 1 cm 0-10 cm.

St8 - CTD (pressure 590.1) and water for salinity and trace elements. One MC cast, 1 good core in each cast. One sampled for aDNA 0-1 cm, one sampled for stained and unstained foraminifera, every 1 cm 0-10 cm. The rest of the core sampled for aDNA has been archived. If to be sampled further, the upper 0.5 cm will equal 0-1 cm as cored.

St6 - CTD (pressure 111.4) and water for salinity and trace elements. Two MC cast, 2 good cores. MC2-A sampled for aDNA 0-1 cm, MC1-A sampled for stained and unstained foraminifera, every 1 cm 0-10 cm. The rest of the core sampled for aDNA (MC2-A) has been archived. If to be sampled further, the upper 0.5 cm will equal 0-1 cm as cored. Lots of “vegetation” seen at the bottom, likely pushed down along the lined from the top during coring.

	St.name	Date			UTC	Depth	Latitude/ N		Longitude/ E		
CAST		year	mon	day	hh:mm	m	deg	min	deg	min	
0	O1	2025	6	23	8:13	543	60	32.050	5	17.925	
1	L6	2025	6	23	10:04	420	60	41.101	5	10.383	
2	L6	2025	6	23	10:45	420	60	41.101	5	10.383	
3	L4	2025	6	23	11:16	251	60	43.521	5	3.159	Tracemetals
4	F6	2025	6	23	12:52	539	60	49.813	5	3.344	
5	F6	2025	6	23	12:52	539	60	49.813	5	3.344	
6	F_Bjørg	2025	6	23	14:19	83	60	45.602	5	11.787	Two casts, forgot water
7	F_Bjørg	2025	6	23	14:25	83	60	45.602	5	11.787	
8	M4	2025	6	23	14:37	654	60	45.912	5	12.767	Bottle 5 did not close
9	M4	2025	6	23	15:20	654	60	45.912	5	12.767	Only bottle 5
10	M4	2025	6	23	15:30	654	60	45.912	5	12.767	
11	M12	2025	6	23	16:02	121	60	48.327	5	17.731	No bottles
12	M35	2025	6	24	06:21	179	60	52.258	5	33.331	
13	M35	2025	6	24	06:45	179	60	52.258	5	33.331	
14	HF	2025	6	24	07:15	121	60	54.284	5	30.658	Bottles were not reset
15	HF	2025	6	24	07:25	121	60	54.284	5	30.658	
16	HF	2025	6	24	07:54	121	60	54.284	5	30.658	
17	HF	2025	6	24	08:17	121	60	54.284	5	30.658	
18	MF24	2025	6	24	09:14	469	60	52.178	5	22.026	
19	MF24	2025	6	24	09:52	469	60	52.178	5	22.026	
20	MF26	2025	6	24	11:02	479	60	52.355	5	24.957	
21	M16	2025	6	24	11:43	297	60	49.533	5	20.754	
22	M16	2025	6	24	12:12	297	60	49.533	5	20.754	
23	M12	2025	6	24	12:49	107	60	48.302	5	17.694	
24	HF02	2025	6	25	12:32	43	60	54.604	5	31.0882	S N4, Trace N5
25	F_B2	2025	6	26	06:29	112	60	88.460	5	12.9676	S & Trace 1
26	L04	2025	6	26	11:00	244	60	43.511	5	3.1462	
27	L05	2025	6	26	11:53	390	60	42.481	5	6.3501	
28	L06	2025	6	26	12:53	425	60	41.176	5	10.3734	
29	St3	2025	6	27	05:30	197	60	33.252	5	15.018	
30	St8	2025	6	27	06:58	590	60	36.472	5	28.599	Trace elements
31	St6	2025	6	27	07:54	111	60	33.190	5	27,0576	Trace elements

Table 1: Details about CTD-stations occupied during HB2025009032.

Table 2b: Stations with only surface samples. We returned to station M26 later in the day to take a full CTD-station.

Cast	Station	Date & time	Latitude	Longitude	Temp	Samples
32	L03	23/6 11:34	60 43.931 N	5 01.556 E	13C	*Methane, DIC, Nutrients, NH4*
33	F03	23/6 12:25	60 51.184	4 52.511	13C	*Methane, DIC, Nutrients, NH4, Salt*
34	F08	23/6 14:01	60 47.44	5 7.845	13.5C	*Methane, DIC, Nutrients, NH4, Salt*
35	M28	24/6 8:50	60 52.54 N	5 26.33 E	13C	*Methane, DIC, Nutrients, NH4, Salt*
36	M26	24/6 8:59	60 52.37 N	5 24.94 E	13C	*Methane, DIC, Nutrients, NH4, Salt*
37	M22	24/6 11:33	60 51.07 N	5 20.99 E	14C	*Methane, DIC, Nutrients, NH4, Salt*

Table 2: Pressure (dbar) where the Niskin bottles were closed.

Cast	Station Name	Bottle 1	Bottle 2	Bottle 3	Bottle 4	Bottle 5	Bottle 6
0	O1	544	544	544	544	544	544
1	L6	420	420	353	303	253	203
2	L6	101	81	51	21	11	6
3	L4	250	250	250	250	250	250
4	F6	541	541	455	405	253	153
5	F6	101	81	51	21	11	6
6	F_Bjørg						
7	F_Bjørg	71	71	71	71	71	71
8	M4	653	653	507	354	254	203
9	M4	256					
10	M4	103	82	53	22	12	7
11	M12						
12	M35	177	177	152	101	81	61
13	M35	51	41	31	21	11	6
14	HF						
15	HF	66	61	55	51	45	40
16	HF	71	36	31	21	11	6
17	HF	119	102	81	51	46	41
18	MF24	470	470	405	304	253	203
19	MF24	101	81	51	21	11	6
20	MF26	478	478	478	303	303	303
21	M16	293	293	253	202	152	101
22	M16	81	51	21	16	11	6
23	M12						
24	HF2	43					

25	F_B2	112					
26	L04	244					
27	L05	390					
28	L06	425					
29	St3	197					
30	St8	590					
31	St6	111					

Table 4 Sediment core stations

Station ID	Lat N	Long E	Depth (m)	MC cast	Core name	Comments
HB25-265-M16	60°49,5406	5°20,7384E	295,5	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-M16	60°49,5406	5°20,7384E	295,5	1	MC1-B	Corer not closed, not used
HB25-265-M16	60°49,5238	5°20,7754E	295,8	2	MC2-A	aDNA 0-1 cm
HB25-265-M16	60°49,5238	5°20,7754E	295,8	2	MC2-B	Not used, sed given to MUJ
HB25-265-M24	60°52,2041	5°22,0414	458,11	1	MC1-A	ARCHIVED, 58 cm
HB25-265-M24	60°52,2041	5°22,0414	458,11	1	MC1-B	ARCHIVED, 48 cm, aDNA
HB25-265-M24	60°52,2122	5°22,0951	456,15	2	MC2-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-M24	60°52,2122	5°22,0951	456,15	2	MC2-B	aDNA 0-1 cm
HB25-265-HF	60°54,2771	5°30,6401	121,76	1	MC1-A	ARCHIVED, 60 cm
HB25-265-HF	60°54,2771	5°30,6401	121,76	1	MC1-B	Not kept
HB25-265-HF	60°54,2648	5°30,6211	122,23	2	MC2-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-HF	60°54,2648	5°30,6211	122,23	2	MC2-B	aDNA 0-1 cm
HB25-265-HF02	60°54,5956	5°31,0700	46,09	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-HF02	60°54,5956	5°31,0700	46,09	1	MC1-B	Several sea urchins at the top, not kept
HB25-265-HF02	60°54,5859	5°31,0590	44,79	2	MC2-A	Empty
HB25-265-HF02	60°54,5859	5°31,0590	44,79	2	MC2-B	aDNA 0-1 cm
HB25-265-M35	60°52,2669	5°33,3478	179,42	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-M35	60°52,2669	5°33,3478	179,42	1	MC1-B	aDNA 0-1 cm
HB25-265-M35	60°52,2550	5°33,3150	177,57	2	MC2-A	ARCHIVED, 35 cm
HB25-265-M35	60°52,2550	5°33,3150	177,57	2	MC2-B	ARCHIVED, 35 cm, aDNA
HB25-265-F_B	60°45,6368	5°11,8197	69,61	1	MC1-A	Empty, no more tries
HB25-265-F_B	60°45,6368	5°11,8197	70,1	1	MC1-B	Empty, no more tries
HB25-265-F_B2	60°48,8520	5°12,9344	124,68	1	MC1-A	ARCHIVED
HB25-265-F_B2	60°48,8520	5°12,9344	124,68	1	MC1-B	aDNA 1/3, 1/3 sampled for living foraminifera (rose bengal), 1/3 sampled for foraminifera 0-1cm; every cm, 1-10 cm halfsampled for living foraminifera (rose bengal), the other half sampled for foraminifera.
HB25-265-F_B2	60°48,8847	5°12,9378	102,65	2	MC1-A	Empty
HB25-265-F_B2	60°48,8847	5°12,9378	102,65	2	MC1-B	Empty

HB25-265-F06	60°49,8261	5°03,3542	540,28	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-F06	60°49,8261	5°03,3542	540,28		MC1-B	aDNA 0-1 cm
HB25-265-F04	60°51,0400	4°57,5252	428,19	1	MC1-A	Good recovery, fell over, thrown away
HB25-265-F04	60°51,0400	4°57,5252	428,19	1	MC1-B	Good recovery, fell over, thrown away
HB25-265-F04	60°51,0311	4°57,5461	428,33	2	MC2-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-F04	60°51,0311	4°57,5461	428,33	2	MC2-B	aDNA 0-1 cm
HB25-265-L04	60°43,5004	5°03,1373	249,7	1	MC1-A	ARCHIVED
HB25-265-L04	60°43,5004	5°03,1373	249,7	1	MC1-B	ARCHIVED
HB25-265-L04	60°43,4973	5°03,1072	248,41	2	MC2-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-L04	60°43,4973	5°03,1072	248,41	2	MC2-B	aDNA 0-1 cm
HB25-265-L06	60°41,1032	5°10,3670	143,6	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-L06	60°41,1032	5°10,3670	143,6	1	MC1-B	aDNA 0-1 cm
HB25-265-St3	60°33,2574	5°14,9984E	198,07	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-St3	60°33,2574	5°14,9984E	198,07	1	MC1-B	aDNA 0-1 cm
HB25-265-O01	60°32,0628N	5°17,1917E	541,76	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-O01	60°32,0628N	5°17,1917E	541,76	1	MC1-B	aDNA 0-1 cm
HB25-265-St8	60°36,4542N	5°28,5617E	591	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-St8	60°36,4542N	5°28,5617E	591	1	MC1-B	aDNA 0-1 cm, the rest archived (upper 0.5 cm equals left. Over 0.-1cm)
HB25-265-St6	60°33.1875N	5°27.0685E	113,65	1	MC1-A	Half sampled for living foraminifera (rose bengal), the other half sampled for foraminifera, every cm, 0-10 cm.
HB25-265-St6	60°33.1875N	5°27.0685E	113,65	1	MC1-B	Overpenetrated, not kept
HB25-265-St6	60°33.2402N	5°26.9940E	114,99	2	MC2-A	aDNA 0-1 cm, the rest archived (upper 0.5 cm equals left. Over 0.-1cm)
HB25-265-St6	60°33.2402N	5°26.9940E	114,99	2	MC2-B	Overpenetrated, not kept